

## Use of phenolic compounds from cocoa pod-husks (*Theobroma cacao* L.) as inhibitors of *Salmonella* spp. in fresh cheese produced in Manabí, Ecuador



Uso de compuestos fenólicos del pericarpio de cacao (*Theobroma cacao* L.) como agentes inhibidores de *Salmonella* spp. en queso fresco producido en Manabí, Ecuador

#### https://doi.org/10.15446/rfnam.v74n3.90287

Stalin Santacruz1\* and Pablo Medrano1

#### ABSTRACT

#### Keywords:

Cheese Cocoa pod-husk Fine aroma cocoa Growth curve measurement Phenolic compounds Sensory evaluation Cocoa pod-husk is a by-product of cocoa processing, underutilized despite its phenolic compounds that can be an alternative to preserve the microbiological quality of food. The aim of this work was to evaluate the in vitro inhibitory activity of phenolic compounds from the cocoa pod-husk against Salmonella spp, which is commonly found in fresh cheese produced in Manabí, Ecuador; as well as the effect on the sensory characteristics of cheese after immersion in a solution of phenolic compounds. In vitro microbiological analyzes of the inhibitory activity of phenolic compounds, showed that the concentrations 1 and 1.5% had the highest zone of inhibition against Salmonella spp., with mean diameters of 10.67 and 11.8 mm, respectively. On the other hand, the growth curve of Salmonella spp. indicated that 2 h were required for complete inhibition of bacteria by phenolic compounds at concentrations of 1 and 1.5%. For the sensory analyzes of cheese treated with phenolic compounds, 56.3% of the panelists accredited the firmness and odor with "I like it", while 37.5% of the panelists qualified the color of the cheese with "I neither like nor dislike". Firmness and odor had higher values of acceptance than color. For 25 and 12.5% of the panelists, firmness and odor were rated as "I like it a lot", respectively, and 56.3% of the panelists conferred the label of "I like it" to both attributes. Cheese color was the lowest rated attribute, given that 12.5% of the panelists chose "I like it a lot" and 25% for "I like it".

#### RESUMEN

#### Palabras clave: Queso Pericarpio de cacao Cacao fino y de aroma Curva de crecimiento Compuestos fenólicos

Análisis sensorial

El pericarpio del cacao es un subproducto generado en la transformación agroindustrial, comúnmente subutilizado, a pesar de los compuestos fenólicos presentes en él que pueden ser una alternativa para preservar la calidad microbiológica de los alimentos. El objetivo del presente trabajo fue evaluar el poder inhibitorio in vitro de compuestos fenólicos provenientes del pericarpio de cacao frente a Salmonela spp., comúnmente presente en queso fresco producido en Manabí. Ecuador: así como el efecto en las características sensoriales del gueso después de su inmersión en una solución de compuestos fenólicos. Los análisis microbiológicos in vitro del poder inhibitorio de los compuestos fenólicos mostraron que concentraciones de 1 and 1,5% tuvieron las mayores zonas de inhibición de Salmonela spp., con medias de 10,67 mm y 11,8 mm de diámetro, respectivamente. Por otro lado, la curva de crecimiento de Salmonela spp. indicó que se requirió de 2 h para una inhibición completa de la bacteria frente a concentraciones de compuestos fenólicos de 1 y 1,5%. Los análisis sensoriales del queso tratado con compuestos fenólicos presentaron que la firmeza y el olor tuvieron mejores calificaciones que el color. La firmeza y el olor se calificaron con "me gusta mucho" por 25% y 12,5% de los panelistas, respectivamente y con "me gusta" por un 56,3% de los panelistas para ambos atributos. El color del queso fue el atributo de menor calificación, con 12,5% de los panelistas que escogieron el nivel de agrado "me gusta mucho" y 25% para el nivel "me gusta".

