Phytochemical Analysis and Antioxidant Potential of Ethylacetate Extract of *Tamarindus Indica* (Tamarind) Leaves by Frap Assay

Mubarak Muhammad Dahiru*, Hadiza Ahmadi, Maimuna Umar Faruk, Huzaifa Aminu Hamman, Abreme Gahana Charles

Department of Science laboratory technology, Adamawa State Polytechnic Yola, Nigeria

Abstract

Oxidative stress is characterized by an imbalance in the generation of free radicals and their subsequent elimination by endogenous antioxidants. It is a characteristic of several diseases, especially during the progression stage, which can lead to fatal effects. This study aims to investigate the phytochemical components and antioxidant capability of *Tamarindus indica* and assess its capability as a candidate for managing diseases associated with oxidative The gravimetric method detected and guantified stress. phytochemicals, while the reducing power assay determined the antioxidant potential. Saponins, steroids, and flavonoids were detected in 6.83 ±0.44, 4.30 ±0.60, and 10.17% ±0.60, respectively, without alkaloids, glycosides, and terpenoids. The antioxidant test showed a concentration-dependent increase in absorbance of both the extract and standard (Ascorbic acid). However, Ascorbic acid had higher absorbance. At 100% concentration, the sample had an absorbance of 0.388 ±0.022, which was lower than the absorbance of Ascorbic acid (0.411 ±0.009) at 40% concentration. It can be concluded that Tamarind leaves could be utilized to manage diseases associated with oxidative stress, evidenced by their antioxidant potential credited to the phytochemical content of the leaves. However, there is a need for further studies to ascertain the exact compounds and their modes of action.

Keywords:	antioxidant;	ethylacetate;	oxidative	stress;
phytochemica	l; Tamarindus ir	ndica		

INTRODUCTION

Oxidative stress is a state with a high level of reactive oxygen species (ROS) accompanied by low levels of endogenous antioxidants due to disturbance in the balance for production and neutralization of the ROS by the antioxidants.¹ Several including diseases, diabetes, neurodegenerative, and cancer, are

Data of article

Received : 01 Nov 2022 Reviewed : 22 Dec 2022 Accepted : 30 Jan 2023

DOI

10.18196/jfaps.v2i1.16708

Type of article: Research

associated with oxidative stress, notably during the progression of the diseases where there is continuous damage to DNA, lipids, and proteins.²⁻⁴ In proteins, oxidative damage goes through carbonylation, modification of the side chain, and integrity of the protein molecule. In contrast, in DNA damage, 8hydroxydeoxyguanosine is formed, which binds to thymidine instead of cytosine,

^{*} Corresponding author, e-mail: mubaraq93@gmail.com

causing mutagenesis.^{5,6} Some antioxidants in the form of enzymes found within cells are crucial in maintaining cellular homeostasis, including catalase, superoxide dismutase, and glutathione peroxidases. However, non-enzymatic forms of antioxidants also exist, which include ascorbic acid, glutathione, and vitamin E.7

In type 2 diabetes, a generation of ROS causes the development and progression of diabetes by disrupting the β -cell regulation signaling and pathway, subsequently causing β-cell dysfunction and insulin resistance.⁸ Generation of ROS is a characteristic of cancer cells due to the rapid cellular division. However, strategies are adopted by these cells to stay beneath a threshold for activation of apoptosis, leading to their proliferation. During cancer progression, there is an overexpression of antioxidant enzyme genes and the production of NADPH, thus emphasizing the effects of oxidative stress on cancer progression.² In cardiovascular diseases, ROS is an important part of signaling in the heart, acting as a second messenger. However, oxidative stress sets in when they are in excess, leading to dysfunction, hypertrophy, cardiac failure.9 apoptosis, and heart In neurodegenerative disease, oxidative stress causes target macromolecules and the oxidation of these molecules; proteins, lipids, and nucleic acids, disturbance in proteasome and mitochondrial function, production of cytokines and inflammatory responses, formation of amyloid β deposition, plaque, and advanced glycation end products, and cell death.¹⁰

Plant and their products are applied in managing diseases through several mechanisms, sometimes attributed to their antioxidant potential.¹¹⁻¹³ Phytochemicals from plants are implicated with therapeutic roles of ailments associated with oxidative stress. which is credited to their several pharmacological effects acting individually synergistically.14 or For example, phytochemicals were previously implicated in managing cancer progression by suppressing DNA damage caused by oxidative stress and modulating signaling to pathways leading carcinogenesis due to their antioxidant effects.¹⁵ Considering the background mentioned above, this study aims to investigate the phytochemical components and antioxidant potential of Tamarind (Tamarindus indica) leaves to assess their capability for managing diseases linked to oxidative stress. It considers the effects of oxidative stress in the progression of several diseases, including diabetes, cardiovascular and neurodegenerative diseases, and cancer.

