Propagation of "Valencia" orange (*Citrus* x *sinensis* Osbeck) by minigraft





Propagación de naranja "Valencia" (*Citrus* x *sinensis* Osbeck) por mini injertos

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ABSTRACT

Keywords: Callus Citrus Nursery Rootstock Scion Union.

Demand for citrus (*Citrus* spp) plants for commercial orchards has increased worldwide due to the need for new plantations, renewal of disease-effected crops, and strict regulation for plant production. To evaluate faster and low-cost propagation alternatives for citrus, "Valencia" orange plants were propagated by using two minigrafting techniques (Cleft and inverted T-budding). Rootstocks were raised from "Cleopatra" mandarin seeds, and scions and buds were isolated from 1-year-old grafted plants. For cleft minigrafts, scions were inserted at 5-7 cm height inside of the decapitated rootstocks and covered with Eppendorf® tubes. For T-budding, buds were inserted at 5-7 cm height under the rootstock cortex cut. Unions were fixed with Parafilm®. Grafted plants were maintained under a shade house (50%) with two daily fog irrigation (2 min each). Treatments were distributed with a completely randomized design. Six weeks after grafting, the percentage of success, the shoot length, and the number of leaves per treatment were registered and analyzed with a T test (α =0.05). Cleft minigraft resulted in a higher success percentage and plants with larger shoots. Cleft minigraft could be considered an alternative for citrus propagation in small and medium size nurseries.

RESUMEN

Palabras clave: La demanda de plantas cítricas (Citrus spp) para cultivos ha incrementado mundialmente debido Callo a las necesidades de nuevas siembras, reemplazo de árboles enfermos en cultivos establecidos, y las estrictas regulaciones para la propagación de plantas. Con el fin de evaluar alternativas de Cítricos propagación más rápidas y de bajo costo para cítricos, plantas de naranja "Valencia" fueron Viveros propagadas usando dos métodos de mini injertación (Hendidura y T-invertida). Los patrones fueron Portainjerto obtenidos de mandarina "cleopatra" y las yemas fueron aisladas de plantas injertadas de 1 año de Yema edad. Para los injertos de hendidura, las yemas fueron insertadas a 5-7 cm de altura en los patrones Unión o portainjertos decapitados y se cubrieron con tubos Eppendorf[®]. Para los injertos en T-invertida, las yemas se insertaron a 5-7 cm de altura debajo de la corteza del patrón. Las uniones se fijaron con Parafilm. Los injertos se mantuvieron en una casa malla (59%) con dos riegos diarios (2 min cada uno). Los tratamientos se distribuyeron con un diseño completamente aleatorizado. Seis semanas después de la injertación, el porcentaje de éxito, la longitud de tallos y el número de hojas por tratamiento fueron registrados y analizados con la prueba de T Student (α =0.05). El mini injerto de hendidura resultó en un mayor porcentaje de éxito y tallos más largos. El mini injerto de hendidura puede considerase como una alternativa para la propagación de cítricos en pequeños y medianos viveros.

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itrus plants are one of the most important MATERIALS AND METHODS fruit crops all over the world (Wu et al., The experiment was carried during the year 2019-2020 in a shade house of the Institute of Applied Biotechnology for the Caribbean of the Universidad de Córdoba (Monteria, Colombia), located at 8° 31 N and 75° 58' W with an elevation of 12 masl. **Rootstock growth**

Seeds for rootstock production were extracted from horticulturally ripened Cleopatra mandarin (Citrus reshni hort. Ex Tanaka) fruits harvested from field grown trees at the Universidad de Córdoba – Berastegui Campus (8°40'26" N 75°46'44" W). Fruits were washed twice with distilled water, hand-squeezed and seeds separated with a plastic sieve. The extracted seeds were profusely washed with sterile-distilled water, air-dried overnight on filter paper, and stored in sterile closed glass flasks for 4 weeks in a conventional fridge at 8 °C. Germination occurred after seeds were sown in plastic tube containers (15×5 cm) filed with peat as substrate. Seedlings were maintained under shade house conditions with a 50% saran light for 6 months with fog irrigation twice a day for 1 min each.

Bud and scion selection

Grafting material was isolated from 2-year-old grafted plants of Valencia orange (*Citrus* x *sinensis* Osbeck) obtained from an authorized citrus plant distributor (Reg. ICA 25290-06V). Plants were maintained in a shade house (50%) with fog irrigation twice a day for 1 minute each.

