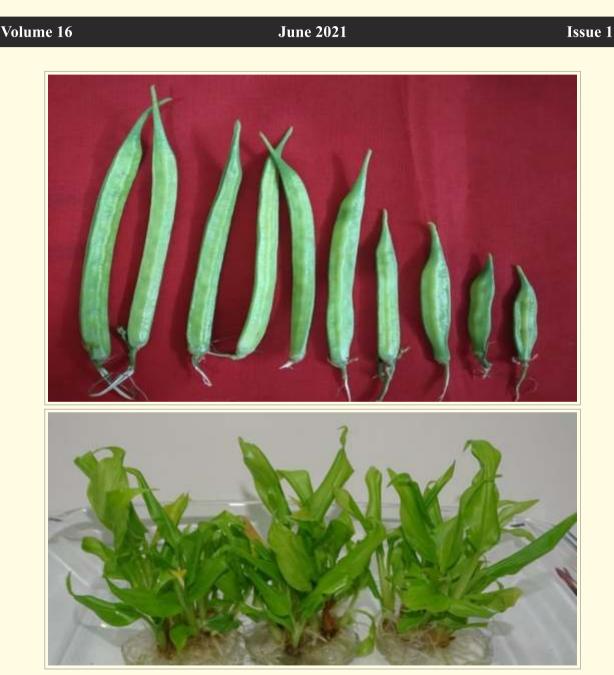
JOURNAL OF HORTICULTURAL SCIENCES





Society for Promotion of Horticulture ICAR - Indian Institute of Horticultural Research, Bengaluru - 560 089



JOURNAL OF HORTICULTURAL SCIENCES

Volume 16	Issue 1	June 2021						
CONTENTS								
In this Issue		i-ii						
Review								
tree holding remark	, Raj Kumar, Kumar A., Rani B., Phogat D.S.,	1-13						
Original Researc	ch in Papers							
(Flacourtia montana	l evaluation of mountain sweet thorn J. Grah) collections an S., Radhika V. and Shetti D.L.	14-25						
		26-35						
e e e e e e e e e e e e e e e e e e e	tudies in amaranthus (<i>Amaranthus</i> spp.) S., Lakshmana D., Nadukeri S. and Hanumanthappa M.	36-44						
iron deficiency and t (Solanum tuberosum	al parameters associated with chlorosis resistance to their effect on yield and related attributes in potato L.) Sharma J., Raveendran M. and Sudhakar D.	45-52						
deficit conditions	nt Okra (<i>Abelmoschus esculentus</i>) cultivars to water Hussain I., Naveed K., Ali S., Mehmood A., Khan M.J.,	53-63						
[Cyamopsis tetragono	for yield and its attributing traits in cluster bean bloba (L.) Taub] through gamma irradiation a S., Sood M., Aghora T.S., Anjanappa M., Rao V.K. and Reddy A.B.	64-68						
cytokinin source and	n protocol for <i>Curcuma mangga</i> : Studies on carbon, d explant size 2., Karthika Devi R. and Pixy J.	69-76						



Effect of fungicide and essential oils amended wax coating on quality and shelf life of sweet orange (<i>Citrus sinensis</i> Osbeck) Bhandari M., Bhandari N. and Dhital M.	77-90
Post-harvest quality and quantification of betalains, phenolic compounds and antioxidant activity in fruits of three cultivars of prickly pear (<i>Opuntia ficus-indica</i> L. Mill) Gonzalez F.P.H., Saucedo V.C., Guerra R.D., Suarez E.J., Soto H.R.M. Lopez J.A., Garcia C.E. and Hernandez R.G.	91-102
Soil microbial community dynamics as influenced by integrated nutrient management practices in sweet basil (Ocimum basilicum L.) cultivation Baraa AL-Mansour and D. Kalaivanan	103-113
Effect of spectral manipulation and seasonal variations on cut foliage production and quality of <i>Philodendron (Philodendron</i> 'Xanadu') Sujatha A. Nair, Laxman R.H. and Sangama	114-120
Short Communications	
Studies on mutagenic sensitivity of seeds of pummelo (<i>Citrus maxima</i> Merr.) Sankaran M., Kalaivanan D. and Sunil Gowda D.C.	121-124
Isolation and characterization of microsatellite markers from Garcinia indica and cross species amplification	125-129

Ravishankar K.V., Vasudeva R., Hemanth B., Nischita P., Sthapit B.R., Parthasarathy V.A. and Rao V.R.

Original Research Paper



Responses of different okra (Abelmoschus esculentus) cultivars to water deficit conditions

Ayub Q.^{*1}, Khan S. M.¹, Hussain I.¹, Naveed K.², Ali S.¹, Mehmood A.², Khan M.J.¹, Haq N. U.³ and Shehzad Q.³

> ¹Department of Horticulture, ²Department of Agronomy, ³Department of Food Science and Technology, The University of Haripur, Pakistan *Corresponding author e-mail: qasimayub.alizai@gmail.com

