Comparative Study on The Phytochemical, Antistaphylococcal and Antioxidant Properties of the Stem Bark of Jatrophacurcas L and Jatrophagossypifolia L

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ABSTRACT: This study was conducted to assess the phytochemical, antistaphylococcal and antioxidant properties of the leaf and stem bark of *Jatrophacurcas* and *Jatrophagossypifolia*. Ethanolic extracts of the leaf and stem bark of *J. curcas* and *J. gossypifolia* were obtained using standard methods. Qualitative and quantitative phytochemical properties of the Jatropha plant was assessed using standard methods. Antistaphylococcal and minimum inhibitory concentrations of the extracts were assessed against *Staphylococcal aureus* obtained from different sources using agar diffusion method. The ferrous ion and hydroxyl radical scavenging activity of the extracts were also determined using standard methods. Quantitative phytochemicals in the two *Jatropha* species do not follow a regular trend. However, saponin was highest in *J. curcas* stem bark (35.64 mg/g) while *J. gossypifolias*tem bark and leaf have the highest flavonoid (31.35 mg/g) and alkaloids (23.20 mg/g) respectively. The antistaphylococcal effect of the combined extract was higher and significantly different (P≤0.05) than when used singly. The highest antistaphylococcal effect (19.83mm) was recorded for the combination of *J. curcas* and *J gossypifolia* leaf (JCL and JGL) against *Staphylococcus aureus* obtained from blood. Antioxidant assay of extracts revealed a concentration-dependent effect. The antioxidant activities of the extracts vary from one extract to the other. The results obtained from this study indicates that bioactive compounds present in *J. curcas* and *J gossypifolia* can be exploited as source of effective antistaphylococcal and antioxidant compounds.

Keywords:Comparative; Antioxidant; Antistaphylococcal; Leaf; Stem bark; Extract; Jatrophacurcas; Jatrophagossypifolia.

دراسة مقارنة حول الخصائص الكيميائية النباتية والمضادة للمكورات العنقودية ومضادات الأكسدة للحاء الجزعي لنباتي من صنف الجاتروفاكوركاس وجاتروفا جوسييفوليا

ف و أويتيو و ز ي تيمانيو

الملخص: أجريت هذه الدراسة لتقييم الخصائص الكيميائية النباتية والمضادة المكورات العنقودية والمضادة للأكسدة من أوراق وسيقان الجاتروفا كوركاس و جاتروفا جوسيبيفوليا. تم الحصول على المستخلص الكحولي من أوراق وسيقان الجاتروفا كوركاس و جاتروفا جوسيبيفوليا باستخدام الطرق القياسية. تم تقييم الخصائص النوعية والكمية الكيميائية النباتية لنبات الجاتروفا باستخدام الطرق القياسية. تم تقييم النشاط المضاد الميكروبي للمكورات العنقودية الذهبية التي تم الحصول عليها من مصادر مختلفة باستخدام طريقة انتشار الأجار. تم تحديد الأنزيمات المضادة للأكسدةو الشوارد الحرة للمستخلصات أيضًا باستخدام الطرق القياسية. كشف الفحص الكيميائي النباتي الكمي أن توزيع المواد الكيميائية النباتية في نوعي جاتروفا تختلف من نوع لأخر. ومع ذلك ، كان السابونين أعلى في لحاء جذع جاتروفا كركاس (35.64 مجم/جم) ، بينما كان لحاء ساق الجاتروفا جوسيفوليا وأوراقه أعلى نسبة من الفلافونويد معنويًا (20.05) على في لحاء جذع جاتروفا كركاس (23.64 مجم/جم) ، بينما كان لحاء ساق الجاتروفا تعقوليا وأوراقه أعلى نسبة من الفلافونويد معنويًا (20.05) عند استخدامه منفردًا. تم تسجيل أعلى تأثير مضاد للمكورات العنقودية المصادل المستخلصين معا أعلى ومختلفًا اختلافًا معنويًا (20.05) عند استخدامه منفردًا. تم تسجيل أعلى تأثير مضاد للمكورات العنقودية (30.05) مع أوراق الجاتروفا جوسيفولياضد المكورات العنقودية الذهبية المأحوذة من الدم. وتبين من نتائج مضادات الأكسدة للمستخلصات انها تتناسب طرديا مع التركونا. تعنوي إرادة المصرول على المستخلصات من مستخلص إلى تأثير مضاد للمكورات العنقودية لكلا المستخلصين معا أعلى ومختلفًا معنويًا (20.05) عند المكورات العنقودية الذهبية المأحوذة من الدم. وتبين من نتائج مضادات الأكسدة للمستخلصات انها تتناسب طرديا مع التركيزات. تختلف جوسيفولياضد المكورات العنقودية الذهبية المأحوذة من الدم. وتبين من نتائج مضادات الأكسدة للمستخلصات انها تتناسب طرديا مع التركيزات. تختلف الأنشطة المضادة للأكسدة في المستخلصات من مستخلص إلى آخر. تشير النتائج التي تم الحصول عليها من هذه الدراسة إلى أن المركبات النشطة بيولوجبًا الأنشطة المضادة للأكسدة في المستخلصات من مستخلص إلى آخر. تشير النتائج التي تم الحصول عليها من هذه الدراسة إلى ألمركبات المكورات المركبات النشطة بيولور. المستخلصة من أوراق وسيقان ا