1 Universidad Laica Eloy Alfaro de Manabí, Manta, Ecuador. stalin.santacruz@gmail.com 💿, medranopablo05@gmail.com

\* Corresponding author



ocoa tree is cultivated in tropical countries with hot and humid environments. Three countries, Cote D'Ivoire, Ghana and Indonesia together cultivate 61% and produce 67% of global traded cocoa (Campos-Vega et al., 2018), considering the top ten countries account for approximately 93% of total world cocoa production (Table 1). The average yield in countries from Latin America and the Caribbean is 0.39 t ha<sup>-1</sup> of cocoa and in the world is 0.44 t ha<sup>-1</sup> of cocoa (Sánchez et al., 2019). The main varieties of cocoa produced and sold in Ecuador are cacao fino y de aroma (fine aroma cocoa) (Clones Nacionales, Pichilingue Tropical Experimental Station, Ecuador) and CCN-51 cocoa (Castro Naranjal Collection, Guerrero, 2015). Ecuador is an important producer of fine aroma cocoa worldwide and is currently recognized for providing more than 60% of its production. Fine aroma cocoa is

recognized worldwide for its aroma, color and quality. The use of the fruit seed to produce chocolate, cosmetics, among others, triggers the generation of by-products. The agroindustry of cocoa generates between 17 and 20 kg of by-products for each 100 kg of cocoa fruit (López, 2014), which currently are used in soil fertilization (Munongo et al., 2017). Nonetheless, several phenolic compounds were identified by LC-MS/MS in cocoa pod-husk by Karim et al. (2014) such as phenolic acids, flavonoids, luteolin, apigenin and linarin. Despite cocoa pod-husk is an important source of phenolic compounds (Hii et al., 2009), with great potential for application in the field of nutrition, health and medicine, due to its antioxidant and antimicrobial characteristics (Niemenak et al., 2006), in many cases the by-products of cocoa are accumulated without any treatment, leading to the generation and transmission of diseases that may affect crops (FEDECACAO, 2013).

Table 1. Countries with the highest production of cocoa (Campos-Vega et al., 2018).

	Production (t)	% total	Cultivation (ha)	% total
Cote D'Ivoire	1,472,313	32.96	2,851,084	27.96
Ghana	858,720	19.23	1,683,765	16.51
Indonesia	656,817	14.71	1,701,351	16.69
Cameroon	291,512	6.53	723,853	7.10
Nigeria	236,521	5.30	838,046	8.22
Brazil	213,843	4.79	720,053	7.06
Ecuador	177,551	3.98	454,257	4.45
Peru	107,922	2.42	125,580	1.23
Dominican	81,246	1.82	172,940	1.70
Colombia	56,163	1.26	165,844	1.63

The fresh cheese is an Ecuadorian artisanal cheese with 37% moisture (Espinosa, 2012) and good acceptance among consumers, especially in the province of Manabí (Ecuadorian coast). The presence of pathogenic microorganisms such as *Salmonella* in this kind of cheese (Zambrano, 2014) together with commercialization of the cheese at inappropriate temperatures can affect the health of the consumer (Lee *et al.*, 2015) and contribute to a high number of people suffering from Salmonellosis in the province of Manabí, Ecuador (Ministerio de Salud Pública, 2014, 2015, 2016, 2017, 2018).

Salmonella and other pathogenic microorganisms have been found in cheese of countries such as Colombia (Instituto Nacional de Salud, 2011), Turkey (Kahraman *et al.*, 2010), France (Domínguez *et al.*, 2009), Germany (Fretz *et al.*, 2010) and Egypt (El-Baz *et al.*, 2017). To deal pathogen microorganisms, natural antimicrobials can be an important alternative to preserve the microbiological quality of food and guarantee consumer safety (Abdalla *et al.*, 2007).

Concerns about the use of synthetic chemical antimicrobials have renewed the interests of consumers using natural and

safe alternatives (Nazzaro *et al.*, 2009). The antimicrobial activity of phenolic compounds is well documented by several authors (Erdemoglu *et al.*, 2007; Xia *et al.*, 2011). The growth of strains of *Salmonella* and *Escherichia coli* is inhibited by phenolic compounds extracted from vegetables, fruits, herbs and spices (Cetin-Karaca and Newman, 2015), cocoa bean (Todorovic *et al.*, 2017) and bean shells (Nsor-Atindana *et al.*, 2012). However, no studies of the use of phenolic compounds of cocoa pod-husk against *Salmonella* growth have been performed.