ABSTRACT

A pot experiment was conducted to evaluate the adverse effects of drought on different okra cultivars and to identify the most suitable okra cultivar for growing in drought conditions. Five okra cultivars namely Pusa Green, Clemson, Sabz Pari, Pusa Swani and Mehak Pari were subjected to three drought levels i.e., control (100% Field capacity),50% and 25% Field Capacity (FC). Physiological parameters like fresh and dry weight of plant and plant height were recorded along with biochemical attributes such as chlorophyll content (a, b, total) carotenoids, total protein, proline content, and Membrane stability index (MSI%). Results showed that drought significantly reduced all the studied parameters and at maximum drought (25% FC) lowest values of fresh weight (12.42g), dry weight (1.22g), plant height (7.86cm), chlorophyll a (9.02mg/g FW), chlorophyll b (18.69mg/g FW), total chlorophyll (27.71mg/g FW), carotenoids (11.80mg/g FW), total protein (2.73mg/g FW), whereas maximum Proline (21.36µg/g FW), and MSI (72%) were observed under the same drought. The results concerning responses of okra cultivars under drought conditions showed that maximum. Fresh weight (15.25g) and Dry weight (2.74g) was observed in Pusa green while maximum Plant height (13.77cm), Chlorophyll a (14.38mg/g FW), Chlorophyll b (24.41mg/g FW), Total Chlorophyll (38.80mg/g FW), Carotenoids (18.57mg/g FW), Total Protein (5.44mg/g FW), Proline (27.78µg/g FW), and MSI (56.33%) were produced by Sabz Pari. Hence it can be concluded that drought causes significant variation on physical and biochemical attributes of okra whereas Sabz Pari showed resistance towards the applied stress and produced better results.

Keywords: Drought, Okra, Oxidative Stress and Proline content

INTRODUCTION

Okra (*Abelmoschus esculentus* L.), belongs to the family Malvaceae, is an important vegetable crop grown during summer season in Pakistan for its nutritious edible pods. It is favored for soft and tender green pods, which are commonly consumed as curries and boiled vegetables (Mounir *et al.*, 2020). Despite their astonishing beautiful flowers, okra pods are rich source of nutrients and medicinal properties. The fresh okra fruit contains carbohydrates (9.6%), protein (2.25%), fiber (1.1%), fat (0.2%) and minerals such as magnesium iron, potassium, calcium, sodium, zinc, nickel and manganese (Khan and Rab 2019).

Fiber found in okra pods reduces cholesterol and risk of cardiac diseases and promotes healthy digestive track. Okra helps in slow absorption of sugar, hence can be consumed as anti-diabetic food (Nawaz *et al.*, 2020).

Many environmental conditions including droughts have much harmful effects on the growth and yield of agricultural crops (Ayub *et al.*, 2018). During plants exposure to drought, many physiological and biochemical alterations occur inside plants on cellular level. These changes include accumulation of ABA, reduction in leaf area and closing of stomata (Meise





et al., 2018). Drought also decreases the rate of leaf growth by making cell walls sclerotic and reduced plant biomass. Like any other abiotic stresses, under drought stress proline accumulates inside plant which helps plants to with stand under stress conditions (Lintunen et al., 2020). Plants which are exposed to drought stress also exhibits lower levels of carbohydrates and starch (Qu et al., 2019). During drought stress protein degradation starts and reduction in chlorophyll takes place (Dawood et al., 2019). Drought stress mostly causes accumulation of ROS (Reactive Oxygen Species) which leads to oxidative stress in chlorophyll and disrupts normal working of plants cells (Stanley and Yuan 2019). ROS also damages lipids, terpenoids, carbohydrates and nucleic acids (Guo et al., 2018).

Okra is one of the most important summer vegetables commonly cultivated in tropical and subtropical plans of Pakistan including Haripur due to its higher nutritional values, ease of cultivation and resistance to harsh environmental conditions. Most of the cultivated lands in Haripur region are rainfed, thus the farming relies mainly on rainfall for water. During the summer followed by low rainfall seasons, these regions face moderate to severe droughts during summer in case of low rainfall. Despite of huge potential of okra production low water availability to the crop causes a great reduction on yield per area and total area under cultivation. Hence this study was conducted to evaluate the adverse effects of drought on different okra cultivars and to identify the most suitable okra cultivar for growing in drought conditions.

MATERIALS AND METHODS

Current study was conducted at Horticulture nursery, Department of Horticulture, The University of Haripur during February-March 2020. For this experiment pots of 20 cm diameter and 15cm depth were used. Each pot was filled with 2kg potting media containing soil, sand and farmyard manure in equal ratio (1:1:1). These pots were kept inside the rainout shelters during night time and during rain to avoid the entry of water to plants. Seeds of five okra cultivars namely Pusa Green, Clemson, Sabz Pari, Pusa Swani and Mehak Pari were collected from National Agriculture Research Council, Islamabad. The experiment was laid out in a split-plot factorial arrangement with Completely Randomized Design with three replications. The drought stress was taken as main plot whereas factorial arrangements of verities were placed as a sub plot.

Imposing drought treatment

Eight seeds of okra were planted in each pot. 10 days after germination plants were thinned to 5 plants per pot. Plants were irrigated up to full field capacity (FC) during 14 days after germination to achieve maximum germination and equilibrium in plant growth, afterward okra plants were subjected to three levels of drought i.e.,100% FC (control/normal irrigation), 50% FC and 25% FC. Field capacity was calculated based on saturation percentage as described by Wilcox (1951).Uniform cultural practices were carried out throughout the research period. After 20 days of drought treatment, plants were uprooted and phenotypic characters viz. plant height, fresh weight, and dry weight were measured as per the standard procedures.

Biochemical analysis

Chlorophyll and carotenoids content was measured by the method explained by Lichtenther (1987). About 0.2 g of grounded leaf sample was extracted with 80% acetone till the residue becomes colourless. Absorbance of the acetone extract was measured at 470, 663 and 646 nm with the help of spectrophotometer (from Germany, Cary-50). Chlorophyll and carotenoids concentration was then measured by following formula.

Chla =
$$(12.25A_{663.2}-2.79A_{646.8})$$

Chlb = $(21.21A_{646.8}-5.1A_{663.2})$
ChlT = chla + chlb
Carotenoids = $\frac{1000A_{470}1.8chla85.02chlb}{198}$

Soluble protein content in the okra leaf sample was measured by the method described by Lowry *et al.* (1951).

