

# Sociobiology

An international journal on social insects

## SHORT NOTE

# Occurrence of Gregarines (Protozoa: Apicomplexa) in the Neotropical Soldierless *Ruptitermes* spp. (Isoptera, Termitidae, Apicotermitinae)

### VANELIZE JANEI, ANA M. COSTA-LEONARDO

Laboratório de Cupins, Departamento de Biologia Geral e Aplicada, Instituto de Biociências, Universidade Estadual Paulista – UNESP, Rio Claro-SP, Brazil

#### **Article History**

#### Edited by

Og DeSouza, UFV, Brazil	
Received	31 May 2022
Initial acceptance	06 June 2022
Final acceptance	27 September 2022
Publication date	28 December 2022

#### Keywords

Parasitism; midgut; Ruptitermes pitan; Ruptitermes reconditus.

#### **Corresponding author**

Ana Maria Costa-Leonardo Laboratório de Cupins Departamento de Biologia Geral e Aplicada Universidade Estadual Paulista – UNESP Av. 24A, nº 1515, CEP: 13506-900 Rio Claro, São Paulo, Brasil. E-mail: ana.costa-leonardo@unesp.br

#### Abstract

This is the first record of gregarine infection in the worker midgut of the Neotropical termites *Ruptitermes pitan* and *Ruptitermes reconditus*. Gregarines have already been described in the gut and hemocoel in workers of other termite species, and although these protozoa are not able to kill these insects, they may affect their fitness. In the present study, gregarines were only observed in the gut lumen or linked to the midgut wall of termite workers. Histological sections of the worker midgut of *Ruptitermes* spp. showed three phases of the gregarine life cycle with the same individual worker displayed up to 4 gamonts and 2 trophozoites.

Termites are part of the most important fauna which contributes to the enhanced productivity in the tropical ecosystem (Bourguignon et al., 2016). They are grouped into lower and higher termites based on the presence of flagellated protist symbionts in the hindgut of the former and absence in the latter. Thus, the higher termites are included in the family Termitidae while lower termites comprise the other families, which are dependent on cellulolytic flagellated protozoa (Lo & Eggleton, 2011).

Gregarines are common as parasites of invertebrates, especially arthropods (Criado-Fornelio et al., 2017). These unicellular organisms may reach a length of 10 mm (Manwell, 1961) and belong to the phylum Apicomplexa, class Conoidasida and order Eugregarinorida (Adl et al., 2019). Generally, the arthropod host becomes infected with these protozoa through ingestion of mature cysts, which liberate sporozoites that reach the hemocoel and target tissue (Schimid-Hempel, 1998). Many researches showed a certain costs for hosts which harbor gregarines, for instance, in damselfly species, the presence of gregarines alters survival and egg production, while it increases susceptibility to a fungal pathogen in the cockroach Blattella germanica (Arcila & Meunier, 2020; Lopes & Alves, 2005). These protozoan parasites affect the fitness of their hosts and can cause a delayed development, decreased survival and reduced reproduction (Zuk, 1987; Cordoba-Aguilar, 2003). According to Gigliolli et al. (2016), the lack of nutrients induced by parasitic gregarines cause the already mentioned effects to the host. However, besides parasitic relationships, gregarines also can develop mutualistic or neutral association with their hosts and the type of this interaction is highly species-specific (Zuk, 1987; Rueckert et al., 2019).