In the present work, the inhibition of *Salmonella* spp. in fresh cheese was studied, after immersing the cheese in a solution of phenolic compounds previously extracted from the pericarp of fine aroma cocoa. Additionally, sensory analyzes were used to examine whether cheese treated with phenolic compounds showed organoleptic differences compared to an untreated cheese.

### MATERIALS AND METHODS

Samples of fine aroma cocoa pericarp (*Theobroma cacao* L.) were collected at the Alegría farm, located in Junín, Manabí province, Ecuador, with 0.9277 south latitude and 80.2058 western longitude. The samples were washed with water and dried in an oven at 50 °C, ground and stored for future analyzes. Fresh cheese was purchased at the Nuevo Tarqui shopping center in the city of Manta, Ecuador.

#### Alcoholic extraction of phenolic compounds

The extraction of phenolic compounds was done according to Santacruz *et al.* (2020). An amount of 15 g of the previously ground cocoa pericarp was mixed with 150 mL of ethanol (95% v/v) and kept under stirring at 130 rpm for 24 h at 25 °C. Afterwards, the sample was centrifuged (SIGMA 2-16P, Germany) for 10 min at 4000 rpm and the supernatant was filtered by vacuum, saving the filtered liquid fraction.

#### Quantification of phenolic compounds

The quantification of phenolic compounds was done according to the Folin-Ciocalteu method proposed by Mahmood *et al.* (2011). A stock solution was prepared by mixing 10 mL of the previous filtered fraction with 5 mL of ethanol (95% v/v) and distilled water to complete a total volume of 100 mL. From the stock solution, 0.1 mL was mixed with 0.5 mL of the Folin-Ciocalteau reagent, allowing it to stand for 5 min. Afterwards, 1 mL of a

sodium carbonate solution (5%) was added and made up to 25 mL with distilled water. The resulting solution was left in dark for 1 h and its absorbance at 760 nm was measured (Spectrophotometer Jenway 6320D, China). The quantification of phenolic compounds was performed using a calibration curve with gallic acid standards, and the results were expressed in mg of gallic acid equivalent (GAE)  $g^{-1}$  cocoa pericarp.

# Inhibition of *Salmonella* spp. growing by phenolic compounds

The antibacterial activity of phenolic compounds against *Salmonella* spp. was performed according to Santacruz and Castro (2018) and the Clinical and Laboratory Standards Institute (CLSI, 2009). *Salmonella* spp. was inoculated into Petri dishes using Salmonella-Shigella agar (HiMedia Laboratories, India) as culture medium and incubated at 37 °C for 24 h. Filter paper disks (Fisher Scientific Q2) of 5 mm diameter were immersed in a solution of phenolic compounds at a specific concentration (see experimental design). The disks were dried and placed in the center of the Petri dishes containing *Salmonella* spp. and incubated at 37 °C for 24 h. Afterwards, the zone of inhibition of the growth by phenolic compounds was measured in triplicate.

#### Bacterial growth curve measurement

Bacterial growth was performed according to Puupponen *et al.* (2001). 10 mL of fresh growth medium (Salmonella-Shigella agar) were inoculated with 5% of culture (frozen *Salmonella* spp. was transferred to liquid media and incubated for 24 h). Phenolic compounds, from alcoholic extraction, were added to the culture media to give final concentrations of 0.5, 1 or 5%. The culture was manually shaken, and afterwards, it was incubated at 37 °C for 24 h. Bacterial growth was followed by taking seven samples from the culture during the incubation period of 24 h (0, 0.5, 1, 2, 4, 6, and 24 h). The samples were plated in Petri dishes containing Salmonella-Shigella agar and incubated at 37 °C for 24 h before bacterial count was recorded.

#### Bacterial growth in cheese

Fresh cheese was immersed in a 1% phenolic compounds solution for 15 min. Afterwards, cheese samples were stored for 8 days, overnight under refrigeration conditions and during the daytime at room temperature (approximately  $25 \,^{\circ}$ C).