Method of Bates *et al.* (1973) was implemented to evaluate the proline content. 5g plant grounded plant sample was mixed in 3% sulfosalicylic acid (aqu) and the mixture was centrifuged at 10,000 rpm. The supernatant was mixed with 2 mL of ninhydrin) and 2 mL of glacial acetic acid, and the solution was boiled for 1h at 100 °C. The reaction is allowed to take place in ice bath and after the completion of



reaction 4mL toluene to extract the mixture and absorbance at 520nm were read by using spectrophotometer.

MSI was measured as per Premachandra *et al.* (1991). Fresh leaf material (1.0 g) was cut into small discs, washed with deionized water and placed in glass test tubes along with blank. 10 mL of deionized water was added to each test tube so that leaf discs get submerged. The test tubes were kept in a water bath for 30 min at 45 °C. After cooling, the electrical conductivity of water (C1) was measured using the conductivity meter. Water was again poured back to the same leaf discs and kept in the water bath at 100 °C for 10 min. The final electrical conductivity was used to calculate membrane stability index using the following formula.

MSI (%) = [1 - (C1/C2)] *100

where,

C1 = Initial electrical conductivity C2 = Final electrical conductivity

The experimental data were subjected to analysis of variance (ANOVA) using windows software Statistix 8.1 with two-factor factorial arrangements. Each treatment was replicated three times. The effects of drought on okra varieties were determined by the Least Significant Difference test (LSD) at pd"0.05, where the F test was significant (Steel and Torrie 1960).

RESULTS

Study was conducted to evaluate the adverse effects of a drought on different okra cultivars and to identify the most suitable okra cultivar for growing in drought conditions. The results obtained for various plant phenotypical and biochemical parameters studied are given in the Table 1.

Okra Cultivars	Plant Height	Fresh Weight	Dry Weight	Chlorop hyll a	Chlorop hyll b	Chlorop hyll total	carote noids	Protein	Proline	MSI	
Pusa Green	11.66c	15.25a	2.74a	12.26b	23.30b	35.56b	16.31b	5.44a	23.58b	51.88b	
Clemson	12.22bc	14.82b	2.65a	9.33c	20.50c	29.88c	13.30c	5.44a	20.34c	55.22a	
Sabz Pari	13.77a	14.23c	2.47c	14.38a	24.41a	38.80a	18.57a	5.44a	27.78a	56.33a	
Pusa Swani	13.33a	14.00c	2.41b	8.51d	16.52d	25.03d	10.20d	5.33a	15.78d	55.88a	
Mehak Pari	12.88ab	13.93c	2.45b	5.43e	13.51e	18.94e	6.91e	4.55a	11.57e	55.77a	
LSD	1.0448	0.3645	0.1166	0.2278	0.4419	0.2790	0.2371	0.7861	0.3912	0.9740	
Drought											
Control	17.73a	16.63a	3.70a	10.98a	20.39a	31.37a	14.28a	8.06a	18.47c	35.93c	
50% FC	12.73b	14.28b	2.72b	9.99b	19.60b	29.85b	13.09b	5.00b	19.61b	56.86b	
25% FC	7.86c	12.42c	1.22c	9.02c	18.69c	27.71c	11.80c	2.73c	21.36a	72.26a	
LSD	0.8093	0.2823	0.0903	0.1765	0.1976	0.2161	0.1837	0.6089	0.3030	0.7545	
LSD (C×D)	1.8097	0.6313	0.2019	0.3946	0.2551	0.4833	0.4107	1.3615	0.6776	1.6871	

Table 1: Comparison of okra varieties and drought stress on growth and biochemical attributes

The results are presented as mean based on n=3, LSD Least significant differences at (p<0.05). Different letters within column indicate significant differences between okra varieties and drought stress (p<0.05)

Plant height (cm)

Plant height is the major character significantly affected due to drought. In the present study, maximum plant height (17.73cm) was observed in control whereas lowest plant height (7.86cm) was recorded in plants subject to 25% FC. Okra cultivar Sabz Pari exhibits highest height (13.77cm) whereas lowest height (11.66 cm) was recorded in Pusa green (Table 1). The combined effects of drought and variety showed that Sabz Pari exhibited highest plant height among all other cultivars under all droughts. Maximum plant height 18, 15 and 8.33cm was recorded in Sabz Pari at 100%, 50% and 25% drought stress respectively,



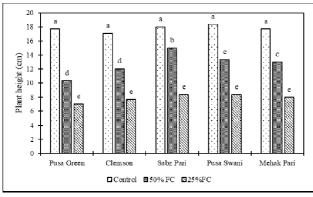


Fig. 1: Effect of drought stress on plant height of okra cultivars

whereas lowest plant height 17.66, 13 and 8 cm was observed in Mehak Pari under 100%, 50% and 25% drought. (Fig. 1).

Fresh weight (g)

Highest plant fresh weight (16.63g) was recorded at control (100%FC) whereas, lowest values (12.42g) were observed in plants subjected to water stress of 25% FC. Analysis of variance showed highly statistical difference (p < 0.01)among okra cultivars for fresh weight and applied drought. Okra cultivar Pusa green showed maximum fresh weight (15.25g) whereas minimum fresh weight (13.93g) was obtained by Mehak Pari (Table 1). Results regarding interaction of drought and okra cultivars indicates that all okra cultivars showed reduction in their fresh weight as drought stress is elevated. Highest fresh weight (17.20g) was recorded in okra cultivar Pusa Green and Clemson under control condition (100% FC) whereas 25% FC drought stress significantly reduced the fresh weight and lowest values of fresh weight (11.26g) were observed in okra cultivar Mehak Pari, meanwhile under same drought level okra cultivars Pusa Green while Clemson retained maximum biomass and showed maximum fresh weight of 14.20g and 13.3g, respectively (Fig. 2). Results of our experiment correspond with findings of Idrees et al. (2010), Singh and Usha (2003) and Munir et al. (2016) who observed similar reduction in fresh weight of lemon grass, wheat and okra, respectively, when exposed to elevating drought.

