



Metals bioremediation potential using Pseudokirchneriella subcapitata

Potencial de biorremediação de metais pela microalga *Pseudokirchneriella subcapitata* Mônica Ansilago¹ , Franciéli Ottonelli¹ , Emerson Machado de Carvalho²

ABSTRACT

Microalgae are unicellular organisms, photosynthesizers that present cell duplication exponentially and biosorption capacity of nutrients dissolved in water. The objective of this work was to evaluate the capacity of the microalga Pseudokirchneriella subcapitata for bioremediation of metals and salts. In this aspect, the reduction of the metals and salts in the synthetic effluents by the microalga P. subcapitata was evaluated: (T1) culture medium (control); (T2) culture medium contaminated with aluminum chloride; (T3) culture medium contaminated with ferrous sulfate; (T4) culture medium contaminated with zinc sulfate; (T5) culture medium contaminated with the combination of aluminum chloride, ferrous sulfate and zinc sulfate. The bioremediation process was evaluated by comparing culture media with suspended microalgae to a filtrate version of the same medium. Iron and zinc metals, as well as nitrogen and phosphorus salts, showed depleted values in the filtered medium, indicating efficiency in the treatment of water by microalgae. Aluminum content was below the limit of detection in all treatments. The cumulative values in the microalgae biomass were, in descending order: nitrogen, zinc, iron and phosphorus, thus indicating the assimilation of the contaminants in the algal biomass. In addition, high biomass production of the microalgae was observed. The highest production rate was verified in the synthetic effluent with the association of metals, indicating a synergy between contaminants, which was probably responsible for reducing the toxic effect on the microalgae. These results indicated high potential for bioremediation by microalga P. subcapitata, besides the possibility of using algal biomass for biotechnological applications.

Keywords: adsorption; biosorption; Chlorophyceae; contaminants.

RESUMO

As microalgas são organismos unicelulares, fotossintetizadores, que apresentam duplicação celular exponencial e capacidade de biossorção de nutrientes dissolvidos na água. O objetivo deste trabalho foi avaliar a capacidade de biorremediação de metais e sais pela microalga Pseudokirchneriella subcapitata. Nesse aspecto, avaliou-se a redução de metais e sais nos efluentes sintéticos pelas microalgas P. subcapitata: (T1) meio de cultura (controle); (T2) meio de cultura contaminado com cloreto de alumínio; (T3) meio de cultura contaminado com sulfato ferroso; (T4) meio de cultura contaminado com sulfato de zinco; (T5) meio de cultura contaminado com a combinação de cloreto de alumínio, sulfato ferroso e sulfato de zinco. O processo de biorremediação foi avaliado comparando o meio de cultura com microalgas em suspensão e o mesmo meio filtrado. Metais de ferro e zinco, assim como sais de nitrogênio e fósforo, apresentaram valores esgotados no meio filtrado, indicando eficiência no tratamento da água por microalgas. O teor de alumínio ficou abaixo do limite de detecção em todos os tratamentos. Os valores acumulados na biomassa de microalgas, em ordem decrescente, foram nitrogênio, zinco, ferro e fósforo, indicando assim a assimilação dos contaminantes na biomassa de algas. Além disso, foi observada alta produção de biomassa das microalgas. A maior taxa de produção foi verificada no efluente sintético com a associação de metais, indicando sinergia entre contaminantes, provavelmente responsável pela redução do efeito tóxico nas microalgas. Esses resultados indicaram alto potencial de biorremediação pela microalga P. subcapitata, além da possibilidade de utilização de biomassa de microalgas para aplicações biotecnológicas.

Palavras-chave: adsorção; biossorção; Chlorophyceae; contaminantes.

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Introduction

Population growth and the development of new production technologies generate continuous environmental impacts. In this scenario, there is a possibility of increasing concentration of contaminants in wastewater, which would require adequate treatment for these effluents (Mahapatra et al., 2014), with the goal of reducing environmental impacts. These contaminants may present a wide variety of organic and inorganic materials, including toxic metals that pose high risks to aquatic environments (Carvalho et al., 2012; Zhang et al., 2016).