Minigrafting and plant growth

To evaluate grafting success percentage, two types of graft, cleft, and T-budding, were performed on 6-monthold rootstocks and, approximately, 20 cm high. For cleft grafting, the rootstock was decapitated at 5-7 cm high, and a vertical downward cut was done in the center of the decapitated stem using a sterile scalpel. The scion, 2-3 cm long tender tissue containing at least one node, was cut from both sides at the basal end with the scalpel into a gently sloping wedge (~0.5 cm) where the cambium vascular tissue was observed. The scion was properly inserted in the rootstock cut, firmly tied with Nescofilm[®], and top covered with a plastic 1.5 mL Eppendorf tube for two weeks. For inverted T-budding, a 1-2 cm vertical downward cut was done with a scalpel

2018). Citrus fresh fruits and orange juice are common components of daily diets due to their high levels of vitamin C, folate, flavanones, hesperidin, and naringin, and numerous reported health benefits (Inglese and Sortino, 2019). Production of citrus in the world in 2018 was around 150 million t where oranges accounted for 50%, mandarins 22%, lemons, and limes 12% and other citrus fruits 6%. In Colombia, production in 2018 was estimated at 75000 t distributed in 20% oranges, 13% mandarins, 14% limes, lemons, and 52% other citrus fruits (FAOSTAT, 2020). In the Córdoba department, citrus fruit production, in the same year, was 1150 t, being "Valencia" orange production more than 90% (Agronet, 2021). A worldwide shortage in citrus plant supply is happening due to strict regulations during the propagation process to prevent the spread of diseases such as CTV (Citrus tristeza virus), CEVd (Citrus Exocortis Viroid) and HLB (Huanglongbing) (Wang, 2021; Vashisth et al., 2020; Folimonova, 2020; ICA, 2019). Citrus orchards are generally established with grafted plants to combine rootstock and cultivar benefits, to ensure fruit quality and uniformity, and to reduce the time for harvesting. (Talon et al., 2020; Barón et al., 2019). Minigraft is a clonal propagation technique that uses young rootstocks to be grafted with small size scion/bud parts to obtain younger plants fully adapted to field conditions, avoiding the maintenance of large size plants for scion production and the ex vitro acclimatization stage of the micropropagation process (Sigueira et al., 2016). Out of the hundreds of grafting techniques. citrus plants are usually grafted by T-budding, a timeconsuming process where a bud is removed from the desired cultivar and inserted underneath of the rootstock cortex to promote callus growth and vascular connection between the two parts; the whole process may last for 24-36 months for plants to be ready for field planting (Alves et al., 2019; Widaryanto et al., 2019). Recent studies on citrus propagation focus on speed up the propagation process and increasing the number of plants produced while complying with the official regulations (Pokhrel et al., 2021; Solonia et al., 2020). The present research aimed to evaluate two types of minigrafts, cleft and T-budding, their viability and success level on the propagation of "Valencia" orange plants as a way to obtain a faster and cost-effective citrus propagation protocol.

in the rootstock stem at about 5-7 cm high, and down terminated with a perpendicular horizontal cut. The bud was removed from young-tender stem shoots by cutting at about 0.3 cm below the bud with the scalpel and making a slicing cut down under the bud finishing about 0.5 cm beyond the but point. The bud piece was inserted by pushing it upward under the two flaps of the rootstock cut and thereafter firmly tied with Nescofilm®. Two weeks after the grafting, for plants where the scion was viable (green), the rootstock stem was chop-down about 1 cm above the grafting, and once the scion bud began to grow the rootstock stem above de scion was completely removed. Grafted plants were maintained in a shade house (50%) with fog irrigation twice a day for 2 min each. The experiment consisted of a one-way factor with two treatments (Cleft and T-budding minigrafts) and 100 replicates per treatment for a total of 300 experimental units, which were distributed with a completely randomized design. Six weeks after the grafting, for each treatment, the number of successfully grafted plants, the average length of the grafted scion shoot, and the average number

of fully expanded leaves were registered and analyzed with a T test (α =0.05).

RESULTS AND DISCUSSION Minigrafting and plant growth

The success of grafting was observed in plants propagated using cleft and T-budding minigrafts (Figure 1). Valencia orange plants propagated with cleft minigraft showed a 75% success while plants propagated by T-budding minigraft resulted in 38% success; the T-test showed that the number of successful cleft minigrafts was statically higher (P=7.77×10⁻⁶) than the number of T-budding successful minigraft showed a significant (P=0.0483) increase in shoot height, and a statistically (P=0.0005) higher number of leaves, compared to plants propagated using T-budding minigrafts (Table 1). Grafted recovered plants showed no morphological abnormality or deficient growth during the evaluation period.