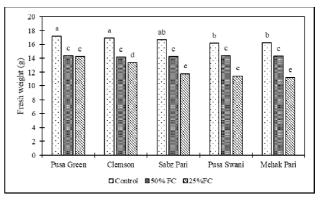


Fig. 2: Effect of drought stress on fresh weight of okra cultivars

Dry Weight of Plants (g)

Results regarding plant dry weight indicate that highest plant dry weight (3.70g) was recorded at control (100% FC) whereas lowest dry weight (1.22g) was observed in plants subjected to maximum water stress of 25% FC. Okra cultivar Pusa green showed maximum dry weight (2.74g) whereas minimum dry weight (2.41g) was obtained for Pusa Swani (Table 1). The interaction effect between drought and okra cultivars suggested a prominent reduction in dry weight for all cultivars. At 100% FC (normal irrigation) and maximum dry weight (3.86g) was recorded in Pusa Green while under same condition minimum dry weight (3.5g) were recorded in Pusa Swani. Meanwhile at highest drought stress (25%FC) least dry weight (0.76g) was recorded by Mehak Pari while under same drought Pusa green produced highest dry weight (1.6g) (Fig. 3). Similar results were observed by Idrees et al. (2010), Singh and Usha (2003) and Munir et al. (2016) in lemon grass, wheat and okra respectively.

Chlorophyll a (mg/g FW)

It was noted that maximum Chlorophyll-a content (10.98 mg/g FW) were recorded in 100% FC whereas minimum Chlorophyll-a content (9.0 mg/g FW) were observed in 25% FC. Okra cultivar Sabz Pari had highest Chlorophyll-a content (14.38 mg/g FW) while lowest values (5.43 mg/g FW) were recorded in Mehak Pari (Table-1). The combined results of drought and okra varieties revealed that there is a gradual reduction in chlorophyll content with increase in drought stress. At 100% FC (normal irrigation), okra cultivars Sabz Pari had maximum chlorophyll (15.23mg/g) while Mehak Pari had lowest chlorophyll content (6.46mg/g) ,. At 25% FC, Sabz Pari



maintained highest chlorophyll content (13.76 mg/g) while Mehak Pari produced lowest values of chlorophyll (4.56 mg/g) (Fig. 4). A reduction in chlorophyll content was also reported in drought stressed cotton (Massacci *et al.*, 2008) and *Catharanthus roseus* (Jaleel *et al.*, 2009).

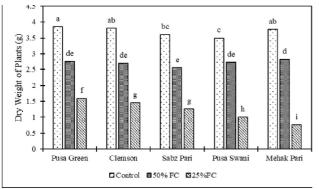


Fig. 3: Effect of drought stress on Dry weight of okra cultivars

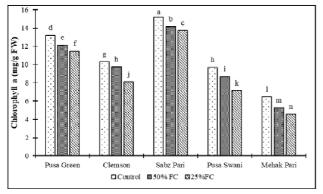


Fig. 4: Effect of drought stress on Chlorophyll a content of okra cultivars

Chlorophyll b (mg/g FW)

Maximum Chlorophyll b content (20.39 mg/g FW) was recorded in 100% FC (normal irrigation) whereas minimum Chlorophyll b content (18.6 mg/g FW) were observed in 25% FC drought. Okra cultivar Sabz Pari had highest Chlorophyll b content (24.41 mg/g FW) while lowest values (13.51 mg/g FW) was recorded in Mehak Pari (Table 1). The combined results of drought and okra varieties revealed that there was a gradual reduction in chlorophyll content with increase in drought stress. At 100% FC (normal irrigation) okra cultivars Sabz Pari showed maximum chlorophyll b (25.7mg/g) while a same irrigation lowest chlorophyll b content (14.56mg/g) were observed in Mehak Pari, on the other hand at highest drought (25% FC) Sabz Pari maintained highest chlorophyll b

content (13.76mg/g) while Mehak Pari produced lowest values of chlorophyll b (4.56mg/g) (Fig. 5). The chlorophyll content decreased to a significant level at higher water deficits sunflower plants (Kiani *et al.*, 2008) and *Vaccinium myrtillus* (Tahkokorpi *et al.*, 2007).

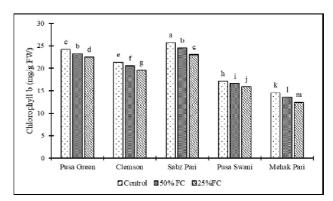


Fig. 5: Effect of drought stress on Chlorophyll-b content of okra cultivars

Total chlorophyll content (mg/g FW)

Effects of drought on total chlorophyll content indicate that maximum Total chlorophyll content (31.37 mg/g FW) was noted under no drought stress (normal irrigation) whereas minimum total chlorophyll (27.71 mg/g FW) was recorded under 25% FC drought (Table 1). Results concerning total chlorophyll content of okra cultivars reveled that Sabz Pari produced highest total chlorophyll content (38.80 mg/g FW) while lowest values (18.94 mg/g FW) were recorded in Mehak Pari (Table 1). The combined results of okra and drought stress revealed that significant reduction was observed with increase in drought stress. Highest total chlorophyll of 40.93, 38.66 and 36.6 mg/g was recorded in Sabz at 100%, 50% and 25% FC conditions, while lowest chlorophyll content 21.03, 18.83 and 16.96 mg/g was obtained by Mehak Pari at 100%, 50% and 25% FC (Fig. 6). Ram et al. (2014) and Amin et al. (2009) also observed that under drought conditions, reduction in chlorophyll content was noted in watermelon and okra respectively.