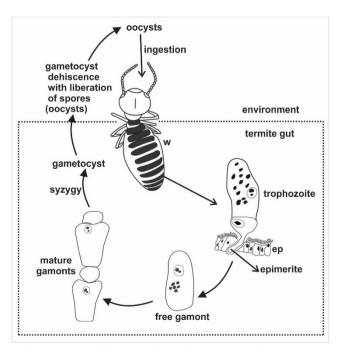


Termites are social cockroaches and according to Bouwma et al. (2005), the behavior of a social insect reflects a selection of parasites since society favors infection transmission among nestmates. However, termite societies have evolved mechanisms to limit or prevent disease agents within their colonies, making such observation unusual (Chouvenc et al., 2011). Previous records of termite gregarine infections were reported in the families Hodotermitidae (Uttangi & Desai, 1962), Rhinotermitidae (Uttangi & Desai, 1962; Huger & Lenz, 1976; Kalavati & Narasimhamurti, 1978; Hall & Hostettler, 1993; Costa-Leonardo et al., 2008), Archotermopsidae (Grassé, 1986) and Termitidae (Kalavati & Narasimhamurti, 1978; Kalavati & Narasimhamurti, 1980; Zhang et al., 2021). Gregarines present a life cycle with an endogenous stage within the insect body and an exogenous stage in the external environment (Fig 1). Infections by gregarines occur when termites and other insects ingest the parasite oocysts (mature cysts), which initiate the infection via the digestive tract (Logan et al., 2012). Then, oocysts release sporozoites, which develop into large stages named trophozoites that are attached to the intestinal wall. Later, when trophozoites mature and become gamonts, they are released from the gut wall and appear free in the gut lumen. The gamonts of opposite sex adhere to one another in a reproductive process called syzygy and form a gametocyst, which is a cyst that forms around gamont pairs. Gametocysts are then expelled in the environment together with the termite feces (Kolman et al., 2015).

The termite genus *Ruptitermes* is endemic in South America and includes 13 species and subspecies with several of them being litter-feeders (Acioli & Constantino, 2015; Constantino, 2022). *Ruptitermes pitan* and *Ruptitermes reconditus* (Apicotermitinae, Termitidae) are soldierless species of termites which have subterranean nests and forage in the open on the surface litter (Matews, 1977; Acioli & Constantino, 2015). The present study is the first record of gregarines in the Neotropical genus *Ruptitermes*, which is included in the poorly studied group Neotropical Apicotermitinae.

Foraging workers of *Ruptitermes reconditus* (Silvestri, 1901) and *Ruptitermes pitan* Acioli & Constantino, 2015 were collected in different areas located at the São Paulo State Campus of University (UNESP), Rio Claro, SP, Brazil (22°23'S, 47°32'W). For histology, 8 abdomens from each termite species were fixed in FAA (absolute alcohol, glacial acetic acid, 40% formaldehyde, in the proportion of 3:1:1) and dehydrated in increasing ethanol concentrations (70, 80, 90 and 95%). Next, the samples were transferred to a Leica historesin solution and stored for seven days in a refrigerator. The abdomens were embedded with historesin (Leica) plus catalyzer for polymerization and sectioned (3µm thick) using a Leica RM 2245 microtome. The sections were stained with toluidine blue/fuchsin and xylidine-Ponceau and documented using a Leica photomicroscope.

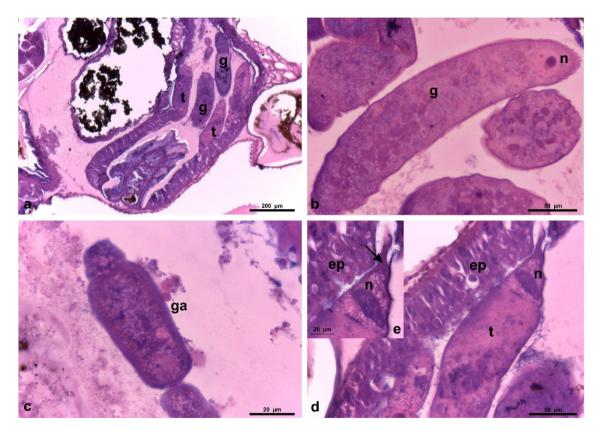
The abdomen histological sections showed the occurrence of gregarines infecting the midgut in both species of *Ruptitermes* (Fig 1). Although gregarines occupy a variety of niches inside their hosts, from extracellular cavities, as gut lumen and coelom, to epicellular and intracellular sites, these eukaryotes were not observed in the coelomic cavity or intracellularly in the gut of *Ruptitermes* spp. workers. We found 4 workers of *R. reconditus* and two of *R. pitan* infected from 8 examined specimens of each species. Gregarines have



**Fig 1**. Schematic representation of the gregarine life cycle which infect workers of *Ruptitermes* spp. Workers (w) ingest oocysts, containing sporozoites which develop into large stages named trophozoites. The trophozoites will attach to the gut epithelial cells (ep) through a special structure known as epimerite. After, the trophozoites will detach from the epithelium and form free gamonts. These free gamonts mature and undergo syzygy, originating the gametocyst that will be released into the environment.

already been described in the gut and hemocoel in workers of several termite species (Grassé, 1982; Costa-Leonardo et al., 2008), and although these protozoa do not cause termite death, they may decrease their longevity (Villanueva, 2004). Studies with wasps of the species *Polybia occidentalis* parasitized by gregarines indicated that infected individuals decrease their foraging rates and, consequently, compromise the productivity of the colony (Bouwma et al., 2005).