METHOD

Reagent

All the reagents utilized in this study were of AnarlaR.

Plant material

Tamarind leaves were collected from the Yolde Pate area of Yola south Local Government, Adamawa State, Nigeria. It was identified by a Forest Technologist from the Forestry Technology State Department of Adamawa Polytechnic, Yola. A voucher specimen was maintained in the departmental with voucher number herbarium ASP/FT/118. The leaves were air-dried and ground to powder using a blender.

Extraction

The plant sample was extracted by maceration of 300 g of the leave powder in 1L of 70% ethyl acetate for 48 h, followed by filtration and concentration to dryness under reduced pressure yielding 9.2 g extract.¹⁶

Qualitative phytochemical Analysis

Phytochemical tests and identification in ethyl acetate extract of Tamarind leaves (EETL) were carried out to detect alkaloids, saponins, steroids, glycosides, terpenoids, and flavonoids, as previously reported.¹⁶

Quantitative phytochemical Analysis Saponins content

Saponins were quantified by the gravimetric method.¹⁷

Steroids content

Steroids were quantified by the gravimetric method.¹⁸

Flavonoids content

Flavonoids were quantified according to a method described previously.¹⁸

Reducing power assay

The reducing power of EETL was carried out by a previously reported method.19 The plant extract varying concentrations of 20, 40, 60, 80, and 100% were prepared in distilled water. 2.5 ml of 0.2 M phosphate buffer (pH 6.6) and 2.5 mL of 1% potassium ferricyanide were added. It was then incubated at 50 °C for 20 min. After incubation, 2.5 mL of 10% trichloroacetic acid was added, then centrifugated for 10 min at 3000 rpm, and the upper layer was collected. Lastly, 2.5 mL of distilled water was added to 2.5 mL of the upper layer solution, followed by adding 0.5 mL of 0.1% FeCl₃ solution. The absorbance was measured at 700 nm using against blank а UV-Vis а spectrophotometer UV-VIS (752 Spectrometer, Shanghai Yoke Instruments Co., Ltd, China). Ascorbic acid was used as standard.

Statistical Analysis

Data obtained in the study were expressed as mean ± standard error of triplicate determinations' mean (± SEM) evaluated with Statistical Package for the Social Sciences (SPSS) version 22 Software.

RESULTS AND DISCUSSION

Phytochemical Analysis of EETL

The results of the quantitative determination of the phytochemical composition of the ethylacetate extract of Tamarind leaves are presented in Table 1. Saponins, steroids, and flavonoids were detected, while alkaloids, glycosides, and terpenoids were absent.

Table	1:	Phytochemicals	detected	in
EETL				

Phytochemical	Inference	
Alkaloids	Absent	
Saponins	Present	
Steroids	Present	
Glycosides	Absent	
Terpenoids	Absent	
Flavonoids	Present	

The phytochemical reported in this study correlates with a previous report on an ethyl acetate extract of Tamarind leaves. However, in their study, alkaloids were detected as absent.²⁰⁻²² A similar study on the ethanol extract of Tamarind reported absence of saponins, although the alkaloids were detected.23 Besides, in the other study, saponins, steroids, and flavonoids were detected in the ethanol extract of Tamarind leaves at the moment alkaloids were also found.24 In the same study, vitamin E was detected as a good antioxidant. The present study partially agreed with this study for detecting saponins, steroids, and flavonoids. The polarity of ethyl acetate was lower than that of ethanol; thus, the difference in solvent might be the reason for not being able to detect alkaloids.²⁵

The phytochemicals quantified in the ethylacetate extract of Tamarind leaves are shown in Table 2. Flavonoids were quantified in the highest (10.17% ±0.60) concentration, followed by saponins which were present in a concentration of $6.8_{3}\%$ ±0.44. Meanwhile, steroids were quantified in the least concentration (4.30% ±0.60).