Carotenoids (mg/g FW)

Results regarding carotenoids (mg/g FW) of okra plant depicts highly significant statistical difference (p < 0.01) among okra cultivars, applied drought and their interaction (Table 1; Fig. 7). It was noted that



maximum carotenoids (14.28mg/g FW) were recorded in 100% FC (control) whereas minimum carotenoids values (11.08 mg/g FW) were observed in 25% FC drought stress. Among the okra cultivars studied maximum carotenoids (18.57 mg/g FW) were observed in Sabz Pari while lowest concentration of carotenoids (6.91 mg/g FW) was recorded in Mehak Pari (Table 1). The combined effect of drought stress and okra cultivars for carotenoids suggested that okra cultivar Sabz Pari showed highest carotenoids (20.43, 18.4 and 16.9 mg/g at 100%, 50% and 25% FC drought, respectively), while lowest carotenoids (5.26,7.23 and 5.26 mg/g at 100%, 50% and 25% FC respectively) were observed in Mehak Pari (Fig. 7). Similar findings were previously obtained by Altaf et al. (2015) in okra, Idrees et al. (2010) in lemon grass and by Ram et al. (2014) in water melon.

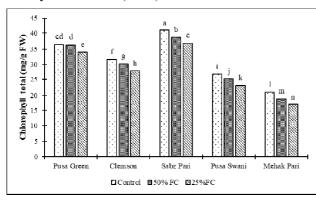


Fig. 6: Effect of drought stress on total Chlorophyll content of okra cultivars

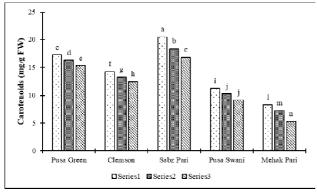


Fig. 7: Effect of drought stress on Carotenoids of okra cultivars

Total Protein (mg/g FW)

It was noted that maximum proteins (8.06 mg/g FW) were recorded in 100% FC whereas minimum protein (2.73 mg/g FW) were observed in 25% FC drought. Results regarding okra cultivars suggested non-significant variation for protein content, maximum

protein (5.44 mg/g FW) were observed in Pusa green, Clemson, Sabz Pari and in Mehak Pari while lowest concentration of protein (4.66 mg/g FW) was recorded in Pusa Swani. The combined results of drought stress and okra cultivars for protein showed that okra cultivar Pusa green showed highest protein (9.0 mg/ g FW) at control condition, while lowest protein (2.33 mg/g FW) was observed in Pusa Swani at highest drought stress i.e., 25% FC (Fig. 8). Amin *et al.* (2009) obtained similar results in okra and Kabiri *et al.* (2014) in *Nigella sativa.*

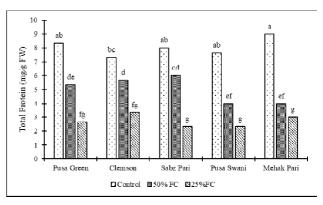


Fig. 8: Effect of drought stress on total protein of okra cultivars

Proline content (µg/g FW)

Maximum proline content $(21.36\mu g/g)$ was noted in those okra plants which were exposed to 25% FC drought whereas minimum proline content $(18.47 \mu g/g)$ was recorded in control plants (100%) FC). Okra cultivars showed significant variations in respect to proline content, okra cultivar Sabz Pari had highest proline content $(27.78\mu g/g)$ while lowest proline value $(11.57 \mu g/g)$ was recorded in Mehak Pari (Table 1). The combined results of drought stress and okra cultivars suggested an increase in proline content of all okra cultivar with increase in drought stress and among all okra cultivar Sabz Pari showed highest proline content of 29.5, 27.63 and 26.23µg/g at 25%, 50% and 100% FC drought respectively, while okra cultivar Mehak Pari showed lowest proline content of 13.1, 11.4 and 10.23µg/g were observed in Mehak Pari at 125%, 50% and 100% FC drought respectively (Fig. 9). Similar observation was recorded by Rokhzadi (2014) in chick pea, Amin et al. (2009) in okra and Idrees et al. (2010) in lemon grass.



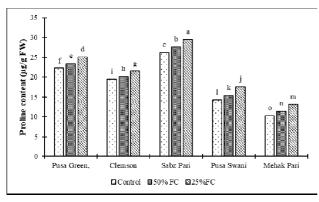


Fig. 9: Effect of drought stress on proline content of okra cultivars

Membrane stability index (%)

Effects of drought on membrane stability index (MSI) indicated that maximum MSI (72.26%) was noted at 25% drought whereas minimum MSI (35.93%) was recorded in control plants. Okra cultivars showed non-significant variations in respect to MSI, okra cultivar Sabz Pari produced highest MSI(56.33%) while lowest MSI values (51.88%) were recorded in Pusa green. The interaction of drought and okra varieties on MSI showed that okra cultivar Mehak Pari produced maximum MSI (73.33%) when grown in 25% FC drought, while least MSI (31.66%) were produced by Pusa green when grown at normal irrigation (Fig. 10). The results of the present study agree with Idress *et al.* (2010) for lemon grass and Sakhabutdinova *et al.* (2006) for wheat.