Free gamonts were observed in the midgut lumen in both species of *Ruptitermes* (Figs 2a, 2b and 3) and trophozoites in the inner wall of the midgut in *R. pitan* (Figs 2a and 2d). Gamonts in syzygy were also observed in the midgut lumen in *R. pitan* (Fig 2c). Previous studies with gregarine infection also displayed some of the cycle phases of as gamonts, oocysts and trophozoites in the midgut of other termites (Kalavati & Narasimhamurti, 1978; 1980; Hall & Hostettler, 1993). Additionally, the stages of oocysts and gametocysts have already been described in the hindgut of Zootermopsis angusticollis and hemocoel of Coptotermes gestroi (Henry, 1933; Costa-Leonardo et al., 2008). Free gamonts, gamonts in syzygy and gametocysts were observed only in the worker caste of the termitid Macrotermes barnevi, being absent in soldiers and reproductives (Zhang et al., 2021). Although the stages of gametocyst and sporozoites were not observed in the present study, it was possible to detect up to 4 gamonts and 2 trophozoites in the same worker. The trophozoite was attached to the mesenteric epithelium through an epimerite, which is a characteristic elongated structure, observed in R. pitan (Figs 2d and 2e). Ruptitermes spp. feed on litter and R. reconditus workers store this food in subterranean nest galleries, a fact that should facilitate the ingestion of mature cysts and, consequently, increase the infection by gregarines. However, future studies on the Ruptitermes genus will be needed to clarify this gregarine infection.



**Fig 2**. Histological sections of the midgut of *Ruptitermes pitan* worker stained with toluidine blue/fuchsin. **a** Free gamonts (g) in the midgut lumen and trophozoite (t) attached to the mesenteric epithelium. **b** Detail of the free gamont (g) in longitudinal and transversal section. **c** Gamonts in syzygy (ga). **d** Trophozoite connected to the epithelium (ep). **e** Detail of the epimerite (arrow) which is a elongated structure used by the trophozoite (t) for connection to the in epithelium (ep). **n** = nucleus

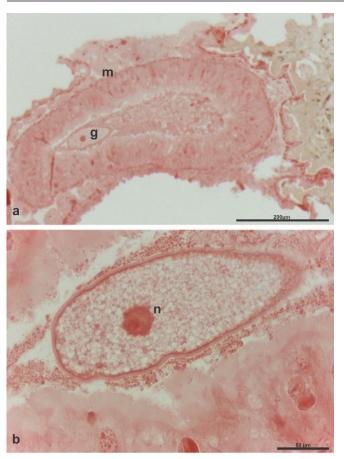
#### Acknowledgments

The authors thank to CNPq (Conselho Nacional de Desenvolvimento Científico e Tecnológico) for financial support on their termite research. They also are grateful to Iago Bueno da Silva and Varlei José Janei for valuable help with the schematic diagram.

#### **Authors' Contributions**

VJ: investigation, visualization, writing-original draft; review & editing.

AMCL: conceptualization, methodology, writing-review & editing, supervision.



**Fig 3**. Histological sections of the worker midgut (m) of *Ruptitermes reconditus* stained with xylidine Ponceau. **a**, **b** Free gamont (g) in the gut lumen. n = nucleus.

#### References

Acioli, A.N.S. & Constantino, R. (2015). A taxonomic revision of the neotropical termite genus *Ruptitermes* (Isoptera, Termitidae, Apicotermitinae). Zootaxa, 4032: 451-492. doi: 10.11646/zootaxa.4032.5.1

Arcila, F. & Meunier, J. (2020). Friend or foe? The apparent benefits of gregarine (Apicomplexa: Sporozoa) infection in the European earwig. International Journal for Parasitology, 50: 461-469. doi: 10.1016/j.ijpara.2020.01.007.

