# The role of microRNAs in COVID-19 with a focus on miR-200c

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# ABSTRACT

**Objective:** Epigenetics is a quickly spreading scientific field, and the study of epigenetic regulation in various diseases such as infectious diseases is emerging. The microribonucleic acids (miRNAs) as one of the types of epigenetic processes bind to their target messenger RNAs (mRNAs) and regulate their stability and/ or translation. This study aims to evaluate non-coding RNAs (ncRNAs) with a focus on miR-200c in COVID-19. In this review, we first define the epigenetics and miRNAs, and then the role of miRNAs in diseases focusing on lung diseases is explained. Finally, in this study, we will investigate the role and position of miRNAs with a focus on miR-200c in viral and severe acute respiratory syndrome–related coronavirus (SARS-CoV2) infections.

**Methods:** Systematic search of MEDLINE, PubMed, Web of Science, Embase, and Cochrane Library was conducted for all relative papers from 2000 to 2021 with the limitations of the English language. Finally, we selected 128 articles which fit the best to our objective of study, among which 5 articles focused on the impact of miR-200c.

**Results:** Due to the therapeutic results of various drugs in different races and populations, epigenetic processes, especially miRNAs, are important. The overall results showed that different types of miRNAs can be effective on the process of various lung diseases through different target pathways and genes. It is likely that amplified levels of miR-200c may lead to decreased angiotensin-converting enzyme-2 (ACE2) expression, which in turn may increase the potential of infection, inflammation, and the complications of coronavirus disease.

Conclusion: miR-200c and its correlation with ACE2 can be used as early prognostic and diagnostic markers.

Keywords: Covid-19, Epigenetic, Lung diseases, miR-200c, miRNAs

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# Introduction

Epigenetics is defined as hereditary changes in gene expression without altering the deoxyribonucleic acid (DNA) sequence (1). Methylation of cytosine in the DNA sequence and the biochemical changes of histones are two critical mechanisms in the epigenetics that play an important role in gene regulation, differentiation, and carcinogenicity (2-5). Another mechanism that affects epigenetics and the gene expression is microribonucleic acids (miRNAs). miRNAs are non-coding endogenous RNAs with a length of 20 to 25 nucleotides. These molecules can bind to untranslated 3' regions (UTRs) and suppress the expression of messenger RNAs (mRNAs) at the posttranscriptional level by pairing a specific base sequence (6,7). miRNAs bind to their target mRNAs and regulate their stability and/or translation. If miRNAs bind completely to their target sequence on the

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mRNA, they can lead to degradation; but in case of binding incorrectly, translational suppression of their target genes occurs by a mechanism that has not yet been fully understood (8). Each miRNA is predicted to have multiple gene targets and each mRNA may be regulated by more than one miRNA (9,10). The miRNAs play a vital role in many important biological processes, including cell proliferation (11), growth (12), differentiation (13), apoptosis (14), metabolism (15), aging (16), signal transduction (17), and viral infections (18). It is estimated that about one-third of genes and their pathways are regulated and controlled by miRNAs. Briefly, miRNAs have a remarkable effect on the genomic and epigenetic mechanisms (19,20).

# The role of miRNAs in diseases focusing on lung diseases

The miRNAs involve in the development, progression, prognosis, diagnosis, and evaluation of therapeutic response in human diseases (21). In recent years, altered expression of the miRNAs has been identified in many human cancers (22),

cardiac hypertrophy and failure (23), metabolic disorders (24), immune system-related diseases, and inflammation (9). Also, the miRNAs have been studied in lung homeostasis, functional development, and various pulmonary diseases including asthma, chronic obstructive pulmonary disease (COPD), cystic fibrosis (CF), idiopathic pulmonary fibrosis (IPF), and lung cancer (25) (Tab. I).

In recent years, an increasing amount of research has shown the impact of miRNAs in the progress of pulmonary diseases (43). Our knowledge of the role of miRNAs in lung diseases has developed step by step. The role of miRNAs in the unique pulmonary cells is thought to be essential in understanding the mechanism of lung function and disease pathogenesis (25). More recently, many studies have begun to report the effects of miRNA transfer via extracellular vesicles. In lung diseases, this transfer was indicated to be facilitated via the intercellular communication between many types of cells in the respiratory system including endothelial cells (44), bronchial epithelial cells (45), mesenchymal stem cells, and others (46).