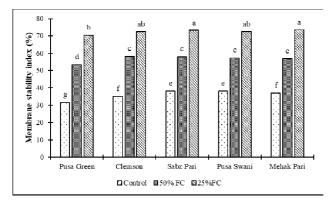


Fig. 10: Effect of drought stress on Membrane Stability index of okra cultivars

DISCUSSION

Reduction in plant biomass and other morphological parameter is clear indication of drought stress on plants and indicates sensitivity towards water deficiency. Our study also confirms that at higher water stress the decrease in plant growth parameters like plant height, fresh weight, and dry weight was observed. Wilting, closing of stomata to prevent transpiration and reduction in cell growth are some key unique responses of plants under drought stress which are produce due to lesser water content, reduced turgor pressure and lower water potential which causes reduced dry and fresh weight and plant height (Guo et al., 2018). Water stress also affects cell division, cell differentiation and growth which might be the reason for a cause for reduced plant height, low dry and fresh weight of okra plants. Lesser availability of water under drought stress to cells might have reduced the photosynthesis due to which plants are unable to acquire desirable biomass and height as suggested by Tanveer et al. (2019).

In case of effects of drought on chlorophyll content of okra it was observed that an increase in drought significantly reduces the chlorophyll pigment concentration and at highest drought the lowest values were observed. Chlorophyll (a, b and total) pigments are responsible for collection and conversion of sunlight into food and energy. Structural and functional integrity of both pigments is directly related to water availability, which is also confirmed by this study that at higher drought a decrease in photosynthetic pigments were observed as suggested by Peiró et al. (2020) and Hussain et al. (2019). Decrease in photosynthetic pigments can be attributed to lesser relative water content of leaves and lower water potential (Trueba et al., 2019). Similarly, stomatal impairment in water deficit plants is responsible for reduced chlorophyll pigment content (Dąbrowski et al., 2019). Drought stress also destabilizes the integrity of protein complexes and increases the activity of chlorophyllase, an enzyme which is responsible for chlorophyll degrading, this eventually leads to reduced chlorophyll concentration. This decrease in chlorophyll content might have caused a reduction in photosynthesis rate, which can lead to lesser availability of food and energy required for development and growth of new tissues and organs, which lead to reduced fresh and dry weight and plant height of okra plants under drought conditions.

Drought stress causes a series of physical and biochemical changes in plant organs and tissues, carotenoids reduction is one of them. This reduction can be related to severity of drought, duration of



exposure and phase of plant growth and genetic resistance capacity of plants towards drought stress (Plazas *et al.*, 2019). Khan *et al.* (2019) also proposed that the reduced carotenoids levels in plants under stress can be due to chloroplast degradation, photo oxidation of chloroplast, chlorophyll synthesis inhibition and increased chlorophyllase activity. Ahmad *et al.* (2019) suggested that activation of LOX and degradation of β -carotene caused the degradation of carotenoids under drought conditions.

Water deficiency under drought condition causes decrease in protein content of plants because water shortage seriously affects the nitrogen metabolism inside plants. The finding of our experiment regarding protein content showed a decrease in protein content with increase in drought stress; this can be attributed to the fact that under drought conditions reduction in polysomal complexes was noted in plant tissues because of lower tissue water content (Khan and Rab 2019). Also, the production of ROS (Reactive Oxygen Species) caused the collapses of protein structure, hence causing an oxidative stress which might be responsible for reduced protein content in stress affected okra plants.

During this study an increase in proline content was recorded in plants subjected to drought. Proline synthesis is greatly associated with the plants response towards stress. Production of osmolytes by plants is a common mechanism adopted to lower the stress; proline is one of these osmolytes which in case of drought stress act as organic reservoir (Zhang *et* *al.*, 2014). It is reported that increase inproline can protect turgor pressure and prevents membrane damage on plants. So, proline accumulation is an adaptation of plants which amplify the tolerance toward drought stress (Singh *et al.*, 2019).

Membrane stability index helps to assess the injury occurred to cell membrane due to a-biotic stress. The integrity of cell membrane allows plant to survive during the continuous or random water deficiencies (Orace and Tehranifar 2020). Decrease of MSI percentage in plants indicates the tolerance of plants towards drought stress (Jafarnia *et al.*, 2018). The increase in MSI of okra plants can be due to production of ROS, and oxidation of cell membrane which caused damage to membrane stability and integrity (Meena *et al.*, 2017).

CONCLUSION

From this experiment it was observed that drought have significant effect on the overall growth and caused serious reduction in biochemical characters of okra cultivars and at higher drought levels maximum reductions in all studied parameters was observed. On the other hand, okra cultivars subjected to different drought levels exhibits prominent variations in all parameters but okra cultivar Sabz Pari showed promising results and showed increased chlorophyll concentration, lower Membrane stability index and higher protein content as compared to other cultivars, Mehak Pari failed to withstand the applied stress and is more sanative to the drought stress.

REFERENCES

- Ahmad, I., Kamran, M., Meng, X., Ali, S., Bilegjargal, B., Cai, T., Liu, T. and Han, Q. 2019. Effects of plant growth regulators on seed filling, endogenous hormone contents and maize production in semiarid regions. J. Plant Growth Regulation, 38(4):1467-1480.
- Altaf, R., Hussain, K., Maryam, U., Nawaz, K. and Siddiqi, E.H. 2015. Effect of different levels of drought on growth, morphology and photosynthetic pigments of lady finger (*Abelmoschus esculentus*). World J. Agri. Sci, 11:198-201.
- Amin, B., Mahleghah, G., Mahmood, H.M.R. and Hossein, M., 2009. Evaluation of interaction

effect of drought stress with ascorbate and salicylic acid on some of physiological and biochemical parameters in okra (*Hibiscus esculentus* L.). *Res. J. Biol. Sci*, **4**:380-387.