Microbiological analysis of *Salmonella* in cheese was performed on samples after 0, 2, 4, 6 and 8 days of storage, according to ISO 6579: 2002 method (ISO, 2002). For the test, 10 g of cheese were weighed and placed in a sterile resealable plastic bag, before adding 90 mL of sterile peptone water. The sample was homogenized manually, and 1 mL of the liquid was transferred to a glass test tube before adding 9 mL of peptone water (10<sup>-1</sup> dilution). The process was repeated to get a dilution of 10<sup>-4</sup>, and immediately 1 mL of the solution was inoculated into Petri dishes and incubated at 37 °C for 24 h prior CFU counting.

#### Sensory analyzes

Cheese samples were previously analyzed to verify the absence of *Salmonella*. Afterwards, the cheese was immersed for 15 min in a 1% phenolic compounds solution before sensory analyzes. First, a duo-trio test ISO 10399: 2004 (ISO, 2004) with 16 semi-trained panelists was done to determine organoleptic differences between cheese samples treated with phenolic compounds and a control sample (cheese with no immersion). Subsequently, attributes of firmness, color, odor and flavor were evaluated using a five-point hedonic scale, with scores ranging from "I like it a lot" (5) to "I dislike it a lot" (1).

#### **Texture analysis**

Puncture tests were performed according to Castro *et al.* (2017), using cheese cubes of edge 2 cm, with slight modifications. A Shimadzu texturometer (Model EZ-LX, Japan) together with a stainless-steel probe of 3 mm diameter and 8 cm length were utilized. The probe speed was 10 mm s<sup>-1</sup> and the maximum penetration force was recorded. Results were the average of three measurements.

#### Experimental design and statistical analyzes

A completely randomized design with a unifactorial arrangement was used. The concentration of phenolic compounds obtained by alcoholic extraction at 3 levels (0.5, 1 and 1.5%) was established as the independent variable (Medrano, 2019), whereas the zone of inhibition and the bacterial growth measurement were the dependent variables. The concentration of phenolic compounds with the best *Salmonella* inhibition was applied to cheese samples, before sensory analysis.

ANOVA and the significance of the difference between means was determined by Tukey test (*P*<0.05) using InfoStat statistics software (Infostat version 2014, Argentina). All measurements were performed in triplicate.

#### **RESULTS AND DISCUSSION**

The results of the inhibition of the growth of Salmonella spp. by phenolic compounds showed that there was a difference between Salmonella inhibition zones at the three concentrations of phenolic compounds (P<0.05) with values between 7.78 and 11.78 mm (Table 2). A positive correlation was also found between the inhibitory effect and the concentration of phenolic compounds by Hernández et al. (2012). The results of the present study agree with those obtained by Nsor-Atindana et al. (2012) for an ethanolic extract of cocoa pericarp at 100 mg mL<sup>1</sup> with zones of inhibition between 10.98 and 12.21 mm against Salmonella, E. coli, S. aureus and B. cereus. The inhibitory effect can be influenced by the type of phenolic compounds (Orihuela, 2016) that acts to a greater or lesser degree on the cell wall of bacteria, resulting in the loss of cell structure and the death of the microorganism. Another explanation for the inhibitory effect may be the diffusion

Table 2. Inhibition of growth of Salmonella spp. by phenolic compounds extracted from the pericarp of cocoa.

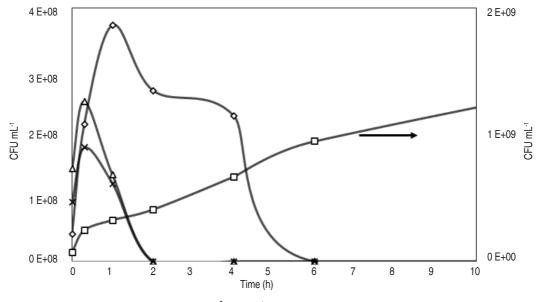
Phenolic compounds in solution (%)	Diameter of inhibition zone (mm)
0.5	7.78±1.15 a
1.0	10.67±1.32 ab
1.5	11.78±1.86 b

Mean value and standard deviations of measurements made with nine replicates. Different letters denote significant differences (P<0.05).

of hydrophilic compounds through the channel proteins of the outer membrane that surrounds the cell wall of gramnegative bacteria (Santos *et al.*, 2014) This cell wall may contribute to the entry of phenolic compounds, which may have resulted in the observed cell death.