Disease	miRNA	Gene target	Expression in disease	Sample	Measurement type	Ref
Asthma	miR-145	RUNX3	Up	PB	Quantitative PCR	(26,27)
	miR-21	IL-12	Up	Serum	qRT-PCR	(49)
	miR-133a	RhoA	Down	hBSMCs	qRT-PCR	(50)
	mir-19a	TGFβR2	Up	BEC	RT-PCR	(28)
	miR-155	IL-13Ra1	Up	Macrophages—monocytes	RT-PCR	(48)
COPD	miR-15b	SMAD7	Up	Lung	qRT-PCR	(29)
	miR-146a	COX-2	Down	PLF	RT-PCR/Northern Blot	(30)
	miR-24-3p	BIM	Down	Lung	RT-PCR	(31)
	miR-93-5	NFKBIA	Up	PBMCs	High-throughput microarray	(32)
CF	miR-126	TOM1	Down	Lung	RT-PCR	(33)
	miR-145	CFTR	Up	Cell line	qRT-PCR	(34)
	miR-138	SIN3A	Down	Cell culture	Quantitative PCR	(61)
	miR-9	ANO1	Up	Bronchial tissues	RT-PCR	(35)
IPF	let-7d	HMGA2	Down	Lung	Microarrays	(36)
	miR-21	Smad, Smad7	Up	Lung	miRNA array/Northern blotting	(37)
	miR-200c	TGF-β1	Down	HLT	miR Array	(38)
	miR-199a-5p	TGF-β	Up	Serum	TaqMan miRNA assay	(39)
Lung cancer	miR-137	SLC22A18	Down	Lung	Bioinformatics analysis and luciferase reporter assay	(40)
	mirRNA-34a	TGFβR2	Down	Tissues	qRT-PCR and Western blot	(41)
	miR-449a	E2F3	Down	Lung cancer tissue	RT-PCR	(42)
	miR-200	ZEB1	Down	Tissue	RT-PCR	(55)

BEC = human bronchial epithelial cells; CF = cystic fibrosis; CFTR = Cystic Fibrosis Transmembrane Channel; COPD = chronic obstructive pulmonary disease; HLT = human lung tissue; IPF = idiopathic pulmonary fibrosis; miRNA = microribonucleic acid; PB = peripheral blood; PLF = primary lung fibroblast; qRT-PCR = quantitative reverse transcription polymerase chain reaction; PBMC = peripheral blood mononuclear cell; hBSMC = Human Bronchial Smooth Muscle Cells.

TABLE I - Relationship between miRNA types and their target genes in different lung diseases

# The miRNAs in Asthma

Asthma is a chronic inflammatory disease of the lungs that is often associated with clinical features such as airway hyperresponsiveness (AHR), airflow obstruction, excessive mucus secretion, and airway wall structural changes (remodeling) (47). Interleukin (IL)-13 and transcription factor signal transducer-and-activation-of-transcription-6 (STAT6)-operated pathways have been shown to play a significant role in regulating the prominent asthma features, for example, AHR and remodeling. miR-155 has been shown to be upregulated in order to target directly the transcription of the IL-13 receptor a1 (IL13Ra1) in human macrophages, reducing the levels of IL13Ra1 protein and decreasing the levels of activated STAT6, which is vital in regulating the IL-13 signaling pathway (48). Inhibition of miR-21 leads to a decrease in Th2 cytokine levels (IL-4, IL-5, and IL-13), the number of inflammatory airway leukocytes and AHR (49). Downregulation of miR-133a was followed by an increased expression of RhoA and subsequently increased bronchial hyperactivity in a murine model of asthma (50). Elevated expression of the miR-155 has also been indicated in murine models of asthma. Additionally, by using antagomir against miR-145, the mucus secretion, Th2 cytokine production, and eosinophil infiltration in the airways decreased (51).

# The miRNAs in lung cancer

Dysfunction of miRNAs is often identified in malignancies, including lung tumor. Lung cancer is the leading cause of cancer-related mortality worldwide and to date the roles of miRNAs in lung cancer have been specified and reviewed widely along with the other diseases. Histologically, lung cancer can be mostly divided into small cell (SCLC) and non-small cell lung cancer (NSCLC). The latter is more common and is subclassified into squamous, adenocarcinoma, and large-cell carcinoma (52). Recent sequencing studies have exposed a very large number of targets for each single miRNA. By regulating the posttranscriptional gene expression, miRNAs strongly involved in wide-ranging pathways with the main effect are on the progressive and carcinogenesis routes (53,54). Concisely, various miRNAs that are recognized as either oncogenes or tumor suppressors in lung cancer are also involved in the immune system response, for instance, the miR-200 family. The low expression of the miR-200 family members in human early-stage lung adenocarcinomas has been correlated with upregulation of PD-L1 (55) and CD8<sup>+</sup> T-cell immunosuppression and metastasis, which resulted in the reduction of tumor load. This finding greatly supported the role of miR-200 as a tumor suppressor.