- Ayub, Q., Khan, S.M., Hussain, A.K.I., Ahmad, Z. and Khan, M.A., 2018. Effect of gibberellic acid and potassium silicate on physiological growth of Okra (*Abelmoschus esculentus* L.) under salinity stress. *Pure and Applied Biology* (*PAB*), 7(1):8-19.
- Bates LS, Waldeen RP, Teare ID. 1973. Rapid determination of free water stress studies. *Plant Soil*, **39**:205-207



- Dąbrowski, P., Baczewska-Dąbrowska, A.H., Kalaji, H.M., Goltsev, V., Paunov, M., Rapacz, M., Wójcik-Jagła, M., Pawluśkiewicz, B., Bąba, W. and Brestic, M., 2019. Exploration of chlorophyll a fluorescence and plant gas exchange parameters as indicators of drought tolerance in perennial ryegrass. *Sensors*, **19(12)**:2736.
- Dawood, M.G., El-Awadi, M.E.S., Sadak, M.S. and El-Lethy, S.R., 2019. Research Article Comparison Between the Physiological Role of Carrot Root Extract and β-carotene in Inducing Helianthus annuus L. Drought Tolerance *Asian J. Biol. Sci.*, **12** (2):231-241.
- Guo, Y.Y., Yu, H.Y., Yang, M.M., Kong, D.S. and Zhang, Y.J. 2018. Effect of drought stress on lipid peroxidation, osmotic adjustment and antioxidant enzyme activity of leaves and roots of *Lycium ruthenicum* Murr. seedling. *Russian J. Plant Physiol.*, **65**(2), pp.244-250.
- Hussain, H.A., Men, S., Hussain, S., Chen, Y., Ali, S., Zhang, S., Zhang, K., Li, Y., Xu, Q., Liao, C. and Wang, L. 2019. Interactive effects of drought and heat stresses on morphophysiological attributes, yield, nutrient uptake and oxidative status in maize hybrids. *Scientific reports*, 9(1):1-12.
- Idrees, M., Khan, M.M.A., Aftab, T., Naeem, M. and Hashmi, N., 2010. Salicylic acid-induced physiological and biochemical changes in lemongrass varieties under water stress. *Journal of Plant Interactions*, **5**(4): 293-303.
- Jafarnia, S., Akbarinia, M., Hosseinpour, B., ModarresSanavi, S.A.M. and Salami, S.A. 2018. Effect of drought stress on some growth, morphological, physiological, and biochemical parameters of two different populations of Quercusbrantii. *iForest-Biogeosciences and Forestry*, **11**(2):212.
- Jaleel, C.A., Manivannan, P., Wahid, A., Farooq, M.W, Somasundaram, R. and Panneerselvam, R. 2009. Drought stress in plants: a review on morphological characteristics and pigments composition. *Int. J. Agric. Biol.*, 11: 100–105
- Kabiri, R., Nasibi, F. and Farahbakhsh, H. 2014. Effect of exogenous salicylic acid on some

physiological parameters and alleviation of drought stress in *Nigella sativa* plant under hydroponic culture. *Plant Protection Science*, **50**(1):43-51.

- Khan, M.A. and Rab, A. 2019. Plant spacing affects the growth and seed production of okra varieties. *Sarhad Journal of Agriculture*, **35**(3):751-756.
- Khan, M.N., Zhang, J., Luo, T., Liu, J., Ni, F., Rizwan, M., Fahad, S. and Hu, L. 2019. Morpho-physiological and biochemical responses of tolerant and sensitive rapeseed cultivars to drought stress during early seedling growth stage. *Acta Physiologiae Plantarum*, 41(2):25.
- Kiani, S.P., Maury, P., Sarrafi, A. and Grieu, P. 2008.
 QTL analysis of chlorophyll fluorescence parameters in sunflower (*Helianthus annuus* L.) under well-watered and water-stressed conditions. *Plant Sci.*, 175: 565–573
- Kusvuran, S. 2012. Influence of drought stress on growth, ion accumulation and antioxidative enzymes in okra genotypes. *International Journal of Agriculture and Biology*, **14**(3)401-406.
- Lichtenther, H.K. 1987. Chlorophylls and carotenoides: Pigments of photosynthesis. *Methods in Enzymology. INRA, EDP Sci*, 57:245-250.
- Lintunen, A., Paljakka, T., Salmon, Y., Dewar, R., Riikonen, A. and Hölttä, T. 2020. The influence of soil temperature and water content on belowground hydraulic conductance and leaf gas exchange in mature trees of three boreal species. *Plant, Cell & Environment*, **43**(3):532-547.
- Lowry OH, Rusenbrough NJ, Far AL, Randall RJ. 1951. Protein measurement with the Folin's reagent. *J Biol Chem.* **193**:265-266.
- Massacci, A., Nabiev, S.M., Pietrosanti, L., Nematov, S.K., Chernikova, T.N., Thor, K. and Leipner, J. 2008. Response of the photosynthetic apparatus of cotton (*Gossypium hirsutum*) to the onset of drought stress under field conditions studied by gas-exchange analysis and chlorophyll fluorescence imaging. *Plant Physiol. Biochem.*, **46**:189–195