ا**لكلمات المفتاحية:** مقارنة؛ مضادات الأكسدة، مضادات المكورات العنقودية ؛ اللحاء الجذعي ؛ جاتروفا كركاس جاتروفا جوسيبيفوليا.



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1. Introduction

Diseases of man caused by microorganisms have become a major threat to human health and existence. *Staphylococcus* species are very important pathogen responsible for bacterial infections in hospitals and communities worldwide. They are very diverse and are implicated in various infection processes especially in immunecompromised individuals and those with implant devices such as shunts and catheters [1]. *Staphylococcus aureus* has been recognized as a versatile microorganism worldwide [2]. It is a human pathogen and a part of the normal flora of human skin [3]. It can colonize and infect both patients and healthy people with life threatening effects [4]. *Staphylococcus*species generally exhibit multiple antibiotic resistances [5, 6].

Resistance of microorganisms to commonly used antimicrobial agents is a major challenge in the treatment of diseases of microbial origin. Hence, the search for antimicrobial agents that are safe and most importantly effective against diseases of microbial origins has doubled/tripled in the last three decades. This is as a result of resistance developed by microorganisms against commonly used antimicrobial agents, the safety of these antimicrobial agents and the side effects associated with the use of antibiotics in the treatment of gastroenteritis and other infections [7].

Plant-based bioactive compounds have recently become of great interest in the search for suitable, safe and friendly alternatives to those existing antimicrobials which are becoming less effective. Researchers have made tremendous efforts to discover new antimicrobial compounds from natural products especially of plant origin. Ncube *et al.* [8] submitted that medicinal plants are the richest bio-resource of drugs of the traditional system of medicines, pharmaceutical intermediates and chemical entities for synthetic drugs. Moreover, many people all over the world have resorted to the use of herbal medicine for the treatment of various health challenges [9].

Jatropha species are among these medicinal plants that have attracted interest as potential sources of antimicrobial agent. They belong to the Family Euphorbiaceae. They contain secondary metabolites such as alkaloids, tannins, flavonoids, steroid, saponins and phenolics which are known to possess medicinal properties [10, 11]. The stem bark, root, leaf, sap, seed and the oil from the seed of *Jatropha* species had been used for treating various ailments ranging from skin diseases, parasitic diseases, urinary tract infections, bleeding of gum and toothaches, treatment of wound and sores, fever and many others [12-18].

Jatropha curcas and *Jatropha gossypifolia* are commonly found in Nigeria. These two plants had found wide use in ethno-medicine especially in rural parts of Nigeria. The present study is therefore aimed at comparative study of the phytochemical properties, antioxidant and antistaphyloccal activities of leaf and stem bark extracts of *J. curcas* and *J. gossypifolia*singly and in combination.

2. Materials and methods

2.1 Collection and preparation of Jatropha species extracts

Fresh leaves and stem bark of *Jatropha curcas* L and *Jatropha gossypifolia* L were collected from a local farm at Oke-Aro in Akure, Ondo State, Nigeria. The plants were identified and authenticated by a plant scientist in the Department of Crop Science and Pest Management, Federal University of Technology, Akure. The stem bark and leaf were rinsed with clean water and sun-dried for three weeks under shade and then pulverized using a mechanical grinder. The pulverized plant material was kept in an air-tight cellophane bag until required. Powdered *Jatropha* species (200 g) of the leaf and stem bark were each placed in 1000 ml of 99.7% ethanol (analytical grade) and kept in conical flasks, each was shaken in a rotary shaker at 121rpm for 24 hrs. After 24 hours, the suspension was filtered with a double-layer muslin cloth and Whatman No. 1 filter paper. The resulting filtrates were concentrated under reduced pressure in a rotary evaporator (RE - 52A; Union Laboratory, England) at 40 °C.

2.2 Test organism, Staphylococcus species

All *Staphylococcus* species were grown on Mannitol salt agar plates at 37 °C for 24 hours. The isolates were maintained on agar slant and stored in the refrigerator at 5 °C until used.