The measurement of bacterial growth curve (Figure 1) shows an increase in CFU mL<sup>-1</sup> for the control sample along storage time. A complete inhibition of *Salmonella* spp. was observed after 2 h in the presence of phenolic compounds at 1 and 1.5%. The concentration of 0.5%

showed a complete inhibition of *Salmonella* after 6 h. Phenolic compounds presented a dose-dependent antibacterial activity (Khan *et al.*, 2021). Puupponen *et al.* (2001) showed that berry extracts at concentrations between 1 and 5 mg mL<sup>-1</sup> were strong inhibitors of *Salm. enteric.* The inhibitory effect over time may be due to affectations of the bacteria cell wall because of the presence of the phenolic group (Puupponen *et al.*, 2001). However, diffusion of phenolic compounds through the cell wall may allow an initial growth of bacteria (Figure 1) (Khan *et al.*, 2021).



 $\square$  0%,  $\Diamond$  0.5%,  $\Delta$  1%, imes 1.5%

Figure 1. Curve of Salmonella growth in the presence of solutions of phenolic compounds.

The control sample (cheese with no immersion) showed presence of *Salmonella* spp. According to Ecuadorian regulations, NTE INEN 1529-15: 2013 (INEN, 2013), *Salmonella* should not be present in cheese. Cheese samples previously immersed in phenolic compound solutions at 1% showed an absence of *Salmonella* at 0 day of storage. In fact, Figure 1 shows no presence of *Salmonella* in cheese treated with phenolic compounds at 0.5% after 2 h of storage.

#### Sensory analyzes

Results of the duo-trio test showed that 15 of 16 panelists did not find difference between cheese previously immersed in a solution of phenolic compounds and the control sample. Texture analysis

displayed that penetration force of control sample was 1.02 N, which was lower than cheese treated with phenolic compounds, showing values between 1.92 and 1.99 N (P<0.05). Instrumental texture of cheese does not have a single dominant characteristic and exhibit at least two primary characteristics (e.g. chewiness and cohesiveness) (Meullenet *et al.*, 1998).

Figure 2 shows the responses of the panelists to the five-point hedonic test about cheese treated with phenolic compounds. Firmness and odor obtained the highest ratings among the evaluated characteristics, since 56.3% of the panelists classified them as "I like it". For the flavor, 37.5% of the panelists gave a rating of "I like it" and an equal percentage for the color of the

cheese with the label of "I neither like nor dislike". The lower color rating could be due to the fact that phenolic compounds developed a yellow color in cheese, color that did not change along storage.

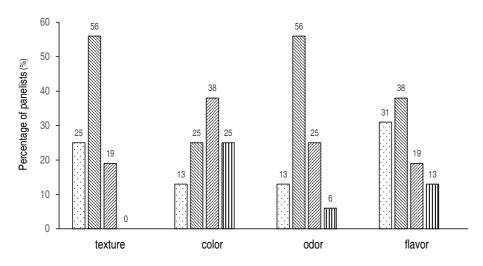


Figure 2. Sensory analyzes. Percentage of panelists and their evaluation of firmness, color, odor and flavor on cheese using a five-point hedonic scale, with scores ranging from "I like it a lot" [] "I like it" (), "I neither like nor dislike" (), "I dislike it" (), "I dislike it a lot" ().

Based on present results, it is difficult to conclude if color had influence either in odor or flavor. Moreover, visual and auditory appreciations can modify the flavor of food but are not intrinsic to it (Auvray and Spence, 2008). Koch and Koch (2008) stated that "In fact, it may be that color has nothing to do with the taste of food or drink". Meanwhile, Bayarri *et al.* (2001) suggested that "the possible influence of color on flavor perception is under discussion and no clear conclusions have been attained yet".

#### CONCLUSIONS

The solutions of phenolic compounds at concentrations of 1 and 1.5% showed the highest *in vitro* inhibition of *Salmonella* spp., requiring 2 h to achieve a complete inhibition of the bacteria.

Sensory analysis showed there was no difference between the cheese treated with phenolic compounds and a control cheese. Firmness and odor had high ratings whereas color had the lowest rating among the evaluated attributes.