Table 2: Quantitation of phytochemical analysis of EETL

Phytochemical	Concentration (%)	
Saponins	6.83 ±0.44	
Steroids	4.30 ±0.60	
Flavonoids	10.17 ±0.60	
	1	

Values are in triplicate determinations ± SEM.

Phytochemicals different exert pharmacological effects, including antioxidant activities through several mechanisms of action. Saponins exert their antioxidant effect by raising the concentration of antioxidant enzymes, decreasing the release of malondialdehyde or lactate dehydrogenase, and restoring Glutathione homeostasis. Thus, they were implicated in repairing oxidative damage in cells and inhibiting cell death.²⁶ Saponins were previously reported to exert an antioxidant effect on Alzheimer's disease (AD) by preventing DNA damage due to formation of the 8hydroxydeoxyguanosine and increasing the expression of endogenous antioxidant enzymes in the brain of mice, making it a potential therapeutic in the therapy of Saponins were also reported to AD.27 possess hepatoprotective effects on alcohol induced-liver injury by reducing the level of ethanol-induced oxidative stress.²⁸ Polymyxin E-induced

nephrotoxicity was previously reported to be decreased by saponins, and a possible mechanism of action was postulated to be inhibiting oxidative stress and cell death through the mitochondrial pathway.²⁹

Furthermore, flavonoids were previously postulated as a crucial metabolite for managing central nervous system disorders by acting gammaas aminobutyric acid (GABA) receptors.³⁰ The flavonoid naringenin exerts an antioxidant effect by neutralizing ROS and promoting the activities of endogenous antioxidant enzymes in diabetes, cardiovascular and neurodegenerative diseases.³¹ In a similar study, flavonoids were reported to exert antioxidant and antiradical effects against ROS and were postulated for application in managing stress.³² oxidative Flavonoids were previously reported to exert an antioxidant effect on streptozotocininduced diabetic rats by direct antiradical effects.33 Hepatoprotective effects of flavonoids against oxidative stress induced by high glucose were previously reported to regulate antioxidant enzymes.34

Reducing the power of EETL

The reducing power of the EETL is presented in Table 3. A concentrationdependent increase in absorbance by the ethyl acetate extract of Tamarind leaves was observed, with the lowest absorbance at 20% concentration (0.235 ±0.008), while the highest was observed at 100% concentration (0.388 ±0.022). However, the absorption of Ascorbic acid was higher the extract, even at 40% than concentration (0.411 ±0.009).

Table 3: Reducing power of EETL			
Concentration	Absorbance		
(%)	Tamarind	Ascorbic	
(70)	leaves	acid	
20	0.235	0.243	
	±0.008	±0.012	
40	0.245	0.411	
	±0.014	±0.009	
60	0.259	0.644	
	±0.008	±0.016	
80	0.298	0.815	
	±0.017	±0.026	
100	0.388	1.131	
	±0.022	±0.018	

49

Values are in triplicate determinations \pm SEM.

Several studies reported different pharmacological effects of Tamarind, which were attributed to the antioxidant potential of the plant. Tamarind leaves revealed a high radical scavenging effect in vitro by DPPH and ABTS. They were suggested to be a natural antioxidant with anti-diabetic properties.³⁵ The antioxidant activity of Tamarind was reported in a similar study and was attributed to the phenolic compounds detected due to an observed correlation between the total phenolic compounds and DPPH radical scavenging inhibition.³⁶ A previous study on the antioxidant potential of Tamarind FRAP leaves using reported а concentration-dependent increase in absorbance, although it was lower compared to Ascorbic acid, which aligns with the result of our study.³⁷ In this study, the absorbance of EETL increased along concentration. However, with the absorbance of the extract was lower than that of ascorbic acid. Besides, several studies reported similar results for FRAP of Tamarind leaves.³⁸⁻⁴²

CONCLUSION

This research investigated the phytochemical components and antioxidant potential of ethylacetate extract of Tamarind leaves for possible application in diseases associated with oxidative stress. Based on the result of this study, Tamarind leaves might be able to be utilized in diseases associated with oxidative stress, evidenced by its established antioxidant potential attributed to the phytochemicals detected in the leave. However, there is a need for further studies to ascertain the exact compounds and their mechanisms of action.

ACKNOWLEDGMENT

The authors would like to thank the Department of science laboratory technology, Adamawa State Polytechnic Yola, for their institutional support.