2.3 Quantitative phytochemical analysis of Jatropha species

Quantitative phytochemical screening of the crude extracts of the stem bark and leaf of *Jatropha* species was performed using standard procedures as described by Harborne [19]; Trease and Evans [20]. The quantity of the following phytochemicals viz; saponin, tannin, flavonoid, cardiac glycosides and alkaloids were assessed in the extracts obtained from the two *Jatropha* species.

2.4 Determination of total phenolic content

The total phenol content of leaf and stem bark was determined (Gallic acid equivalent) as described by Singleton *et al.* [21] with slight modifications. Briefly, 200 μ L of the extract dissolved in 10% DMSO (240 μ gmL⁻¹) was incubated with 1.0 ml of Folin-Ciocalteu reagent (diluted 10 times) and 800 μ L of 0.7 mol L⁻¹ Na₂CO₃ for 30 minutes at room temperature. Then, the absorbance was measured at 765 nm on a Shimadzu UV mini 1240 spectrophotometer (Shimadzu, Japan). All measurements were done in triplicates. Results are expressed as mg GAE / 100 g dry ethanol extracts.

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2.5 Determination of antistaphylococcalproperties of Jatropha species

Antistaphylococcal activity of stem bark and leafextracts of *Jatropha* species was determined by agar well diffusion method as described by Abubakar*et al.* [22]. *Staphylococcus* species obtained from different sources were cultivated on nutrient broth at 32 °C for 18 hours. The inoculum size was adjusted by serial dilution to obtain 0.5 McFarland turbidity standards. The extract was reconstituted in 20% v/v of Dimethyl sulfoxide (DMSO). An aliquot of 0.1 mL containing organism was aseptically transferred and evenly spread onto the dried surface of the sterile Mueller Hinton agar plate. A well of 8 mm was bored in the agar plate with a sterile cork borer. Each extract was sterilized through a membrane filter (0.22 μ m) and 0.1 mL was aseptically introduced into the well in the Petri dishes already inoculated with *Staphylococcus* species with the aid of a micropipette. A volume of 0.1 mL of Ciprofloxacin was used as positive control while 20% of DMSO served as a negative control. The plates were incubated at 37 °C for 24 hours. The diameter of the inhibition zones was measured in millimeters.

2.6 Antioxidant assay

The following antioxidant assays were performed on the stem bark and leaf extracts obtained from *Jatropha* species.

2.7 Hydroxyl radical scavenging ability of Stem Bark and Leaf extracts of Jatropha species

The determination of the scavenging effect on hydroxyl radicals was carried out as described by Oyetayo *et al.* [23]. The reaction mixtures in a final volume of 1.0 ml, containing 0.4 ml of 20 mmol/ml sodium phosphate buffer (pH 7.4), 0.1 ml of 0.125-2 mg/ml extracts, 0.1 ml of 60 nmol/L deoxyribose, 0.1 ml of 10 mmol/L hydrogen peroxide, 0.1 ml of 1 mmol/L ferric chloride, 0.1 ml of 1.04 mmol/L EDTA and 0.1 ml of 2 mmol/L ascorbic acid was incubated at 37 °C for 1 hour. Solutions of FeCl₂ and ascorbic acid were made up immediately before use in de-ionized water. The reaction was stopped by adding 1 ml of 17 mmol/L thiobarbituricacid (TBA) and 1 ml of 17 mmol/L trichloroacetic acid (TCA). The mixture was boiled for 15 min, cooled in ice and then the absorbance was measured at 532 nm using a UNICO 2100 spectrophotometer (As).

2.8 Ferrous ion chelating ability assay

The Fe²⁺chelating ability of leaf and stem bark ethanol extracts was determined by employing a modified method of Puntel *et al.* [24]. Freshly prepared 500 μ mol L⁻¹ FeSO₄ was added to a solution containing 168 μ L of 0.1 molL⁻¹Tris-HCl (pH 7.4), together with 218 μ L of saline and an ethanol extract (1-5 mg/ml). The solution was incubated for 5 minutes, followed by the addition of 13 μ L of 0.25%, 1,10 phenantroline (w/v). Absorbance was read at 510 nm. Fe²⁺chelating ability was expressed as percentage inhibition.

2.9 Statistical analysis

Experiments were carried out in replicates and data obtained were analyzed by one way analysis of variance (ANOVA) and means were separated by Duncan multiple range test (SPSS 17.0 version). Differences were considered significant at $P \le 0.05$.