Grafting is a plant propagation technique used for centuries, especially on evergreen plants (Barón *et al.*,



Figure 1. Grafted plants of "Valencia" orange by cleft (Left) and T-budding (Right) minigrafts.

Table 1. Percentage of success, shoot height and leaf number of Valencia orange plants propagated using T-budding and cleft minigrafts.

Minigraft	Number	Succeeded	Success (%)	Shoot height (cm)	Leaves (Number)
T-budding	300	114	38 b	1.62 b	2.56 b
Cleft	300	225	75 a	2.80 a	4.20 b

*Numbers with the same letter are not different according to T (Student) test (α =0.05).

2019). Massive propagation of citrus plants for crops is based on T-budding 40-50 cm rootstocks with scion buds of specific cultivars, in an 18-24 months process to obtain plant material ready to plant in field crops, after mother plants have been carefully selected (Kamanga et al., 2017). Attempts to speed up the process include the use of tissue culture techniques such as rootstock production through micropropagation (Vashisth et al., 2020) and in vitro micrografting; however, low multiplication rates and time for ex vitro plantlet recovery are still a challenge (Chamandoosti, 2020; Sangma et al., 2020). The success of grafting is founded on a vascular reconnection between rootstock and scions, a process that involves hormones, molecular factors. and even whole genome transfer at the grafting area (Rasool et al., 2020; Gautier et al., 2019). Xylem tissue formations, callus proliferation at the graft union, and vascular bundles fiber growth are reported to be regulated by auxins, cytokines, and gibberellins during graft formation (Sharma and Zheng, 2019). An increased accumulation of stilbene metabolites at the graft union as a result of a re programming of the metabolome at the graft interface to support wounding stress, callus cell proliferation and the healing process were observed when grafting grapevine plants (Prodhomme et al., 2019). Therefore, tissue regeneration that supports grafting healing, decreases with plant aging due to a lack, or reduced, expression of several transcription factors that promote the expression of products, especially auxins, that contribute to callus formation at the wound site (Ibañez et al., 2020).

Demand for citrus plants for orchard plantation is increased worldwide due to difficulties to comply with strict regulations implemented to avoid the spread of diseases or for replanting dead or declining trees (Bhandari *et al.*, 2021). In Colombia, legal measures enforce that mother plants to provide bud and scions must be isolated with anti-aphid fabric mesh to avoid incidence of pests (ICA, 2019). These measures significantly increase the cost of plant production leaving middle and small plant propagation operations out of business. Evidence of this situation is the ICA database of nurseries where in the Córdoba department appears only one nursery reported in 2021 (ICA, 2021), indicating that new citrus crops in the area are established with non-locally produced plant material. A lower disease incidence and better field performance are usually reported when crops are planted with locally produced grafted plants (Ramírez-Jiménez et al., 2020; Noor et al., 2019). Minigrafting has been used to accelerate plant propagation in fruit species (Belmonte-Ureña et al., 2020), for the diagnosis of plant diseases (Spano et al., 2020), as a strategy for molecular biology studies (Bartusch et al., 2020; Tsaballa et al., 2021) and as a mechanism for somatic embryos rescue (Raharjo and Litz, 2005). The use of minigraft in the propagation of citrus cultivars has not been previously reported. In the present study, it was observed that propagation of Valencia orange using cleft and T-budding minigraft was possible, with a 75 and 38% of success, respectively; a higher percentage of success in cleft grafting could be the result of a better contact of cambial tissues of the rootstock and the scion compared to the contact between the inserted bud and the inner layer of the rootstock cortex in T-budding; however histological analysis is recommended to completely guarantee it. Percentages of success obtained from this study are prone to increase with some adjustments when massive propagation is implemented. The minigraft technique may provide nurseries and propagators with a suitable mechanism to propagate citrus plants at the local level, reduce costs for mother plant maintenance, and speed up the propagation process by using younger rootstocks; however, it is recommended to evaluate the performance of propagated plants at the nursery and field crop level.

CONCLUSION

The results of the present study of the propagation of "Valencia" orange plants using Cleft minigraft showed a higher percentage of success compared to T-budding, and the plants recovered by this method developed a significant higher number of shoots than plants propagated by T-budding. Cleft minigraft is a viable alternative for plant propagators to produce grafted citrus plants while complying with official regulations.