Adl, S.M., Bass, D., Lane, C.E., Lukeš, J., Schoch, C.L., Sminorv,
A., Agatha, S., Berney, C., Brown, M.W., Burki, F., Cárdenas,
P., Čepička, I., Chistyakova, L., del Campo, J., Dunthorn, M.,
Edvardsen, B., Eglit, Y., Guillou, L., Hampl, V., Heiss, A.A.,
Hoppenrath, M., James, T.Y., Karnkowska, A., Karpov, S.,
Kim, E., Kolisko, M., Kudryavtsev, A., Lahr, D.J.G., Lara, E.,
Le Gall, L., Lynn, D.H., Mann, D.G., Massana, R., Mitchell,
E.A.D., Morrow, C., Park, J.S., Pawlowski, J.W., Powell, M.J.,
Richter, D.J., Rueckert, S., Shadwick, L., Shimano, S., Spiegel,
F.W., Torruella, G., Youssef, N., Zlatogursky, V. & Zhang,
Q. (2019). Revisions to the Classification, Nomenclature, and
Diversity of Eukaryotes. Journal of Eukaryotic Microbiology,
66: 4-119. doi: 10.1111/jeu.12691.

Bourguignon, T., Šobotník, J., Dahlsjö, C.A.L. & Roisin, Y. (2016). The soldierless Apicotermitinae: insights into a poorly known and ecologically dominant tropical taxon. Insectes Sociaux, 63: 39-50. doi: 10.1007/s00040-015-0446-y

Bouwma, A.M., Howard, K.J. & Jeanne, R.L. (2005). Parasitism in a social wasp: effect of gregarines on foraging behavior, colony productivity, and adult mortality. Behavioral Ecology and Sociobiology, 59: 222-233. doi: 10.1007/s002 65-005-0028-5

Chouvenc, T., Su, N.-Y. & Grace, J.K. (2011). Fifty years of attempted biological control of termites - Analysis of a failure. Biological Control, 59: 69-82. doi: 10.1016/j.biocontrol.2011. 06.015

Constantino, R. (2022). On-Line Termites Database. Disponível em: <a href="http://www.unb.br/ib/zoo/docente/constant/catal/catnew.html">http://www.unb.br/ib/zoo/docente/constant/catal/catnew.html</a>>. Access in: 18. august. 2022

Cordoba-Aguilar, A. (2003). A description of male and female genitalia and reconstruction of copulatory and fertilization events in *Calopteryx haemorrhoidalis* (Zygoptera: Calopterygidae). Odonatologica, 32: 205-214.

Costa-Leonardo, A.M., Casarin, F.E. & Constantini, J.P. (2008). Record of a gregarine (Apicomplexa: Neogregarinida) in the abdomen of the termite *Coptotermes gestroi* (Isoptera, Rhinotermitidae). Journal of Invertebrate Pathology, 97: 114-118. doi: 10.1016/j.jip.2007.08.002

Criado-Fornelio, A., Verdú-Expósito, C., Martin-Pérez, T., Heredero-Bermejo, I., Pérez-Serrano, J., Guàrdia-Valle, L. & Panisello-Panisello, M. (2017). A survey for gregarines (Protozoa: Apicomplexa) in arthropods in Spain. Parasitology Research, 116: 99-110. doi: 10.1007/s00436-016-5266-0

Gigliolli, A.A.S., Julio, A.H.F. & Conte, H. (2016). The life cycle of Gregarina cuneata in the midgut of *Tribolium castaneum* and the effects of parasitism on the development of insects. Bulletin of Entomological Research, 106: 258-267. doi: 10.1017/S0007485315001121

Grassé, P.-P. (1982). Les glandes tégumentaires. In: Grassé P-P (ed) Termitologia, Tome I. Mason, Paris, pp 85-129.

Grassé, P.-P. (1986). Termitologia, vol III. Masson, Paris, 715p.