In order to reduce the amount of contaminants in wastewater, conventional measures have been adopted, but these are often unsustainable from an environmental, economic and energetic point of view. Thus, there is great interest in the use of microorganisms/ biological materials in effluent treatment processes, due to economic feasibility and low risk to the environment. The use of microorganisms in bioremediation processes is considered a clean and sustainable technology. In addition, this technique makes it possible to recycle nutrients efficiently and add value to the biomass produced (Ramachandra et al., 2013).

Microalgae have high applicability in the field of biotechnology, due to the high rate of biomass production and higher growth tendency (two to ten times) in relation to terrestrial plants. Consequently, the ability of these organisms to absorb solar energy increases the fixation of CO_2 by their metabolism (Priyadarshani and Rath, 2012; Sathasivam et al., 2019), and it can increase the production of carbohydrates, proteins, amino acids, lipids and other compounds of interest in algal biomass. Data from the literature suggests high efficiency in the use of microalgae for the reduction of contaminants resulting to effluent treatment. It is possible to observe representative values in the removal of mineral salts (Leong et al., 2018; Mohammadi et al., 2018; Ridley et al., 2018; Saavedra et al., 2018), metals (Peng et al., 2017; Saavedra et al., 2018; Shen et al., 2018), pesticides (González et al., 2012), pharmaceutical compounds (Escapa et al., 2017), oils (Ammar et al., 2018), among others.

Microalgae have been noted for producing lipids (Abdelaziz et al., 2014), proteins, carbohydrates, pigments and carotenoids (β -carotene, lutein, chlorophyll, etc.), vitamins (A, B1, B6, folic acid, etc.), antioxidants (catalases, polyphenols, etc.) and other interesting molecules. These bioactive compounds are essential inputs for the food, pharmaceutical and cosmetic industries. In addition, they are an energy source (Priyadarshani and Rath, 2012) and present high market value due to the low costs of the production process. In this strategy, it is possible to use the organism for production of bioactive compounds and surfactants, for different uses in biofuels, biofertilizers, biopolymers, biofilms, among many others (Carvalho et al., 2012; Schmitz et al., 2012; Gouveia et al., 2016).

Some factors are important to regulate the kinetics growth of microalgae and contribute with high production of algal biomass; for instance, the concentration of nutrients present in the medium, the luminosity and the temperature (Carvalho et al., 2012; Wang et al., 2014; Ansilago et al., 2016). Currently, one of the major obstacles in the production of industrial scale microalgae lies in the high cost of culture medium. An alternative culture medium that has been widely used is chemical fertilizer based in nitrogen, phosphorus and potassium (NPK), due to the high concentrations of micro and macronutrients. Besides, it presents low cost, availability in the market and facility in the preparation of culture medium (Sipaúba-Tavares et al., 2009; Carvalho et al., 2012; Ansilago et al., 2016). In order for it to become even more economically attractive, it is necessary to add value to the cultivation process.

Different species of microalgae were used in wastewater treatment processes. Leong et al. (2018) obtained results of removal of up to 98% of nitrogen in domestic effluent using the microalga *Chlorella vulgaris*. Saavedra et al. (2018) tested the removal of metals in effluent, obtaining efficiency in manganese (99.4%), arsenic (40.7%), barium (38.6%), zinc (91.90) and copper (88%) removal, using the microalgae *C. vulgaris*, *Scendesmus almeriensis* and *Chlorophyceae* spp. The microalga *C. vulgaris* was also used for mercury bioremediation, obtaining removal values of 62.85 and 94.74% (Peng et al., 2017).

The species *Pseudokirchneriella subcapitata* has been reported in nutrient removal studies concerning elements such as nitrogen and phosphorus (Gonçalves et al., 2016), as well as in toxicity studies on media containing toxic metals (Gao et al., 2016; Sousa et al., 2018). It is a half moon-shaped, single-celled green algae, with a single chloroplast containing chlorophyll *a* and *b* (Granados et al., 2008). This microalga has been used in biotechnological trials to evaluate its tolerance and production under conditions of contamination by toxic metals (Lima, 2010; Carvalho et al., 2012). The data obtained is indicative of the potential of *P. subcapitata* in the complementary treatment of domestic or industrial wastewater.