3. Results

The distribution of the phytochemicals in the two *Jatropha* species does not follow a regular trend as revealed by quantitative phytochemical screening (Table 1). Saponin was highest and significantly different ($P \le 0.05$) in *J. curcas* stem bark (35.64 mg/100 g) when compared to other extracts, while *J. gossypifolia* stem bark and leaf has the highest flavonoid (31.35 mg/100 g) and alkaloid (23.20 mg/100g) respectively. The phenolic contents of the different parts of the *Jatropha* species vary and are significantly different ($P \le 0.05$). The leaf extract of *J. curcas* has the highest total phenol (2.2 mgGAE/g) while the least was recorded in the leaf extract of *J. gossypifolia* (0.8 mgGAE/g) (Figure 1).

Table 1. Quantitative Phytochemical Contents of Leaf and Stem Bark Extracts of Two Jatropha Species.

Phytochemicals	JCL	JCS	JGL	JGS
Saponin	4.30 ± 0.29^{a}	$35.64 \pm 1.56^{\circ}$	14.03 ± 1.21^{b}	13.67±0.41 ^b
Tannin	0.93 ± 0.03^{a}	$0.98{\pm}0.12^{a}$	$7.20{\pm}0.93^{b}$	$9.02 \pm 0.23^{\circ}$
Flavonoid	1.11 ± 0.05^{a}	10.15 ± 0.28^{b}	22.81±0.37 ^c	31.35 ± 2.38^{d}
Cardiac glycosides	7.73 ± 0.32^{d}	1.95 ± 0.06^{a}	5.15 ± 0.14^{b}	6.85±0.23 ^c
Alkaloid	3.87 ± 0.11^{a}	5.05 ± 0.06^{a}	23.20 ± 2.42^{b}	2.90±0.11 ^a

Key

JCL: J. curcas leaf extract, JCS: J. curcas stem bark extract.

JGL: J.gossypifolia leaf extract, JGS: J. gossypifolia stem bark extract.

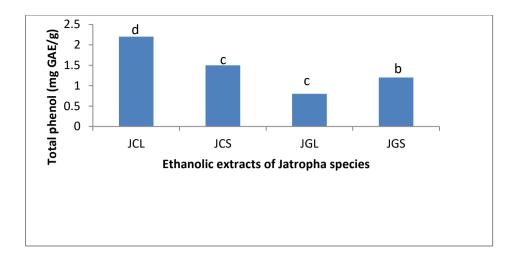


Figure1. Total Phenolics of extracts obtained from Jatropha species.

Bar with different superscript are significantly different (P≤0.05) **Key** JCL: *J. curcas* leaf extract JCS: *J. curcas* stem bark extract JGL: *J.gossypifolia* leaf extract JGS: *J. gossypifolia* stem bark extract

There were significant differences between and within groups in the antistaphylococcal activity of the extracts against Staphylococci used in this study except for *J. curcas* leaf extract (JCL) and *J. gossypifolia* stem bark extract (JGS) where there were no significant differences ($P \ge 0.05$) within the two groups (Table 2). The antistaphylococcal activity of the combined extracts was higher and significantly different ($P \le 0.05$) than when used singly. The highest antistaphylococcal effect (19.83 mm) was obtained for the combination of the leaf extracts of the two *Jatropha* species (JCL and JGL) against *S. aureus* isolated from blood. *J.curcas* stem bark extract (JCS) displayed the least antistaphylococcal effect of 7.00mm against *S.aureus* isolated from urine.

Table 2. Antistaphylococcal Property of the Stem Bark and Leaf extract of Jatropha species.

Source of Staph.	JGL	JCL	JGS	JCS	JCL&JGL	JCS&JGS	СРХ
POW	10.50±1.73	15.25±0.96	16.25±2.22	9.50±4.04	15.75±2.63	18.25±0.50	21.25±2.22
Skin Swab	11.00 ± 1.93	13.13±3.36	14.00 ± 3.42	11.50 ± 4.31	17.75 ± 4.03	14.00 ± 1.07	24.25±1.67
Urine	12.00 ± 1.43	11.80 ± 4.66	11.60 ± 4.72	7.00 ± 1.00	15.00 ± 1.22	14.20 ± 1.10	32.60±1.14
Blood	12.50 ± 0.55	13.67±1.03	15.00 ± 2.19	12.17 ± 2.64	19.83 ± 2.99	16.33±1.37	22.83±1.17
Nose	10.93 ± 1.81	14.62 ± 2.45	14.65 ± 3.16	12.46±3.16	$15.04{\pm}1.93$	14.92 ± 2.17	24.23±1.75
ATCC	14.33 ± 0.58	12.67±0.58	12.33±1.53	7.33±1.53	17.00 ± 1.00	13.83±0.76	27.33±1.16
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Key

POW: Post Operative Wound JCL: J. curcas leaf extract JCS: J. curcas stem bark extract JGL: J.gossypifolia leaf extract JGS: J. gossypifolia stem bark extract CPX: Ciprofloxacin

The antioxidant capacity of the extracts is revealed in Figures 2 and 3. Ferric ion chelating activity of the two *Jatropha* species was observed to be concentration dependent. The higher ferric ion chelating effect (72%) was displayed by extract obtained from the stem bark of *J. curcas* (JCS) while the least (56%) was recorded for extract obtained from *J. gossypifolia* stem bark extract (JGS) at a concentration of 5 mg/mL. Hydroxyl ion scavenging activities of the *Jatropha* species were also concentration dependent. The highest hydroxyl ion scavenging ability (88%) was obtained for *J. curcas* lea extract (JCL) while the least (55%) was obtained for *J. gossypifolia* leaf extract (JGL).