# The miRNAs in COPD

COPD is an inflammatory progressive lung disease that is prompted by chronic inflammation exposure of the airways to stimuli including cigarette smoking and other noxious gases. An increasing number of studies have demonstrated that injured cells such as endothelial and epithelial cells participate seriously in the pathogenesis of COPD (56). The exposure of the respiratory epithelial cells to the harmful agents like cigarette smoke leads to the release of proinflammatory and inflammatory cytokines such as IL-1, IL-6, IL-8, and tumor necrosis factor (TNF)- $\alpha$  (57,58).

#### The miRNAs and CF

In the Caucasian community, CF is the most frequent deadly hereditary disease. It is caused by a recessive mutation in the CFTR (Cystic Fibrosis Transmembrane Channel) gene, which codes for a chloride channel (59). miRNAs can target CFTR directly or indirectly for regulating CF. Several miRNAs can complementarily and directly regulate CFTR expression such as miR-145 (via SMAD3 and TGF- $\beta$ ), miR-223 (via CFTR mRNA), miR-9 (via Anoctamin 1), and miR-494 (via Solute Carrier family 12Member 2 (SLC12A2)), alone or together. However, miR-509-3p and miR-494 downregulate CFTR expression (60). Some miRNAs like miR-138 can also repress the biosynthesis intermediary actors, such as the transcription factor SIN3A (SIN3 transcription regulator family member A) and CFTR (61).

#### The miRNAs and infections

Recent advances in molecular mechanisms point to the importance of miRNAs in the lung and respiratory infections. Acute viral respiratory infections (AVRIs) are the most common causes of acute respiratory symptoms (62). Changes in the regulation of miRNA expression in the epithelial cells of human rhinovirus (hRV), influenza (IV), human metapneumovirus, human coronavirus, and respiratory syncytial virus infections are associated with the pathogenesis of acute respiratory diseases (63). For example, the expression of host miRNAs changes in response to IV stimulation. These miRNAs directly or indirectly target viral and host genes to regulate virus replication, stimulate or suppress innate immune responses and cell apoptosis during the viral infection (64,65). IV increases the expression of miR-4276 by upregulating two proteins involved in the apoptotic pathway, Cas9 and Cocx6c (74), and eventually leads to increased virus replication and apoptosis. Furthermore, a number of specific cellular miRNAs in IV-infected cells including miR-323, mir-491, and miR-654 target the protected region of viral PB1 gene to prevent the virus from replicating in MDCK cells (76).

Another mechanism in IV infection is the altered expression of cellular miRNAs and their effect on important signaling pathways associated with the immune system (66). In hRV infections, miRNAs result in antiviral responses by modulating the immune response (miR-128 and miR-155) as well as controlling virus entry into the infected lung cells (miR-23b) (67).

RSV causes viral respiratory disease in infants and young children (68), modulating the expression of host cell miRNAs for antiviral responses and virus replication similar to the miRNAs mentioned above (69,70). For instance, miR-125a regulates nuclear factor kappa B (NF-κB) signaling pathway by suppressing A20 inhibitor protein (CCL5) as an important cytokine in both innate and compatible immune systems (71). Coronaviruses cause a wide range of respiratory infections, from mild upper respiratory tract infections to severe lower respiratory tract infections (72). Table II shows the four major