Although the microalga *P. subcapitata* presents desirable characteristics for wastewater bioremediation, experiments in controlled environments are still needed to evaluate its ability to remove nutrients of water, whether salts and/or metals, as well as to evaluate the production of algal biomass in liquid media substrates with the presence of contaminants. For this purpose, the objective of this study was to evaluate the bioremediation capacity of metals and salts by *P. subcapitata* microalgae in laboratory culture.

Materials and Methods

The *P. subcapitata* inoculum was obtained from the Laboratory of Algae Physiology at Universidade Federal de São Carlos (UFS-Car), isolated from the Broa Dam (São Carlos, SP, Brazil). The microalga was cultured and maintained in standard medium CHU₁₂ (Chu, 1942) in the laboratory of the Center for Biodiversity Research (CPBio) at Universidade Estadual de Mato Grosso do Sul (UEMS/Dourados, MS, Brazil). The cultivation system was static nonaxenic, with constant aeration and room temperature ($22 \pm 2.0^{\circ}$ C). The tests were maintained in a BOD incubator with photoperiod control of 2,500 LUX provided by white fluorescent lamps (12 h light/ 12 h dark).

The culture medium of the microalgae was prepared by adding 1 mL of NPK stock solution to 1 liter of distilled water that was autoclaved at 120°C for 20 minutes (*e.g.*, Ansilago et al., 2016). The NPK stock solution was prepared with 0.70 gL⁻¹ of chemical fertilizer N:P:K (20-5-20 gL⁻¹), according to Sipaúba-Tavares and Rocha (1993) and Carvalho et al. (2012). The rates of daily growth were obtained by Equation 1:

$$(N_{n}-N_{1})/T$$
(1)

Where:

 $\rm N_{_n}$ = the algal density value at the desired sample time (number of microalgae cells);

N₁ = the algal density value at the initial time of the experiment (number of microalgae cells);

T = the desired sample time (days).

The elaboration of each treatment is described in Table 1. Treatments were performed in triplicate. The value used from each contaminant was established based on the limit allowed by CONAMA's Resolution nº 357/2005 (Brasil, 2005), which was doubled.

For the metal analysis, 100 mL was collected from each Erlenmeyer flask at the beginning of the test (day 1) and at the end of the test (day 21). On day 1, the sample was collected before insertion of *P. subcapitata*. On the last day of the test, the samples were divided into two equal fractions: one filtered (Whatman microfiber filter, chemically inert, with porosity of 0.45 μ M) and another unfiltered (Figure 1), in order to get the Percentage of nutrient removal (% R) between suspension medium and filtered medium on day 21.

The analysis of metals (zinc, aluminum and iron) was determined by flame atomic absorption spectrometry techniques, EAA-flame, according to Welz and Sperling (1999). The total phosphorus analyzes were read by visible ultraviolet spectroscopy, UV-VIS, according to Soares et al. (2001). For the total nitrogen analyses, the Kjeldhal microdistillation technique was used, as described in Mantovani et al. (2005), and then titrated by sodium hydroxide (NaOH); subsequently, the value consumed was converted to mg L⁻¹ of mineral nitrogen, according to Tedesco et al. (1995).

To evaluate the potential bioremediation in each treatment, the concentration values of each nutrient (mg L^{-1}) were compared in the microalgae suspension samples and in the filtered samples. For this, the percentage of removal was used through Equation 2:

$$\%R = \frac{C_0 - C_e}{C_0} (100) \tag{2}$$

Where:

 C_0 and C_e = the concentrations of nutrients in the liquid phase (mg L⁻¹) on the 21st day, when microalgae reach the stationary phase of growth, in the suspended material and in the filtrate one, respectively. The data was evaluated via analysis of variance (ANOVA) and Tukey's test, in the statistical program GENES, version DOS and Visual Basic 5.0.