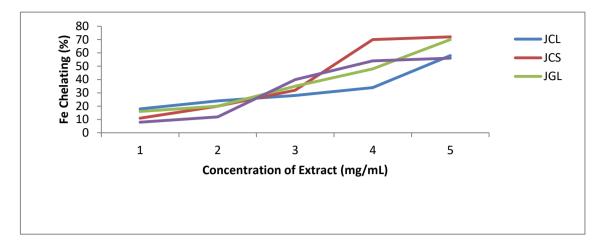


Figure 2. Ferric ion Chelating Ability of of the Stem Bark and Leaf Extracts of Two Jatropha species.

Key

JCL: *J. curcas* leaf extract JCS: *J. curcas* stem bark extract

JGL: J.gossypifolia leaf extract

JGS: J. gossypifolia stem bark extract

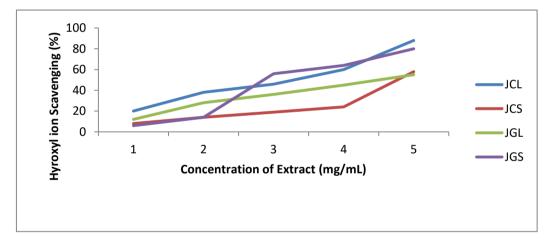


Figure 3. Hydroxyl ion Scavenging of Leaf and Stem Bark Extracts of Two Jatropha Species.

Key JCL: J.curcas leaf extract JCS: J.curcas stem bark extract JGL: J.gossypifolia leaf extract JGS:J.gossypifoliastem bark extract

4. Discussion

The medicinal properties of plants have been identified since time immemorial and they are now recognized as a source of bioactive compounds which can be exploited for the development of novel biopharmaceutical agents. Plants synthesize secondary metabolites, many of which are important in promoting good health in animals and humans [25]. These secondary metabolites such as alkaloids, tannins, anthraquinones, saponin, glycosides and so on are important for the survival of plants in their ecosystem. *Jatropha* species are important medicinal plants that have folkloric applications.

In the present report, quantitative phytochemical screening of ethanolic extracts obtained from the leaf and stem bark of two *Jatropha* species, *J. curcas* and *J. gossypifolia*, was not similar to the report of Atamgba *et al.* [26] which revealed that the phytochemicals are more present in leaf than in the stem bark. However, in the present report, saponin and flavonoid were higher in *J. curcas* stem bark (35.64 mg/100 g) and *J. gossypifolia* stem bark extract (31.35

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mg/100g) while cardiac glycosides and alkaloids were highest in the leaves of *J. curcas* (7.73 mg/100 g) and *J. gossypifolia* (23.20 mg/100 g) respectively. There was no relative increase in the phytochemical content as one moved from the stem bark to the leaf. Secondary metabolites such as alkaloids, tannins, anthraquinones, saponin, and glycosides produced by plants have potent antimicrobial effects [27]. These secondary metabolites are known to exert considerable antimicrobial activity through different mechanisms [28]. Specifically, saponins are stored in plant cells as inactive precursors but are readily converted into biologically active antibiotics by enzymes in response to pathogen attacks [29, 30]

The total phenol content obtained in the leaf and stem bark of the two *Jatropha* species varies from 0.8 mgGAE/g to 2.2 mgGAE/g. Othman *et al.* [31] had earlier reported the total phenol content in leaves and stem of *J. curcas* plant collected from Malaysia as 1.33 mg GAE/g and 0.11 mg GAE/g respectively. Akhtar *et al.* [32] also reported higher level of total phenol in the leaf of *J. curcas* than the stem bark as observed in this study. The total phenol content in the leaf (2.2 mgGAE/g) and stem bark (1.5 mgGAE/g) of *J. curcas* were however higher and significantly different from what was obtained in *J. gossypifolia* leaf and stem bark. *J. curcas* had been reported to have high content of phenolic compounds [33, 34]. Recently, Vega-Ruizet al. [35] also reported that *J. cinerea* and *J. cordata* two species of *Jatropha* collected in Mexico are important source of phenolic acids and flavonoids.