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Viral disease		miRNA	Gene target	Effects on gene regulation	Pathways	Ref
		– miRNA-4276	- COX6C	– Up	<ul> <li>Inhibits COX6C and caspase-9 and promoting viral replication</li> </ul>	(74)
		– miR-323, miR-491	- PB1	— Up	<ul> <li>Inhibits replication of virus</li> </ul>	(75)
		– let-7c	- M1	— Up	<ul> <li>Reduces virus replication by degrading M1 mRNA</li> </ul>	(76)
Influenza virus		– miR-146a-5	- TRAF6	— Up	<ul> <li>Negatively regulates innate immune and inflammatory responses</li> </ul>	(77)
		– miR-576-3p	- AP1G1	– Down	<ul> <li>Regulates virus entry</li> </ul>	(78)
		– miR-21-3p	- HDAC8	– Down	<ul> <li>Suppresses IAV replication</li> </ul>	(79)
		– miR-132, miR-200c	– MAPK3, IRAK1	– Up	<ul> <li>Regulates antiviral response</li> </ul>	(64)
Rhinoviruses		– miR-128 , miR-155	– SMAD2, EGFR	— Up	<ul> <li>Regulates the immune response against RV-1B and inhibits virus replication</li> </ul>	(80)
		– miR-23b	- VLDLR	– Up	<ul> <li>Prevents viral infection by decreasing the VLDLR</li> </ul>	(81)
		■ let-7f	CCL7, SOCS3	– Up	<ul> <li>Antiviral host response</li> </ul>	(82)
		■ miR-30, let-7i	– IL-13, TLR4, RUNX2	— Up	<ul> <li>Induces miRNAs to involve in the immune response pathways such as NF-kB and type I IFNs</li> </ul>	(83-85)
RSV		■ miR-221	– NGF, TrkA	– Down	<ul> <li>Promotes viral replication</li> </ul>	(86)
		■ miR-125a	– TNFAIP3	– Down	<ul> <li>Inhibits NF-κB signaling pathway and results in reducing macrophage activation</li> </ul>	(87)
	OC43	miR-9	— NF-kB	Up	N protein of virus binds to miR-9 and modulates NF-kB expression	(88)
	SARS	miR-17, miR-574-5p, miR-214	<ul> <li>Virulent proteins, including N, S, M, and E</li> </ul>	Up	<ul> <li>Suppresses viral replication that may aid evasion of immune surveillance until successful infection of other cells</li> </ul>	(89,90)
Coronavirus	MERS	<ul> <li>miR-16-1-3p, miR-26a-1-</li> <li>3p, miR-425-5p, miR-1275,</li> <li>miR-2277-5p, miR-500b-5p,</li> <li>miR627-5p, miR-1257, miR-1275</li> </ul>	– MAP3K9, MYO15B, SPOCK1	Up	<ul> <li>miRNA-mRNA network significantly impacts MERS-CoV replication</li> </ul>	(91)
	MERS	miR628-5p, miR-18a-3p, hsa-miR332-3p	– Viral mRNA	Up	<ul> <li>These miRNAs may downregulate viral gene expression resulting in the inhibition of viral replication</li> </ul>	(92)
	SARS- CoV2	■ miR-146a-5p	IL-6	Down	<ul> <li>Acts as a negative regulator of NF-κB as the transcription factor of the IL-6 gene</li> </ul>	(93)
		■ miR-200c	ACE2	Up	<ul> <li>Overexpression of miR-200c induces downregulation of ACE2 in human cells</li> </ul>	
		■ miR-1202	SARS-CoV2 ORF1a/b	Up	<ul> <li>Targets SARS-CoV2 genome</li> </ul>	(94)
		let-7d-5p	– TMPRSS2	Up	<ul> <li>Expression of let-7d-5p negatively correlates with TMPRSS2 expression 91</li> </ul>	(95)

TABLE II - Relationship between different types of miRNAs and their target genes in well-known viral lung infections

IL = interleukin; MERS = Middle East respiratory syndrome; miRNA = microribonucleic acid; NF-κB = nuclear factor kappa B; RSV = respiratory syncytial virus; SARS-CoV = severe acute respiratory syndrome-related coronavirus; IAV = Influenza A viruses. categories of the pulmonary virus families and some of the most important miRNAs that change the expression of the genes involved in infections with these viruses. Severe acute respiratory syndrome coronaviruses (SARS-CoV) use host cell miRNAs to escape removal by the immune system (89).

In Middle East respiratory syndrome coronavirus (MERS-CoV) infection, cellular miRNAs act as an antiviral therapeutic agent (92). The functional mechanisms of miRNAs in SARS-CoV2 as the causative agent of COVID-19 are diverse. For example, increased miR-200c expression in the disease downregulates the expression of angiotensin-converting enzyme (ACE2) protein that is the receptor essential for the virus entry into the cell (73).