The chemical soil analysis manual of the Paraná Agronomic Institute (Pavan et al., 1992) was used as methodology for the analyses. The laboratory analyses were performed in the Laboratory of Environmental and Instrumental Chemistry at Universidade Estadual do Oeste do Paraná (Unioeste), Marechal Cândido Rondon, PR, Brazil.

Results and Discussion

To evaluate the bioremediation potential of the *P. subcapitata* microalgae, the capacities of production in the contaminated medium and removal of contaminants were considered.

The daily growth rate of *P. subcapitata* indicated that the treatment contaminated with aluminum, iron and zinc simultaneously (T5) showed the best algal biomass doubling rate. In addition to the control, this treatment was significantly superior to the other treat-

Treatments	Adw	NPK	MPs	Contaminants		
				AlCl ₃	FeSO ₄	ZnSO ₄
T1	400	50	50	-	-	-
T2	400	50	50	0.32	-	-
Т3	400	50	50	-	0.32	-
T4	400	50	50	-	-	0.6
T5	400	50	50	0.32	0.32	0.6

Table 1 – Composition of treatments used to evaluate microalgae bioremediation by *P. subcapitata*.

Adw: autoclaved distilled water (mL); NPK: cultivation medium with nitrogen, phosphorus and potassium 20-5-20 g L⁻¹, respectively (mL); MPs: inoculum with the microalgae *P. subcapitata* (mL); contaminants: synthetic effluent based on aluminum chloride (AlCl₃), iron sulphate (FeSO₄) and zinc sulphate (ZnSO₄) (g L⁻¹).

ments in the final period of the experiment (21st day of experiment). Treatments contaminated only with aluminum (T2), iron (T3) or zinc (T5) showed significantly lower growth rates than control over almost the entire experimental period (Table 2).

Table 2 also shows the analyses resulting from growth curves of *P. subcapitata*. The analysis of the algal growth curves by the covariance analysis indicated that there was no significant difference between the control and the treatment contaminated with aluminum,

iron and zinc simultaneously (T5), corroborating the observations mentioned above. The exponential growth rate (K) indicated better productivity for treatment contaminated simultaneously with the metals and for control, while treatment with zinc contamination presented poor performance. The specific growth rate (μ max) also indicated satisfactory production values for control and treatment contaminated with metals simultaneously, as well as treatment contaminated only with iron (T3).



Figure 1 – Schematization of the methodology used for analysis of metals and salts present in microalgae in suspension and filtered culture medium. FAAS: Flame Atomic Absorption Spectrometry; UV-VIS: ultraviolet-visible spectrophotometry.

Table 2 – Analysis of variance (ANOVA) of the daily growth rate of *P. subcapitata* microalgae (mean \pm standard error) of (T1) culture medium (control); (T2) culture medium contaminated with aluminum chloride; (T3) culture medium contaminated with ferrous sulfate; (T4) culture medium contaminated with zinc sulfate; (T5) culture medium contaminated with the combination of aluminum chloride, ferrous sulfate and zinc sulfate, in each sampling period, followed by analysis of covariance (Ancova), exponential growth rate (k) and specific maximum growth (μ max) of the 21 days of experiment*.

	Anova		Treatments					
	F	р	T1	T2	Т3	T4	Т5	
3 nd	11.1	< 0.05	4.8 a ± 0.16	$-3.4 c \pm 0.83$	$-2.1 \text{ c} \pm 0.61$	$0.1 \text{ b} \pm 0.55$	$-2.2 \text{ c} \pm 0.52$	
6 th	53.4	< 0.05	$2.2~b\pm0.37$	$2.3 b \pm 0.19$	$0.1~\mathrm{c}\pm0.43$	$0.4 c \pm 0.33$	4.8 a ± 0.23	
9 th	13.5	< 0.05	6.0 a ± 0.16	$1.9 \text{ b} \pm 0.1$	$1.1 \text{ b} \pm 0.54$	$-2.0 \text{ c} \pm 0.16$	$2.2 \text{ b} \pm 0.25$	
12^{th}	43.4	< 0.05	$3.2 \text{ ab} \pm 0.06$	$2.4~\mathrm{b}\pm0.45$	$0.2 c \pm 0.22$	$0.3 c \pm 0.12$	$4.0 a \pm 0.31$	
15^{th}	48.0	< 0.05	$3.0 a \pm 0.47$	$0.3 c \pm 0.38$	$1.2 \text{ bc} \pm 0.13$	$0.5 c \pm 0.10$	$2.3 \text{ ab} \pm 0.09$	
18^{th}	71.5	< 0.05	$2.2~ab\pm0.35$	$2.3 \text{ ab} \pm 0.25$	$1.6~\mathrm{b}\pm0.17$	0.0 ± 0.19	3.3 a ± 0.20	
21 st	99.3	< 0.05	$4.6 a \pm 0.28$	2.9 b ±0.009	$1.4 c \pm 0.25$	$1.0 c \pm 0.04$	5.3 a ± 0.19	
Ancova	14.19	< 0.001	AB	BC	BC	С	А	
k			0,19	0,13	0,10	0,051	0,24	
μmax			0,91	0,69	0,89	0,50	0,90	