Hall, D. & Hostettler, N. (1993). Septate Gregarines from *Reticulitermes flavipes* and *Reticulitermes virginicus* (Isoptera: Rhinotermitidae). Journal of Eukaryotic Microbiology, 40: 29-33. doi: 10.1111/j.1550-7408.1993.tb04878.x

Henry, D.P. (1933). *Hirmocystis termitis* (Leidy) and *Kofoidina ovata* Gen. Nov., Sp, Nov. from Termites. Archiv für Protistenkunde, 80: 101-115.

Huger, V.A.M. & Lenz, M. (1976). Nachweis einer Cölomgregarine (Protozoa: Sporozoa) bei der australischen Termite *Coptotermes acinaciformis* (Isoptera: Rhinotermitidae). Zeitschrift für Angewandte Entomologie, 81: 252-258. doi: 10.1111/j.1439-0418.1976.tb04234.x Kalavati, C. & Narasimhamurti, C.C. (1978). Three new species of gregarines from termites. Proceeding of the Indian Academy of Sciences, 87B: 397-404.

Kalavati, C. & Narasimhamurti, C.C. (1980). A new species of septate gregarine, *Sphaerocystis odontotermi* n.sp. from the gut of a xylophagous termite, *Odontotermes* sp. Proceedings of the Indian Academy of Sciences, 89: 25-28

Kolman, J.A., Clopton, R.E. & Clopton, D.T. (2015). Effects of developmental temperature on gametocysts and oocysts of two species of gregarines *Blabericola migrator* and *Blabericola cubensis* (Apicomplexa: Eugregarinida: Blabericolidae) parasitizing blaberid cockroaches (Dictyoptera:Blaberidae). Journal of Parasitology, 101: 651-657. doi: 10.1645/14-673

Lo, N. & Eggleton, P. (2011). Termite Phylogenetics and Cocladogenesis with Symbionts. In D. Bignell, Y. Roisin & N. Lo (Eds.), Biology of Termites: a Modern Synthesis (pp 27-50). Springer: Dordrecht. doi: 10.1007/978-90-481-3977-4 2

Logan, J.D., Janovy, J. & Bunker, B.E. (2012). The life cycle and fitness domain of gregarine (Apicomplexa) parasites. Ecological Modelling, 233: 31-40. doi: 10.1016/j.ecolmodel.2012.03.018

Lopes, R.B. & Alves, S.B. (2005). Effect of *Gregarina* sp. parasitism on the susceptibility of *Blattella germanica* to some control agents. Journal of Invertebrate Pathology, 88: 261-264. doi: 10.1016/j.jip.2005.01.010

Manwell, R.D. (1961). Introduction to Protozoology. St. Martin's Press, New York.

Matews, A.G.A. (1977). Studies on termites from the Mato Grosso State, Brazil. Academia Brasileira de Ciências, Rio de Janeiro.

Rueckert, S., Betts, E.L. & Tsaousis, A.D. (2019). The Symbiotic Spectrum: Where Do the Gregarines Fit? Trends in Parasitology, 35: 687-694. doi: 10.1016/j.pt.2019.06.013.

Schimid-Hempel, P. (1998). Parasites in Social Insects. Princeton, NJ, 392p.

Silvestri, F. (1901). Nota preliminare sui Termitidi sudamericani. Bollettino dei Musei di Zoologia e Anatomia Comparata della Università di Torino, 16: 1-8.

Uttangi, J.C. & Desai, R.N. (1962). On some gregarines from the non xylophagous termite *Speculitermes Cyclops sinhalensis* from Dharwar, India. Journal of Animal Morphology and Physiology, 9: 97-109.

Villanueva, F.R. (2004). Generalidades y potencialidad en biocontrol de las gregarinas entomoparásitas. Ciencia UANL, 7: 355-360.

Zhang, S., Lin, Z., Huang, Q., Shen, Y. & Ni, J. (2021). First record of gregarine protists (Apicomplexa: Sporozoa) in Asian fungus-growing termite *Macrotermes barneyi* (Blattaria: Termitidae). Scientific Reports, 11: 1-9. doi: 10.1038/s41598-020-79671-7

Zuk, M. (1987). The effects of gregarine parasites on longevity, weight loss, fecundity and developmental time in the field crickets *Gryllus veletis* and *G. pennsylvanicus*. Ecological Entomology, 12: 349-354. doi: 10.1111/j.1365-2311.1987.tb01014.x