- Meena, K.K., Sorty, A.M., Bitla, U.M., Choudhary, K., Gupta, P., Pareek, A., Singh, D.P., Prabha, R., Sahu, P.K., Gupta, V.K. and Singh, H.B. 2017. Abiotic stress responses and microbe-mediated mitigation in plants: the omics strategies. *Frontiers in Plant Science*, 8:172.
- Meise, P., Seddig, S., Uptmoor, R., Ordon, F. and Schum, A. 2018. Impact of nitrogen supply on leaf water relations and physiological traits in a set of potato (*Solanum tuberosum* L.) cultivars under drought stress. *Journal of Agronomy and Crop Science*, **204**(4), pp.359-374.
- Mounir, S., Ghandour, A., Téllez-Pérez, C., Aly, A.A., Mujumdar, A.S. and Allaf, K. 2020. Phytochemicals, chlorophyll pigments, antioxidant activity, relative expansion ratio, and microstructure of dried okra pods: swelldrying by instant controlled pressure drop versus conventional shade drying. *Drying Technology*, 10.1080/07373937.2020.1756843
- Munir, M., Amjad, M., Ziaf, K. and Ahmad, A. 2016. Improving okra productivity by mitigating drought through foliar application of salicylic acid.*Pakistan Journal of Agricultural Sciences*, 53(4):879-886.
- Nawaz, A., Ali, H., Sufyan, M., Gogi, M.D., Arif, M.J., Ali, A., Qasim, M., Islam, W., Ali, N., Bodla, I. and Zaynab, M. 2020. In-vitro assessment of food consumption, utilization indices and losses promises of leafworm, *Spodoptera litura* (Fab.), on okra crop. *Journal* of Asia-Pacific Entomology, 23(1):60-66.
- Oraee, A. and Tehranifar, A. 2020. Evaluating the potential drought tolerance of pansy through its physiological and biochemical responses to drought and recovery periods. *Scientia Horticulturae*, **265**:109225.
- Peiró, R., Jiménez, C., Perpiñà, G., Soler, J.X. and Gisbert, C. 2020. Evaluation of the genetic diversity and root architecture under osmotic stress of common grapevine rootstocks and clones. *Scientia Horticulturae*, **266**:109283.
- Plazas, M., Nguyen, H.T., González-Orenga, S., Fita, A., Vicente, O., Prohens, J. and Boscaiu, M. 2019. Comparative analysis of the responses to water stress in eggplant (Solanum melongena) cultivars. Plant Physiology and Biochemistry, 143:72-82.

- Premachandra, G.S., Saneoka, H., Kanaya, M. and Ogata, S., 1991. Cell membrane stability and leaf surface wax content as affected by increasing water deficits in maize. *Journal of Experimental Botany*, **42**(2):167-171.
- Qu, X., Wang, H., Chen, M., Liao, J., Yuan, J. and Niu, G., 2019. Drought stress-induced physiological and metabolic changes in leaves of two oil tea cultivars. *Journal of the American Society for Horticultural Science*, 144(6):439-447.
- Ram, A., Verma, P. and Gadi, B.R. 2014. Effect of fluoride and salicylic acid on seedling growth and biochemical parameters of watermelon (*Citrullus lanatus*). *Fluoride*, 47(1):49-55.
- Rokhzadi, A. 2014. Response of chickpea (*Cicer* arietinum L.) to exogenous salicylic acid and ascorbic acid under vegetative and reproductive drought stress conditions. Journal of Applied Botany and Food Quality, **87**:80-86
- Sakhabutdinova, A.R., Fatkhutdinova, D.R., Bezrukova, M.V. and Shakirova, F.M. 2003.
 Salicylic acid prevents the damaging action of stress factors on wheat plants. *Bulg J Plant Physiol*, 21:314-319.
- Singh, A., Kumar, A., Yadav, S. and Singh, I.K. 2019. Reactive oxygen species-mediated signaling during abiotic stress. *Plant Gene*, **18**:100173.
- Singh, B. and Usha, K., 2003. Salicylic acid induced physiological and biochemical changes in wheat seedlings under water stress. *Plant Growth Regulation*, **39**(2):137-141.
- Stanley, L. and Yuan, Y.W., 2019. Transcriptional regulation of carotenoid biosynthesis in plants: So many regulators, so little consensus. *Frontiers in plant science*, 10, p.1017.
- Steel, R.G.D. and Torrie, J.H., 1960. Principles and procedures of statistics. *Principles and procedures of statistics*. McGraw-Hill Book Company, Inc., p.481.
- Tahkokorpi, M., K. Taulavuori, K. Laine and E. Taulavuori, 2007. Aftereffects of droughtrelated winter stress in previous and current year stems of *Vaccinium myrtillus* L. *Environ. Exp. Bot.*, **61**: 85–93



- Tanveer, M., Shahzad, B., Sharma, A. and Khan, E.A. 2019. 24-Epibrassinolide application in plants: An implication for improving drought stress tolerance in plants. *Plant Physiology and Biochemistry*, 135:295-303.
- Trueba, S., Pan, R., Scoffoni, C., John, G.P., Davis, S.D. and Sack, L. 2019. Thresholds for leaf damage due to dehydration: declines of hydraulic function, stomatal conductance and cellular integrity precede those for

photochemistry. New Phytologist, **223**(1):134-149.

- Wilcox, L.V. 1951. A method for calculating the saturation percentage from the weight of a known volume of saturated soil paste. *Soil Science*, **72**(3):233-238
- Zhang, L., Peng, J., Chen, T.T., Zhao, X.H., Zhang, S.P., Liu, S.D., Dong, H.L., Feng, L. and Yu, S.X. 2014. Effect of drought stress on lipid peroxidation and proline content in cotton roots. J. Anim. Plant Sci, 24(6):1729-1736.

(Received on 17.11.2020, Revised on 10.04.2021, Accepted on 24.06.2021)