*Analysis of variance at 95% confidence followed by Tukey's test, represented by lowercase letters in comparison in the lines, where equal letters indicate statistically equal means and different letters have statistically different means between them. Analysis of covariance in the algal growth curve at 99% reliability, where upper case letters indicate statistically equal means, and different letters have statistically different means between them. It was verified, however, that the growth curves of the algal biomass have an exponential behavior only for the medium without contaminant (control) and for the medium contaminated with all the metals simultaneously. It has been observed that all trace elements, even those with biological function, such as zinc, when in higher concentrations, can cause toxicity to organisms. On the other hand, the treatment contaminated with aluminum, iron and zinc (T5) presented the best production, superior even to the control one. In this case, it is possible to observe an antagonistic dynamic, in which the effect of zinc on the exposure of other chemicals, such as iron and aluminum, resulted in the reduction of its toxic effect to microalgae, bringing about effects different from those expected for the action of contaminants alone, which result from synergistic, potentiation, antagonistic and additive interactions (Mozeto and Zagato, 2008).

Another important factor to be analysed in the production of microalgae and in the process of contaminant bioremediation is the monitoring of the pH of the culture medium. It is observed in Figure 2 that zinc and iron contamination raised the pH of the culture medium. However, a pH buffering in the culture medium is observed on the 3rd day of experiment. It is possible to verify the extent to which microalgae absorbed nutrients from the medium, carried out chemical reactions and excreted residues, tending to alter its acid-base balance, which can be verified by pH fluctuation. Carvalho et al. (2012) observed that the microalga *P. subcapitata* had considerable growth in an acidic medium, even playing a role in capping the medium.

The increase in density of the algal biomass increases the fixation of CO_2 through photosynthesis, providing greater dissociation of carbonate (CO_2^{-2}) and bicarbonates (HCO_2^{-}) ions, which induces the removal of carbonic acid, and may even precipitate metals in the form of carbonates, followed by the release of OH⁻ ions for the neutralization of the medium (Mota and Von Sperling, 2009; Gardner et al., 2011). All these chemical reactions may explain the increase and subsequent stabilization of pH in all treatments.

Table 3 presents the percentage of removal of metals and salts during the process of biomass production of the microalgae *P*.



Figure 2 – Ionic potential of hydrogen in culture media used for the production of *P. subcapitata*.

	Fe	Al	Zn	Ν	Р
T1	$39.7^{\text{B}} \pm 1,1$	< LD	$-600.1^{\circ} \pm 98,1$	$78.9^{\text{A}} \pm 1.6$	$19.6^{\text{ns}} \pm 0.9$
T2	$-8.7^{\circ} \pm 18,7$	< LD	-433.3 ^c ± 38,5	$21.9^{\circ} \pm 3.4$	$9.2^{ns} \pm 1,1$
Т3	75.3 ^A ± 3,4	< LD	$-183.3^{\rm BC} \pm 67.3$	$6.9^{\text{D}} \pm 4.0$	$17.5^{ns} \pm 3.7$
T4	$-16.9^{\circ} \pm 18,7$	< LD	$-46.6^{\text{B}} \pm 81.3$	$46.0^{\text{B}} \pm 1.6$	$12.8^{ns} \pm 4.3$
T5	$27.3^{\text{B}} \pm 6.9$	< LD	$88,9^{A} \pm 1,8$	$57.2^{\text{B}} \pm 0.9$	$13.4^{ns} \pm 3.8$
Р	< 0.05	-	< 0.05	< 0.05	0.25
F	84.10	-	11.41	121.50	1.59

Table 3 – Percentage of nutrient removal (% R) between suspension medium and filtered medium on day 21 of experiment (mean \pm standard error).