REFERENCES

- Sies H. Oxidative stress: a concept in redox biology and medicine. Redox biology. 2015;4:180-3. <u>https://doi.org/10.1016/j.redox.2015.0</u> <u>1.002</u>
- Hayes JD, Dinkova-Kostova AT, Tew KD. Oxidative Stress in Cancer. Cancer Cell. 2020 ;38(2):167-97. <u>https://doi.org/10.1016/j.ccell.2020.0</u> <u>6.001</u>
- 3. Salim S. Oxidative Stress and the Central Nervous System. J Pharmacol Exp Ther. 2017;360(1):201. https://doi.org/10.1124/jpet.116.237503
- 4. Bhatti JS, Sehrawat A, Mishra J, Sidhu
 IS, Navik U, Khullar N, et al. Oxidative
 stress in the pathophysiology of type
 2 diabetes and related complications:

Current therapeutics strategies and future perspectives. Free Radical Biol Med. 2022. https://doi.org/10.1016/j.freeradbiom

<u>ed.2022.03.019</u>

- 5. Pisoschi AM, Pop A. The role of antioxidants in the chemistry of oxidative stress: A review. European Journal of Medicinal Chemistry. 2015 ;97:55-74-<u>https://doi.org/10.1016/j.ejmech.2015</u> .04.040
- Tsapralis D, Panayiotides I, Peros G, Liakakos T, Karamitopoulou E. Human epidermal growth factor receptor-2 gene amplification in gastric cancer using tissue microarray technology. World journal of gastroenterology: WJG. 2012;18(2):150.

https://doi.org/10.3748/wjg.v18.i2.150

- Adwas AA, Elsayed A, Azab AE, Quwaydir FA. Oxidative stress and antioxidant mechanisms in human body. J Appl Biotechnol Bioeng. 2019;6(1):43-7. <u>https://doi.org/10.15406/jabb.2019.0</u> 6.00173
- Zhang P, Li T, Wu X, Nice EC, Huang C, Zhang Y. Oxidative stress and diabetes: antioxidative strategies. Frontiers of Medicine. 2020;14(5):583-600. <u>https://doi.org/10.1007/s11684-</u> 019-0729-1
- 9. D'Oria R, Schipani R, Leonardini A, Natalicchio A, Perrini S, Cignarelli A, et al. The Role of Oxidative Stress in Cardiac Disease: From Physiological Response to Injury Factor. Oxidative Medicine and Cellular Longevity.

2020;2020:5732956.

https://doi.org/10.1155/2020/5732956

- 10. Yaribeygi H, Panahi Y, Javadi B, Sahebkar A. The Underlying Role of Oxidative Stress in Neurodegeneration: A Mechanistic Review. CNS & Neurological Disorders - Drug Targets- CNS & Neurological Disorders). 2018;17(3):207-15. https://doi.org/10.2174/187152731766 6180425122557
- 11. Mileo AM, Miccadei S. Polyphenols as Modulator of Oxidative Stress in Cancer Disease: New Therapeutic Strategies. Oxidative Medicine and Cellular Longevity. 2016 ;2016:6475624.

https://doi.org/10.1155/2016/6475624

- Chang X, Zhang T, Zhang W, Zhao Z, Sun J. Natural drugs as a treatment strategy for cardiovascular disease through the regulation of oxidative stress. Oxidative Medicine and Cellular Longevity. 2020;2020. https://doi.org/10.1155/2020/5430407
- Sarrafchi A, Bahmani M, Shirzad H, Rafieian-Kopaei M. Oxidative stress and Parkinson's disease: New hopes in treatment with herbal antioxidants. Curr Pharm Des. 2016 ;22(2):238-46.

https://doi.org/10.2174/138161282266 6151112151653

14. Chandran R, Abrahamse H. Identifying plant-based natural medicine against oxidative stress and neurodegenerative disorders. Oxidative Medicine and Cellular Longevity. 2020;2020. <u>https://doi.org/10.1155/2020/8648742</u> 51 Mubarak Muhammad Dahiru, Hadiza Ahmadi, Maimuna Umar Faruk, Huzaifa Huzaifa Aminu Hamman, Abreme Gahana Charles | Phytochemical Analysis and Antioxidant Potential of Ethylacetate Extract of *Tamarindus Indica* (Tamarind) Leaves by Frap Assay