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REFERENCES

Agronet. 2021. Resultados de las evaluaciones agropecuarias municipales del año 2017 del departamento de Córdoba. https://www.agronet.gov.co/Paginas/ProduccionNacionalDpto.aspx. accessed: May 2020.

Alves S, Girardi A, Alves F, Soranz R and Coletta H. 2019. Advances in citrus propagation in Brazil. Revista Brasileira Fruticultura Jaboticabal 41(6): (e-422) http://doi.org/10.1590/0100-29452019422

Barón D, Esteves A, Pina A and Ferreira G. 2019. An overview of grafting re-establishment in woody plant species. Scentia Horticulturae 243:84-91 https://doi.org/10.1016/j.scienta.2018.08.012

Bartusch K, Trenner J, Melnyk C and Quint M. 2020. Cut and paste: temperature-enhanced cotyledon micrografting for *Arabidopsis thaliana* seedling. Plant Methods 16(12):1-11. https://doi.org/10.1186/s13007-020-0562-1

Belmonte-Ureña L, Garrido-Cardenas J and Camacho-Ferrer F. 2020. Analysis of world research on grafting in horticultural plants. Hortscience 55(1):112-120. https://doi.org/10.21273/ HORTSCI14533-19

Bhandari N, Basnet M and Khanal S. 2021. Standardization of grafting time of mandarin (*Citrus reticulata* Blanco). International Journal of Fruit Science 21(1):599-608. https://doi.org/10.1080/155 38362.2021.1875964

Chamandoosti F. 2020. Citrus tissue culture with two different approaches. International Journal of Biosciences and Biotechnology 8(1): 19-30. https://doi.org/10.24843/JJBB.2020.v08.i01.p03

Folimonova S. 2020. Citrus tristeza virus (CTV): A large RNA virus with complex biology turned into valuable tool for crop protection. PLoS Pathogens 16(4). https://doi.org/10.1371/journal. ppat.1008416

FAOSTAT - Food Agricultural Organization. 2020. Cultivos. http://www.fao.org/faostat/es/#data/QC. accessed: September 2020.

Gautier A, Chambaud C, Brocard L, Ollat N, Gambetta G, Deirot S and Cookson S. 2019. Merging genotypes: graft union formation and scion-rootstock interactions. Journal of Experimental Botany 70(3):747-755. https://doi.org/10.1093/jxb/ery422

Ibañez S, Carneros E, Testillano P and Pérez-Pérez J. 2020. Advances in plant regeneration: Shake, rattle and roll. Plants 9(7):1-19. https://doi.org/10.3390/plants9070897

ICA - Instituto Colombiano Agropecuario. 2019. Resolución 12816 de 2019 "Requisitos para el registro ante el ICA de los viveros y/o huertos básicos productores y/o comercializadores de semilla sexual y/o asexual (material vegetal de propagación) de cítricos, así como los requisitos fitosanitarios para la conservación, producción, certificación y distribución de material de propagación de cítricos en viveros, en el territorio nacional". In: https://www. ica.gov.co/normatividad/normas-ica/resoluciones-oficinasnacionales/2019/2019r12816 Accessed: January 2020.

ICA - Instituto Colombiano Agropecuario. 2021. Listado de viveros registrados. In: https://www.ica.gov.co/getdoc/08d0b08ff704-4e0f-bfb2-14f861fb5215/certificacion-de-semillas.aspx . Accessed: January 2021.

Inglese P and Sortino G. 2019. Citrus history, taxonomy, breedeing and fruit quality. Oxford Research Encyclopedia of Environmental Science. 1-22. http://doi.org/10.1093/acrefore/9780199389414.013.221

Kamanga R, Chilembwe E and Chisangak. 2017. Comparative

success of budding and grafting *Citrus sinensis*: Effect of scion's number of buds on bud take, growth and sturdiness of seedlings. Journal of Horticulture 4(3):1-6 http://doi.org/10.4172/2376-0354.1000206

Noor R, Wang Z, Umair M, Yaseen M, Ameen M, Rehman S, Khan M, Imran M, Ahmed W and Sun Y. 2019. Interactive effects of grafting techniques and scion-rootstocks combinations on vegetative growth, yield and quality of cucumber (*Cucumis sativus* L.) Agronomy 9(6):1-26 http://doi.org/10.3390/agronomy9060288