#### REFERENCES

Abdalla A, Darwish S, Ayad E, and El-Hamahmy R. 2007. Egyptian mango by-product 2: Antioxidant and antimicrobial activities of extract and oil from mango seed kernel. Food Chemistry 103: 1141-1152. https://doi.org/10.1016/j.foodchem.2006.10.026 Auvray M and Spence C. 2008. The multisensory perception of flavor. Consciousness and Cognition 17: 1016-1031. https://doi. org/10.1016/j.concog.2007.06.005

Bayarri S, Calvo C, Costell E and Durán L. 2001. Influence of color on perception of sweetness and fruit flavor of fruit drinks. Food Science and Technology International 7: 399-404. https://doi.org/10.1106/ JJWN-FFRQ-JBMC-LQ5R

Campos-Vega R, Nieto-Figueroa K and, Oomah B. 2018. Cocoa (*Theobroma cacao* L.) pod husk: Renewable source of bioactive compounds. Trends in Food Science & Technology 81: 172-184. https://doi.org/10.1016/j.tifs.2018.09.022

Castro M, Mantuano M, Coloma J and Santacruz S. 2017. Utilisation of cassava starch edible films containing salicylic acid on papaya (*Carica papaya* L.) preservation. Revista Politécnica 39: 7-12. https:// revistapolitecnica.epn.edu.ec/ojs2/index.php/revista\_politecnica2/ article/view/712

Cetin-Karaca H and Newman M. 2015. Antimicrobial efficacy of plant phenolic compounds against *Salmonella* and *Escherichia coli*. Food Bioscience 11: 8-16. https://doi.org/10.1016/j.fbio.2015.03.002

CLSI. 2009. Clinical and Laboratory Standards Institute. Method for antifungal disk diffusion susceptibility testing of yeasts; Approved Guideline-Second Edition. Wayne, PA: Clinical and Laboratory Standards Institute CLSI document M44–A2. 23 p. In: Clinical and Laboratory Standards Institute. Method for antifungal disk diffusion susceptibility testing of yeasts; Approved Guideline-Second Edition. Wayne, PA: Clinical and Laboratory Standards Institute CLSI document M44–A2.

Domínguez M, Jourdan-Da Silva N, Vaillant V, Pihier N, Kermin C, Weill F, Delmas G, Kerouanton A, Brisabois A and de Valk H. 2009. Outbreak of *Salmonella enterica* Serotype Montevideo Infections in France linked to consumption of cheese made from raw milk. Foodborne Pathogens and Disease 6: 121-128. https://doi.org/10.1089/fpd.2008.0086

El-Baz A, El-Sherbini M, Abdelkhalek A and Al-Ashmawy M.

2017. Prevalence and molecular characterization of *Salmonella serovars* in milk and cheese in Mansoura city, Egypt. Journal of Advanced Veterinary and Animal Research 4: 45-51. https://doi. org/10.5455/javar.2017.d189

Erdemoglu N, Ozkan S and Tosun F. 2007. Alkaloid profile and antimicrobial activity of *Lupinus angustifolius* L. alkaloid extract. Phytochemistry Reviews 6: 197-201. https://doi.org/10.1007/s11101-006-9055-8

Espinosa C. 2012. Estudio del queso manaba y su aplicación gastronómica. (Tesis). Universidad Técnica Equinoccial, Quito, Ecuador. 124 p.

FEDECACAO. 2013. Federación Nacional de Cacaoteros. Guía ambiental para el cultivo del cacao. Segunda edición. Ministerio de Agricultura y Desarrollo rural. Colombia. 126 p.

Fretz R, Sagel U, Ruppitsch W, Pietzka A, Stöger A, Huhulescu S, Heuberger S, Pichler J, Much P and Pfaff G. 2010. Listeriosis outbreak caused by acid curd cheese Quargel, Austria and Germany 2009. Eurosurveillance 15: 1-2. https://doi.org/10.2807/ese.15.05.19477-en

Guerrero A. 2015. Sistema de Información Nacional Agropecuario. Panorama agroeconómico del Ecuador una visión del 2015. Ministerio de Agricultura, Ganadería, Acuacultura y Pesca. Quito, Ecuador.