The ethanolic extracts of the two *Jatropha* species show good antistaphylococcal effects. There was no significant difference in the antistaphylococcal activities exhibited by the leaf and stem extracts of the plant. However, combination of the leaf extracts and stem bark extracts of the two *Jatropha* species exhibited higher and significantly higher antistaphylococcal effect. Rampadarath *et al.* [36] reported inhibition of the growth of *S. aureus, S. epidermidis, Pseudomonas aeruginosa, Escherichia coli, Bacillus cereus, B. subtilis* and *Proteus vulgaris* by methanolic extract of the leaves of *J. curcas*. Moreover, Igbinosa [37] had earlier observed the antibacterial effect of the crude root extracts of *J. carcus*. In another report, the essential oil obtained from J. gossypifolia was found to exhibit strong antibacterial activity against *Escherichia coli, Enterococcus faecium*, and *S. aureus* [38]. The antibacterial effect observed in this study may lay credence to the ethnomedicinal uses of *Jatropha* species in the treatment of wounds and sore. Felger and Moser in 1973 [35] reported the use of *J. cinerea* roots by the Seri ethnic group in the state of Sonora (Mexico) to cure dysentery and the sap to treat mouth ulcers. Another species of *Jatropha, J. cordata* root is also used by ethnic groups in the state of Sonora, Mexico to combat toothache and the stem and leaves are used to cure gum disease [39].

Extracts obtained from the leaf and stem bark of *J. curcas* and *J. gossypifolia* exhibited concentration-dependent antioxidant activity. Sunday *et al.* [40] had earlier observed that extracts of *J.curcas*(Leaves and Stem bark extract) and *J. gossypifolia* (Leaves and Stem bark extract) exhibited effective antiradicals' potencies against the different oxidants, indicating they are good electron donors. In another recent report, extracts of leaves and stems from *J. cinerea* and *J. cordata* collected in Mexico were found to possess good antioxidant activity [35]. The antioxidant activities of plants had been attributed to the total phenol and other antioxidant compounds such as volatile oils, amino acids, vitamins and others [41]. The total phenol obtained in this study was high enough to elicit appreciable antioxidant activity. It has been reported that a significant correlation between antioxidant activity and total phenolic contents is prove that phenolic compounds is a major contributor to the antioxidant activity of different parts of *Jatrophas* pecies [32].

5. Conclusion

The results obtained from this study conclusively indicate that extracts obtained from *J. curcas* and *J. gossypifolia* possess effective antistaphylococcal and antioxidant properties. There was no much difference in the antistaphylococcal and antioxidant properties of ethanolic extracts obtained from the two *Jatropha* species; however, the combination of the extracts produced the better antistaphylococcal effect. The bioactive compounds present in the two *Jatropha* species may therefore be exploited as a source of effective antistaphylococcal and antioxidant compounds.

Conflict of interest

The authors declare no conflict of interest.

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References

1. Akinkunmi, E.O. and Lamikanran, A. Species Distribution and Antibiotic resistance in Coagulase–negative Staphylococci colonizing the Gastrointestinal Tract of Children in Ile-Ife, Nigeria. *Tropical Journal of Pharmaceutical Research* 2010, **9**(1), 35-43.