T1: control – no contaminants; T2: treatment contaminated with aluminum; T3: treatment contaminated with iron; T4: treatment contaminated with zinc; T5: treatment contaminated with aluminum + iron + zinc; < LD: elements that have an absorbance value below the detection limit. The analysis of variance (p < 0.05) was performed, followed by Tukey's test in comparison with the lines, where the same letters indicate statistically equal means and different letters present statistically different means.

subcapitata adsorbed or (bio)sorbed by microalgae. In the analysis of the iron content in the culture medium, values of removal were significantly higher in treatment contaminated with iron (T3), followed by control (T1) and treatment contaminated by all metals simultaneously (T5). Thus, it is possible to infer that the microalgae is able to adsorb or (bio)sorb iron from the medium, even in high concentrations. The capture of metal ions involves some biosorption mechanisms, which are based on ion exchange, coordination, complexation, adsorption and chemical precipitation (Silva et al., 2013).

The results of the aluminum analysis in the medium were below the limit of detection in all treatments, rendering the biore-

Main microalgae used	Objective	Growing medium Main results		Reference	
<i>Oocystis</i> sp.	Biomass production and	Effluent from	Removal of up to 32% sulfate	Mohammadi et al.	
Chorella sp.	sulfate removal	power plant	Biomass production of 50 mg L ⁻¹	(2018)	
Scenedesmus intermedius	Adaptation and removal of lindane	BG11	Resistance by rare mutations after exposure period up to 40 mg L ⁻¹ ; subsequent removal by up to 99% of lindane	González et al. (2012)	
Chlorella vulgaris	Simultaneous cultivation with activated sludge for bioremediation and lipid productionRemoval of up to 98% of nitrogen; lipid yield up to 130 mg L-1		Leong et al. (2018)		
Chlorella vulgaris	Absorption of arsenic,		Removal of 99.4% for manganese	Saavedra et al. (2018)	
Scenedesmus almeriensis	boron, copper, manganese and zinc from water by	Synthetic medium based on the water composition of the Loa river (Chile).	40.7% for arsenic 38.6% for boron		
Chlorophyceae spp.	different green microalgae		91.9% for zinc 88% for copper		
Chlorella vulgaris	Bioremediation of mercury by mineralized microalgae	Blue green medium (BG11) contaminating with mercury	Removal of mercury from 62.85% to 94.74%	Peng et al. (2017)	
Nannochloropsis oculata	Oil removal and Chemical	Oil effluent	Removal up to 89% of oil and up to 90% of COD	Ammar et al. (2018)	
Isochrysis galbana	oxygen demand (COD)		Removal of up to 82% of oil and up to 83% of COD		
Phaeodactylum tricornutum	Growth capacity in nitrate- containing medium	Modified F/2 medium	Removal of up to 1700 mg L ⁻¹ of nitrate in 100 L bioreactor	Ridley et al. (2018)	
Chlorella sp.	Efficiency in removal of cadmium	Immobilization in pellets derived from a complex of the botanical genus <i>Eichhornia</i>	Removal up to 92.45% cadmium	Shen et al. (2018)	
Chlorella sorokiniana	Response of microalgae to high concentrations of paracetamol (PC) and salicylic acid (AS)	Mann and Myers Medium	Removal up to 69% for PC and up to 98% for AS	Escapa et al. (2017)	

Table 4 – C	Compilation	of studies in	bioremediation	using different	t microalgae.
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mediation process inconclusive. However, it is possible that aluminum is able to associate with other elements present, making the reading by the method adopted inefficient.