Hernández J, Cubillos, J y Milian P. 2012. Aislamiento de cepas de *Rhizobium* spp. asociados a dos leguminosas forrajeras en el Centro Biotecnológico del Caribe. Revista Colombiana de Microbiología Tropical 2: 51-56.

Hii C, Law C, Suzannah S, Misnawi J and Cloke M. 2009. Polyphenols in cocoa (*Theobroma cacao L.*). Asian Journal of Food and Agro-Industry 2(4): 702-722. https://www.ajofai.info/Abstract/ Polyphenols%20in%20cocoa%20(theobroma%20cacao%20l.).pdf

INEN. 2013. Instituto Ecuatoriano de Normalización. Norma Técnica Ecuatoriana NTE INEN. 1529–15:2013. Control microbiológico de los alimentos. *Salmonella*. Método de detección. Quito, Ecuador. 17 p.

Instituto Nacional de Salud. 2011. Evaluación de riesgos de *Listeria monocytogenes* en queso fresco en Colombia. Ministerio de Salud y Protección Social. Bogotá, Colombia. 67 p.

ISO. 2002. International Standard ISO 6579:2002. Microbiology of food and animal feeding stuffs. Horizontal method for the detection of *Salmonella* spp. International Organization for Standardization. 27 p

ISO. 2004. International Standard ISO 10399:2004. Sensory analysis: Methodology: Duo-trío test, Second Edition. International Organization for Standardization. 19 p

Kahraman T, Ozmen G, Ozinan B and Omer Goksoy E. 2010. Prevalence of *Salmonella* spp. and *Listeria monocytogenes* in different cheese types produced in Turkey. British Food Journal 112: 1230-1236. https://doi.org/10.1108/00070701011088214

Karim A, Azlan A, Ismail A, Hashim P, Abd Gani S, Zainudin B and Abdullah N. 2014. Phenolic composition, antioxidant, anti-wrinkles and tyrosinase inhibitory activities of cocoa pod extract. BMC Complementary and Alternative Medicine 14:381. https://bmccomplementmedtherapies.biomedcentral.com/ articles/10.1186/1472-6882-14-381

Khan F, Javed S, Hanif A, Faizi S, Yousuf M and Kazmi S. 2021. Antimicrobial activity of phenolic compounds isolated from *Caesalpinia pulcherrima*. Research Square 1:1-12. https://doi.

org/10.21203/rs.3.rs-154525/v1

Koch C and Koch E. 2008. Preconceptions of taste based on color. Journal of Psychology: Interdisciplinary and Applied 137: 233-242. https://doi.org/10.1080/00223980309600611

Lee K, Runyon M, Herrman T, Phillips R and Hsieh J. 2015. Review of *Salmonella* detection and identification methods: Aspects of rapid emergency response and food safety. Food control 47: 264-276. https://doi.org/10.1016/j.foodcont.2014.07.011

López V. 2014. El tratamiento de desechos sólidos de derivados del cacao y la productividad en la planta de chocolates más choco. (Tesis de maestría). Universidad Técnica de Ambato. Ambato, Ecuador. 159 p.

Mahmood A, Ngah, N and Omar M. 2011. Phytochemicals constituent and antioxidant activities in *Musa x paradisiaca* flower. European Journal of Scientific Research 66, 331-318.

Medrano P. 2019. Utilización de compuestos fenólicos de la cáscara del cacao (*Theobroma cacao*) como agentes inhibidores de *Salmonella* spp. en queso fresco. (Tesis). Universidad Laica Eloy Alfaro de Manabí. Manta, Ecuador. 43 p

Meullenet J, Lyon B, Carpenter J and Lyon C. 1998. Relationship between sensory and instrumental texture profile attributes. Journal of Sensory Studies 13:77-93. https://doi.org/10.1111/j.1745-459X.1998. tb00076.x

Ministerio de Salud Pública. 2014. Gaceta epidemiológica Ecuador. Ministerio de Salud Pública. Quito, Ecuador. 40 p.

Ministerio de Salud Pública. 2015. Gaceta epidemiológica Ecuador. Ministerio de Salud Pública. Quito, Ecuador. 27 p.

Ministerio de Salud Pública. 2016. Gaceta epidemiológica Ecuador. Ministerio de Salud Pública. Quito, Ecuador. 31 p.