- 15. Chikara S, Nagaprashantha LD, Singhal J, Horne D, Awasthi S, Singhal SS. Oxidative stress and dietary phytochemicals: Role in cancer chemoprevention and treatment. Cancer Lett. 2018 ;413:122-34. https://doi.org/10.1016/j.canlet.2017. 11.002
- 16. Evans WC. Trease and Evans' pharmacognosy: Elsevier Health Sciences; 2009.
- 17. Obadoni B, Ochuko P. Phytochemical studies and comparative efficacy of the crude extracts of some haemostatic plants in Edo and Delta States of Nigeria. Global Journal of pure and applied sciences. 2002;8(2):203-8.

https://doi.org/10.4314/gjpas.v8i2.16033

- Harborne A. Phytochemical methods a guide to modern techniques of plant analysis: springer science & business media; 1998.
- Oyaizu M. Studies on products of browning reaction antioxidative activities of products of browning reaction prepared from glucosamine. The Japanese journal of nutrition and dietetics. 1986;44(6):307-15. <u>https://doi.org/10.5264/eiyogakuzash</u> i.44.307
- 20. Mehdi MAH, Alarabi FY, Farooqui M, Pradhan V. Phytochemical screening and antiamebic studies of Tamarindus indica of leaves extract. Asian J Pharm Clin Res. 2019;12(2):507-12. https://doi.org/10.22159/ajpcr.2019.v 12i2.29684
- 21. 21.Adeniyi OV, Olaifa FE, Emikpe BO, Ogunbanwo ST. Phytochemical components and antibacterial activity

of Tamarindus indica Linn. extracts against some pathogens. environment. 2017;7:14. https://doi.org/10.9734/BJI/2017/30618

22. Ugoh SC, Jaruma IM. Phytochemical screening and antibacterial activity of the fruit and leaf extract of Tamarindus indica (Linn). Report and Opinion. 2013;5(8):18-27.

- 23. Chigurupati S, Eric WKY, Jahidul IM, Shantini V, Kesavanarayanan KS, Venkata RRM, et al. Screening antimicrobial potential for Malaysian originated Tamarindus indica ethanolic leaves extract. Asian J Pharm Clin Res. 2018;11(3). https://doi.org/10.22159/ajpcr.2018.v 11i3.22614
- 24. Kagoro MPL, Muhammed BN, Auwal AA, Beskeni DR, Gongden JJ. PHYTOCHEMICAL SCREENING AND ANALYSES OF GC-MS А SUBFRACTION OF 70% ETHANOL EXTRACT OF Tamarindus indica (Linn.) Leaf. Journal of Chemical Nigeria. Society of 2022;47(1). https://doi.org/10.46602/jcsn.v47i1.695
- 25. Herawati D, Pudjiastuti P. Effect of Different Solvents On The Phytochemical Compounds of Sargassum sp. From Yogyakarta and East Nusa Tenggara. Journal of Physics: Conference Series. 2021 ;1783(1):012001. https://doi.org/10.1088/1742-

6596/1783/1/012001

Cui Y, Liu B, Sun X, Li Z, Chen Y, Guo Z, et al. Protective effects of alfalfa saponins on oxidative stress-induced apoptotic cells. Food & function.

2020;11(9):8133-40. https://doi.org/10.1039/DoFO01797C

27. Huang J-L, Jing X, Tian X, Qin M-C, Xu
Z-H, Wu D-P, et al. Neuroprotective Properties of Panax notoginseng Saponins via Preventing Oxidative Stress Injury in SAMP8 Mice. Evidence-Based Complementary and Alternative Medicine. 2017 ;2017:8713561.

https://doi.org/10.1155/2017/8713561

- 28. Wang M, Zhang X-J, Liu F, Hu Y, He C, Li P, et al. Saponins isolated from the leaves of Panax notoginseng protect against alcoholic liver injury via inhibiting ethanol-induced oxidative stress and gut-derived endotoxinmediated inflammation. Journal of Functional Foods. 2015 ;19:214-24. https://doi.org/10.1016/j.jff.2015.09.029
- Zhang Y, Chi X, Wang Z, Bi S, Wang Y, Shi F, et al. Protective effects of Panax notoginseng saponins on PME-Induced nephrotoxicity in mice. Biomedicine & Pharmacotherapy. 2019;116:108970.