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- 2. Diekema, D.J., Pfaller, M.A., Schmitz, F.J, Smayevsky, J., Bell, J., Jones, R.N., Beach, M. Sentry Participant Group.Survey of infections due to *Staphylococcus* species. Frequencyof occurrence and antimicrobial Susceptibility of isolates collected in the United States, Canada, Latin America, Europe and the Western Pacific Region for the Sentry.
- 3. Oyetayo, V.O. and Akingbesote, E.T. Assessment of the antistaphylococcal properties and bioactive compounds of raw and fermented *Trametes polyzona* (Pers.) Justo extracts. Microb. Biosyst. 2022, **7**, 1-7.
- 4. Daka, D, Solomon, G. and Dawit, Y. Antibiotic-resistance *Staphylococcus* aureus isolated fromcow's milk in the Hawassa area, South Ethiopia. *Ann. Clin. Microbiol. Antimicrob* 2012, 11(26-31). doi:10.1186/1476-0711-11-26.
- Jhora, S.T. and Paul, S. Urinary Tract Infections Caused by *Staphylococcus saprophyticus* and their antimicrobial sensitivity pattern in Young Adult Women. *Bangladesh Journal of Medical Microbiology* 2011, 5(1), 21-5. doi.org/10.3329/bjmm.v5i1.15817.
- 6. Julie, A.C. and Trevor, P.A. Case report of a methicillin- and multiresistant, mecA positive, *Staphylococcus saprophyticus* and retrospective review of 2011-2012 isolates. NZ J Med. Lab Sci. 2013, **67**, 104-8.
- Oyetayo, V.O. and Ogundare, A.O. Antifungal Property of Selected Nigerian Medicinal Plants. *In: Antifungal Metabolites from Plants* (Eds. M. Razzaghi-Abyaneh and M. Rai). Springer-Verlag Berlin Heidelberg 2013, Chapter 3. pp 59-77. DOI: 10.1007/978-3-642-38076-1-3.
- Ncube, N.S., Afolayan, A.J. and Okoh, A.T. Assessment techniques of antimicrobial properties of natural compounds of plant origin. Current methods and future trends. *African Journal of Biotechnology* 2008, 7(12),1797-1806.
- 9. World Health Organisation (WHO) Guidelines on Safety Monitoring of Herbal Medicines in Pharmacovigilance Systems. Geneva, Switzerland; World Health Organisation 2004.
- 10. Nwokocha, A., Blessing I.O., Agbagwa and Okoli B.E. Comparative phytochemical screening of *Jatropha L.* species in the Niger Delta. *Research Journal of Phytochemistry* 2011, **5**, 104-114.
- 11. Gupta, D.D., Haque, M.D., Islam, N.M., Rahman, S., Hassan, A.K.M.M. and Shibib, B.A. Alkaloid and steroid from the stem bark of *Jatrophacurcas* (Euphorbiaceae). *Journal of Pharmaceutical Science* 2011, **10**(1), 9-11.
- 12. Oliver-Beaver, B. Medicinal plants in tropical West Africa. Cambridge University Press, London 1986.
- 13. Rajore, S. and Batra, A. *Jatropha curcas*: A plant of immense potential value. *Journal of Ecology, Taxonomy and Botany* 2003, **27**, 36-41.
- 14. Thomas, R., Sah, N.K. and Sharma, P.B. Therapeutic biology of *Jatropha curcas*: a mini review. *Current Pharmaceutical Biotechnology*. 2008, **9**, 315-324.
- 15. Jongschaapet, R.E., Corre, W.J., Bindraban P.S and Bradenburg, W.A.Claims and facts on *Jatrophacurcas* L. *Plant Research International*. 2007, **7**, 20-28.
- 16. Okolie, R.I., Aigbe, O., Ohafu-Obode, J.O. and Mensah, J.K. Medicinal herbs used for managing some common ailment among Esan people Edo state, Nigeria. *Pakistan Journal of Nutrition* 2007, **6**(5), 470-490.
- 17. Nayak, B.S. and. Patel, K.N. Pharmacognosis studies of *Jatrophacurcas*leaves. *International Journal of Pharmtech Research* 2010, **2**(1), 140-143.
- 18. Agbogidi, O.M and Ekeke E.A. *Jatropha curcas: Linn* an important but neglected plant species in Nigeria. *Journal of Biological and Chemical Research* 2011, **28**(1), 52-62.
- 19. Harbone, J.B. Method of extraction and isolation in phytochemical techniques. 3rd ed. Chapman and Hall: London 1998.
- Trease G.E., Evans M.C. Pharmacognosy, fourteenth ed. Elsevier, New Delhi, India. http://dl.konkur.in/post/Book/MedicalScience/Trease-and-Evans-Pharmacognosy 2005, 16th-Edition-%5 Bkonkur.in %5D.pdf
- Singleton, V., Orthofer, R. and Lamuela-Raventos, R. Analysis of total phenols and other oxidation substrates and antioxidants by means of Folin-Ciocalteu reagent. In L. Packer (Ed.). Oxidants and Antioxidants, part A, Methods in Enzymology (Vol. 299). NewYork: Academic Press 1999, pp. 152-178.
- 22. Abubakar, Z, Ogidi C.O, Oyetayo V.O. Assessment of Antistaphylococcal Activity of Ethanolic Extract of *Lenzites quercina* on Clinical *Staphylococcus* species. Clinical Phytoscience 2016, **2**, 8.
- 23. Oyetayo, V.O. Free radical scavenging and antimicrobial properties of extracts of wild mushrooms. *Brazilian Journal of Microbiology* 2009, **40**, 380-386.
- 24. Puntel, R.L., Nogueira, C.W and, Rocha JBT. Krebs cycle intermediates modulate thiobarbituric acid reactive species (TBARS) production in rat brain in vitro. Neurochemical Research 2005, **30**(2), 225-235. http://dx.doi.org/101007/s11064-004-2445-7.
- Oyetayo, V.O. Antimicrobials from Wild edible plants of Nigeria. In "*NaturalAntimicrobials in Food quality and Food Safety*" (Eds Mahendra Rai and Michael L.Chikindas), CAB International, UK 2011, Chapter 17, pp 261-276. ISBN: 9781845937690.
- 26. Atamgba, A.A., Margaret, A.A., Kayode, D., Amonor, J.W.The biomedical significance of the phytochemical, proximate and mineral compositions of the leaf, stem bark and root of *Jatrophacurcas*. *Asian Pacific Journal of Tropical Biomedicine* 2015, **5**(8), 650-657.