The percentage of zinc removal was effective only in the treatment contaminated with all metals (T5), which showed a reduction of almost 90%. Other treatments, including treatment contaminated with only zinc (T4), had negative removal values. Gao et al. (2016) observed, in their study with P. subcaptata, an elevation of zinc, without toxic potential, in the microalgae with greater exposure to phosphorus, due to potentially induced by Zn-P complexation or precipitation inside the cell. Saavedra et al. (2018) also observed removal of up to 91.9% for zinc in a study carried out with microalgae of the class Chlorophyceae. Other studies indicate that dead algae biomass may be even more efficient at retaining and accumulating metal elements than living cells and tissues (e.g., Cossich, 2000). This process occurs due to the changes in the nature of the cell surface because of the absence of active transport of the dead microalgae, causing better adsorption of metals efficiency by the algal biomass.

The removal of nitrogen in the culture medium was significantly higher in the control (T1). The value of phosphorus removal was also higher in the control, but it did not differ significantly from the previous treatments. It was possible to observe that, in treatments contaminated with metals, the biosorption or adsorption of salts of nitrogen and phosphorus was low. The maximum reduction value recorded was 78.9% for nitrogen and 19.6% for phosphorus, all in the control treatment. This fact may have resulted from the Redfield ratio ($C_{106}H_{118}O_{45}N_{16}P$), which means that algae, on average, demand 16 times more nitrogen than phosphorus (Redfield, 1958; Sperling, 2001). In a study by Wang et al. (2014), a reduction of up to 95% and 95.7% for phosphorus and nitrogen, respectively, was observed in a wastewater treatment system using the microalgae genera *Chlorella* and *Micractinium*.

Based on the results, it is possible to observe a complex relationship between microalgae bioremediation processes and the antagonistic and synergistic effects of the contaminants, with each other and with the microalgae itself. However, it is important to establish standards of control and monitoring for these contaminants in the effluent, so that it is possible to understand the metabolism and the efficiency of this system. In Table 4, we present a compilation of studies using different species of microalgae for the specific purpose of culturing, adaptation and bioremediation in media containing metals and other organic and inorganic compounds that can be biosorbed. Through these studies, it can be established that wherever there was insertion and/or presence of stressors in the culture medium of the microalgae, positive results were obtained for the removal of these contaminants and consequent increase in the production of algal biomass. This corroborates the results obtained in the present study, since the treatment contaminated with all metals (iron, aluminum and zinc) (T5) obtained a higher rate of algal biomass duplication, which can indicate synergy between the contaminants, thus reducing the toxic effects.

Conclusion

The experimental results showed that the microalga *P. sub-capitata* presented bioremediation capacity (either by adsorption and/or biosorption) and algal biomass production. The contaminants with higher concentration indices in the culture medium obtained a higher percentage of removal (N > Zn > Fe > P > Al). It was also possible to observe that, on a bench scale, the microalgae were able to develop in the presence of the toxic metals inserted in the medium.

In addition, the highest production rate was verified in the synthetic effluent with the association of metals, indicating possible synergy between aluminum, iron and zinc, which was probably responsible for reducing the toxic effect on the microalgae, meaning biomass gain for further biotechnological applications.

However, more detailed studies are needed to improve the technique and the methodology used, requiring shorter test times and the use of other contaminating metals for influence and toxicity analysis.

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Contribution of authors:

Ansilago, M.: Conceptualization, Methodology, Validation, Formal analysis, Investigation, Resources, Data curation, Writing – original draft, Writing – review & editing, Visualization, Project administration, Funding acquisition. Ottonelli, F.: Conceptualization, Methodology, Validation, Formal analysis, Investigation, Resources, Data curation, Writing – original draft. Carvalho, E.M.: Conceptualization, Methodology, Validation, Formal analysis, Investigation, Resources, Data curation, Writing – original draft. Carvalho, E.M.: Conceptualization, Methodology, Validation, Formal analysis, Investigation, Resources, Data curation, Writing – original draft, Writing – review & editing, Visualization, Supervision, Project administration.

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