https://doi.org/10.1016/j.biopha.2019. 108970

- 30. Diniz TC, Silva JC, Lima-Saraiva SRGd, Ribeiro FPRdA, Pacheco AGM, de Freitas RM, et al. The Role of Flavonoids on Oxidative Stress in Epilepsy. Oxidative Medicine and Cellular Longevity. 2015;2015:171756. https://doi.org/10.1155/2015/171756
- 31. Zaidun NH, Thent ZC, Latiff AA. Combating oxidative stress disorders with citrus flavonoid: Naringenin. Life Sci. 2018;208:111-22. https://doi.org/10.1016/j.lfs.2018.07.017

32. Huyut Z, Beydemir Ş, Gülçin İ. Antioxidant and Antiradical Properties of Selected Flavonoids and Phenolic Compounds. Biochemistry Research International. 2017 ;2017:7616791.

https://doi.org/10.1155/2017/7616791

- 33. Al-Numair KS, Chandramohan G, Veeramani C, Alsaif MA. Ameliorative effect of kaempferol, a flavonoid, on oxidative stress in streptozotocininduced diabetic rats. Redox Report. 2015 ;20(5):198-209. https://doi.org/10.1179/1351000214Y. 0000000117
- 34. Cordero-Herrera I, Martín MA, Goya L, Ramos S. Cocoa flavonoids protect hepatic cells against high-glucoseinduced oxidative stress: Relevance of MAPKs. Mol Nutr Food Res. 2015; 59(4):597-609. https://doi.org/10.1001/mpfr.201400400

https://doi.org/10.1002/mnfr.201400492

- 35. Chigurupati S, Yiik EWK, Vijayabalan S, Selvarajan KK, Alhowail A, Nanda SS, et al. Antioxidant and antidiabetic properties of Tamarindus indica leaf ethanolic extract from Malaysia. Southeast Asian Journal of Tropical Medicine and Public Health. 2020; 51(4):559-69.
- 36. Leng LY, binti Nadzri N, bin Shaari AR, Yee KC. Antioxidant capacity and total phenolic content of fresh, ovendried and stir-fried tamarind leaves. Current Research in Nutrition and Food Science Journal. 2017; 5(3):282-7. https://doi.org/10.12944/CRNFSJ.5.3.13
- 37. Amadou IM, Papa MG, Alioune DF, Modou OK, Kady DB, Abdou S, et al. Antioxidative activity of Tamarindus indica L. extract and chemical

Mubarak Muhammad Dahiru, Hadiza Ahmadi, Maimuna Umar Faruk, Huzaifa Huzaifa Aminu Hamman,Abreme Gahana Charles | Phytochemical Analysis and Antioxidant Potential of Ethylacetate Extract of *Tamarindus Indica* (Tamarind) Leaves by Frap Assay

fractions. African Journal of Biochemistry Research. 2017; 11(2):6-11. https://doi.org/10.5897/AJBR2016.0896

38. Nahar L, Nasrin F, Zahan R, Haque A, Haque E, Mosaddik A. Comparative study of antidiabetic activity of Cajanus cajan and Tamarindus indica in alloxan-induced diabetic mice with a reference to in vitro antioxidant activity. Pharmacognosy research. 2014;6(2):180. https://doi.org/10.4103/0974-

8490.129043

- 39. Yeasmen N, Islam MN. Ethanol as a solvent and hot extraction technique preserved the antioxidant properties of tamarind (Tamarindus indica) seed. Journal of Advanced Veterinary and Animal Research. 2015 ;2(3):332-7. https://doi.org/10.5455/javar.2015.b103
- 40. Lim CY, Mat Junit S, Abdulla MA, Abdul Aziz A. In Vivo Biochemical and Gene Expression Analyses of the Antioxidant Activities and Hypocholesterolemic Properties of Tamarindus indica Fruit Pulp Extract. PLOS ONE. 2013;8(7):e70058. https://doi.org/10.1371/journal.pone.0 070058
- 41. Fagbemi KO, Aina DA, Adeoye-Isijola MO, Naidoo KK, Coopoosamy RM, Olajuyigbe OO. Bioactive compounds, antibacterial and antioxidant activities of methanol extract of Tamarindus indica Linn. Scientific Reports. 2022 ;12(1):9432. https://doi.org/10.1038/s41598-022-13716-x
- 42. Vasant RA, Narasimhacharya AVRL. Ameliorative effect of tamarind leaf on fluoride-induced metabolic

alterations. Environ Health Prevent Med. 2012;17(6):484-93. https://doi.org/10.1007/512199-012-0277-7

53