Pokhrel S, Meyering B, Bowman K and Albrecht U. 2021. Horticultural attributes and root architecture of fiel-grown "Valencia" trees grafted on different rootstocks propagated by seed, cuttings and tissue culture. Hortscience 56(2):163-172. https://doi.org/10.21273/ HORTSCI15507-20

Prodhomme D, Fonayet J, Hevin C, Franc C, Hilbert G, de Revel G, Richard T, Ollat N and Cookson S. 2019. Metabolite profiling during graft union formation reveals the reprogramming of primary metabolism and the induction of stilbene synthesis at the graft interface in grapevine. BMC Plant Biology 19:599. https://doi. org/10.1186/s12870-019-2055-9

Raharjo S and Litz R. 2005. Micrografting and ex vitro grafting for somatic embryo rescue and plantlet recovery in avocado (*Persea americana*). Plant Cell Tissue and Organ Culture 82:1-9. https://doi. org/10.1007/s11240-004-5486-3

Ramírez-Jiménez A, Barrera-Sánchez C y Córdoba-Gaona O. 2020. Yield and yield components of tomato grafted plants in the high Andean region of Colombia. Revista Colombiana de Ciencias Hortícolas 14(3):375-384. https://doi.org/10.17584/rcch.2020v14i3.11671

Rasool A, Mansoor S, Bhat K, Hassan G, Baba T, Alyemeni M, Alsahli A, El-Serehy, Paray B and Ahmad P. 2020. Mechanisms underlying graft union formation and rootstock scion interaction in horticultural plants. Frontiers in Plant Sciences 11. https://doi. org/10.3389/fpls.2020.590847

Sangma S, Pereira L, Dang J and Mathew B. 2020. Evaluation of explants for *in vitro* propagation of *Citrus indica* Tanaka – An endangered species. Plant Tissue Culture and Biotechnology 30(1):87-96. https://doi.org/10.3329/ptcb.v30i1.47794

Sharma A and Zheng B. 2019. Molecular response during plant grafting and its regulations by auxins, cytokinins and gibberellins. Biomolecules 9(9):1-20. https://doi.org/10.3390/biom9090397

Siqueira G, Sales C, Rangel C, Dias B and Soares W. 2016. Production of guava mini-grafted on intra or interspecific rootstock Revista Brasileira de Fruticultura 39(1):1-6. https://doi. org/10.1590/0100-29452017635

Solonia F, Ciacciulli A, Poles L, Pappalardo H, La Malfa S and Licciardello C. 2020. New plant breeding techniques in citrus for the improvement of important agronomic traits. A review. Frontiers in Plant Sciences 11. https://doi.org/10.3389/fpls.2020.01234

Spano R, Ferrara M, Gallitelli D and Mascia T. 2020. The role of grafting in the resistance of tomato to viruses Plants 9 (8):1-20. https://doi.org/10.3390/plants9081042

Talon M, Caruso M and Gmitter F. 2020. The genus citrus. 1st Edition Woodhead Publishing, Cambridge. pp. 105-127.

Tsaballa A, Xanthopoulou A, Madesis P, Tsaftaris A and Nianiou-Obeidat I. 2021. Vegetable grafting from a molecular point of view: The involvement of epigenetics in rootstock-scion interaction. Frontiers in Plant Science 11. https://doi.org/10.3389/fpls.2020.621999

Vashisth T, Chun C and Ozores M. 2020. Florida citrus nurseriey

trends and strategies to enhance production of field-transplant ready citrus plants. Horticulturae 6(1):1-16. https://doi.org/10.3390/ horticulturae6010008

Wang N. 2021. A promising plant defense peptide against citrus Huanglongbing disease. PNAS Proceedings of the National Academy of Sciences of the United States of America 118(6). https://doi.org/10.1073/ pnas.2026483118

Widaryanto E, Humaidah A, Saitama A and Hidayatullah A. 2019.

Techniques for accelerating of scion growth in Pummelo grafting (*Citrus maxima* (Burm.) Merr.). Asian Journal of Plant Sciences 18(1):46-51. https://doi.org/10.3923/ajps.2019.46.51

Wu G, Terol J, Ibanez V, López-García A, Pérez-Román E, Borredá C, Domingo C, Tadeo F, Carbonell-Caballero J, Alonso R, Curk F, Du D, Ollitrault P, Roose M, Dopazo J, Gmitter F, Rokhsar D and Talon M. 2018. Genomics of the origin and evolution of Citrus. Nature 554:311-316. https://doi.org/10.1038/nature25447