Ministerio de Salud Pública. 2017. Gaceta epidemiológica Ecuador. Ministerio de Salud Pública. Quito, Ecuador. 19 p.

Ministerio de Salud Pública. 2018. Gaceta epidemiológica Ecuador. Ministerio de Salud Pública. Quito, Ecuador. 21 p.

Munongo M, Nkeng G and Njukeng J. 2017. Production and characterization of compost manure and biochar from cocoa Pod Husks. International Journal of Advanced Scientific Research and Management 2:26-31. http://ijasrm.com/wp-content/ uploads/2017/02/IJASRM\_V2S1\_185\_26\_31.pdf

Nazzaro F, Caliendo G, Arnesi G, Veronesi A, Sarzi P and Fratianni F. 2009. Comparative content of some bioactive compounds in two varieties of *Capsicum annuum* L. Sweet pepper and evaluation of their antimicrobial and mutagenic activities. Journal of Food Biochemistry 33:852-868. https://doi.org/10.1111/j.1745-4514.2009.00259.x

Niemenak N, Rohsius Ch, Elwers S, Ndoumou D and Lieberei R. 2006. Comparative study of different cocoa (*Theobroma cacao L.*) clones in terms of their phenolics and anthocianins contents. Journal of Food Composition and Analysis 19: 612-619. https://doi. org/10.1016/j.jfca.2005.02.006

Nsor-Atindana J, Zhong F and Mothibe K. 2012. Quantification of total polyphenolic content and antimicrobial activity of cocoa (*Theobroma cacao L.*) bean shells. Pakistan Journal of Nutrition 11(7): 672-677.

Orihuela J. 2016. Actividad inhibitoria del extracto etanolico de *Theobroma cacao L*. sobre el crecimiento y adherencia in vitro de *Streptococcus mutans* a esmalte dentario. (Tesis). Universidad Mayor de San Marcos. Lima, Perú. 166 p

Puupponen R, Nohynek L, Meier C, Kahkonen M, Heinonen M,

Hopia A, and Oksman K. 2001. Antimicrobial properties of phenolic compounds from berries. Journal of Applied Microbiology 90: 494-507. https://doi.org/10.1046/j.1365-2672.2001.01271.x

Sánchez V, Zambano J y Iglesias C. 2019. La cadena de valor del cacao en América Latina y El Caribe. FONTAGRO, ESPOL, INIAP. Quito. 100 p.

Santacruz S, Cárdenas G and Mero V. 2020. Phenolic compounds and oil of orange and passion fruit seeds. Rev. Fac. Agron. (LUZ) 37: 51-68. https://produccioncientificaluz.org/index. php/agronomia/article/view/32289/33698

Santacruz S and Castro M. 2018. Viability of free and encapsulated *Lactobacillus acidophilus* incorporated to cassava starch edible films and its application to Manaba fresh white cheese, Manta, Ecuador. Food Science and Technology 93: 570–572. https://doi.org/10.1016/j.lwt.2018.04.016

Santos R, Oliveira D, Sodré G, Gosmann G, Brendel M and

Pungartnik C. 2014. Antimicrobial activity of fermented *Theobroma cacao* pod husk extract. Genetics and Molecular Research 13: 7725-7735. https://doi.org/10.4238/2014.September.26.10

Todorovic V, Milenkovic M, Vidovic B, Todorovic Z and Sobajic S. 2017. Correlation between antimicrobial, antioxidant activity, and polyphenols of alkalized/nonalkalized coccoa powders. Journal of Food Science 82: 1020-1027. https://doi.org/10.1111/1750-3841.13672

Xia D, Wu X, Shi J, Yang Q and Zhang Y. 2011. Phenolic compounds from the edible seeds extract of Chinese Mei (*Prunus mume* Sieb. et Zucc) and their antimicrobial activity. LWT - Food Science and Technology 44:347-349. https://doi.org/10.1016/j. lwt.2010.05.017

Zambrano M. 2014. Determinación de la presencia de *Salmonella* en queso fresco comercializado en el cantón Chone provincia de Manabí entre mayo y julio del 2014. (Tesis). Universidad Católica de Santiago de Guayaquil. Guayaquil, Ecuador. 41 p