- Oyetayo, V.O. Antimicrobials from Wild edible plants of Nigeria. In *Natural Antimicrobials in Food quality and Food Safety* (Eds Mahendra Rai and Michael L. Chikindas), CAB International, UK 2010, Chapter 17, pp 261-276. ISBN: 9781845937690.
- Oyi, A.R., Onaolapo, J.A., Haruna, A.K. and Morah, C.O. Antimicrobial screening and stability studies of crude extract of *Jatropha curcas* Linn latex (Euphorbiaceae). Nigerian. *Journal of Pharmaceutical Science* 2007, 6, 14-20.
- 29. Cowan. M.M. Plant products as antimicrobial agents. ClinMicrobiol Rev 1999, 12(4), 564-582.
- 30. Arif, T., Bhosalea, J.D., Kumara, N., Mandala, T.K., Bendreb, R.S., Lavekara, G.S. and Dabura, R. Natural products-antifungal agents derived from plants. J Asian Nat Prod Res 2009, **11**(7), 621-638.
- Othman, A.R., Abdullah, N., Ahmad, S., Ismail, I.S. and Zakaria, M.P. Elucidation of *in-vitro* anti-inflammatory bioactive compounds isolated from *Jatropha curcas* L. plant root. BMC Complement. Altern. Med. 2015, 15, 1-10.
- Akhtar, P., Yaakob, Z., Ahmed, Y., Shahinuzzaman, M., Mohammad, M.K. and Hyder, Z.Total Phenolic Contents and Free Radical Scavenging Activity of Different Parts of *Jatropha* species. *Asian Journal of Chemistry* 2018, 30(2), 365-370.
- Rebecca, R., Samuel, D.D., Bello, Y.M., Simeon, O.K. Qualitative Phytochemistry and Antibacterial Resistance Pattern of Leaves and Stem Bark Extracts of *Jatropha curcas*. *American Journal of Microbiology Research*. 2016, 4, 143-146.
- 34. Oyama, M.O., Malachi, O.I. and Oladejo, A.A. Phytochemical Screening and Antimicrobial Activity of Leaf Extract of *Jatropha curcas*. *Journal of Advances in Medical Pharmaceutical Sciences*. 2016, **8**, 1-6.
- 35. Vega-Ruiz, Y.C., Hayano-Kanashiro, C.H., Gámez-Meza, N. and Medina-Juárez, L.A. Determination of Chemical Constituents and Antioxidant Activities of Leaves and Stems from *Jatropha cinerea* (Ortega) Müll. Arg and *Jatropha cordata* (Ortega) Müll. Arg. Plants 2021, **10**, 212. https://doi.org/10.3390/plants10020212.
- Rampadarath, S., Puchooa, D. and Ranghoo- Sanmukhiya, M. Antimicrobial, Phytochemicaland Insecticidal Properties of *Jatropha* species and Wild *Ricinus communis* L. Found in Mauritius. *International Journal of Pharmacognosy and Phytochemical Research* 2014, 6(4), 831-840.
- 37. Igbinosa, O. Antimicrobial activity and phytochemical screening of stem bark extractsfrom *Jatrophacurcas* Linn. *African Journal of Pharmacy and Pharmacology*, 2009, **3**, 58-62.
- Okoh, S.O., Iweriebor, B.C., Okoh, O.O, Nwodo, U.U., and Okoh, A.I. Antibacterial and Antioxidant Properties of the Leaves and Stem Essential Oils of *Jatropha gossypifolia* L. BioMed Research International 2016, Article ID 9392716, 9 pages. http://dx.doi.org/10.1155/2016/9392716.
- 39. Johnson, M.B. *Jatropha* (Euphorbiaceae) in Southwestern United States and Adjacent Northern Mexico; University of Arizona: Tucson, AZ, USA, 1998.
- 40. Sunday, O.O., Benson C.I., Omobola, O.O., Uchechukwu, U.N. and Anthony, I.O. Antibacterial and Antioxidant Properties of the Leaves and Stem Essential Oils of *Jatropha gossypifolia* L. Biomedical Research International 2016, **1**, 939.
- Rao, A.S., Reddy, S.G., Babu, P.P. and Reddy, A.R.. The antioxidant and antiproliferative activities of methanolic extracts from Njavara rice bran *BMC Complement. Altern. Med.*, 10, 4, 2010, https://doi.org/10.1186/1472-6882-10-4.

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