# Association of miR-200c with the genes involved in inflammation (ACE2, IL-6)

miR-200c-3p is a member of the miR-200 family with two clusters miR-200a/b/429 and miR-200c/141. The miR-200c-3p is one of the most important miRNAs of the second cluster. Studies on the miR-200 family have shown that it has a variety of roles in cancer progression, drug resistance, and oxidative stress (96,97). The results of various studies have revealed the crucial role of the miR-200c epithelial-mesenchymal transmission, proliferation, metastasis, apoptosis, autophagy, and therapeutic resistance in several types of cancer (98). The miR-200c is also measured as a biomarker to predict disease progression, diagnosis, and response to therapy in several cancers, both in tissues and in body fluids (blood, urine) (96).

Studies using miRNAs can contribute not only to the understanding of virus-host interactions but also to the stratification of the different severities of COVID-19. In this sense, miR-200c-3p, which has been associated with viral infections, including influenza A, offers itself as a candidate for the study of COVID-19. The analysis of its expression in groups of patients presenting different levels of disease aggressiveness could contribute to a better screening of patients affected by SARS-CoV2. Thus, in Pimenta's study, which aimed to analyze the expression of miR-200c-3p in saliva samples from patients with COVID-19, the results showed that the expression pattern of miR-200c-3p increased with disease severity (99).

Furthermore, the significant impact of miR-200c-3p in acute respiratory distress syndrome (ARDS) was discovered, which proposes it as a potential factor in SARS-COV-2 research and is considered as a potential diagnostic agent for SARS-COV-2 studies (100). In a study of the H5N1 avian influenza virus (AIV) ACE, serum levels of miRNA-200c-3p were found to increase in the virus causing acute pulmonary injury and ARDS. This miRNA binds to the 3'-UTR locus of the ACE2 gene, and inhibits the expression of this protein and thus exacerbates the disease (100-102).

The ACE2 gene was first identified from complementary DNA in the left ventricle of the human heart (102). ACE2 inactivates angiotensin II (Ang II) by cleavage and produces Ang 1-7 (103).

Ang II binds to type 1 and type 2 Ang II receptors with high affinity and is involved in regulating blood pressure, body fluid balance, inflammation, cell proliferation, hypertrophy, and fibrosis (104-106). ACE2 has been shown to neutralize the development of severe ARDS caused by AIV, coronavirus, and sepsis in mice (106). ACE2 has also been reported as a receptor for the SARS-CoV2 virus to enter the pneumocytes (107).

# The role of miR-200c in lung inflammation and lung diseases

MiR-200c, alongside with miR-141, is placed in the intragenic zone of chromosome 12. MiR-200c family has beneficial effects on preventing drug resistance, cancer development, and oxidative stress. It consists of two clusters: (1) miR-200c/141 cluster including miR-141-3p and miR-141-5p, miR-200c-3p, miR-200c-5p on chromosome 12p13.31; (2) miR-200a/ b/429 cluster including miR-200a-3p, miR-200a-5p, miR-200b-3p, miR-200b-5p, and miR-429 on chromosome 1p36.33 (108).

miR-200, like ACE2, is greatly expressed in the epithelial cells of the pneumocytes, mainly in type II alveolar epithelial cells. The expression of miR-200 has a crucial role in the differentiation of type II alveolar epithelial cells in fetal lungs, which are important components of the renin-angiotensin system signaling pathway all over the body. miR-200 displays several important effects in the body such as anti-remodeling, anti-inflammatory, and anti-proliferative through reduction of angiotensin II levels (Fig. 1) (109). Remarkable points in this issue are about controlling COVID-19 patients' mortality rates and disease severity, by upregulating ACE2 levels with using angiotensin receptor blockers or ACE2 blockers (110). miR-200 is the exact and direct target of ACE2 at 3'-UTR of ACE2 mRNA which by binding to its locus results in the depression of ACE2 expression as a receptor responsible for ARDS incidence. Normally, ACE2 catalyzes the conversion of AgII to Ag1-7. Later, Ag1-7 binds to mitochondrial assembly (MAS) receptors resulting in Ag1-7 protective effects including anti-proliferation, anti-necrotic and anti-hypertrophic as well as vasodilation and declining of proinflammatory cytokine secretion. SARS-CoV2 inhibits this pathway and worsens AgII adverse effects on lung tissue during the acute phase of the disease. It was reported that SARS-CoV2 induces the secretion of IL-6, TNF- $\alpha$ , IL-1 $\beta$  (102,111-113). Activation of NF-kB pathway, an important factor in ARDS pathogenesis, is one of the noticeable pathways leading to the upregulation of miR-200c-3p. Increased expression of miR-200c-3p occurred when the ACE2 expression decreased (100) (Fig. 1).

These mechanisms include increased mir-200c expression, inhibition of ACE2 expression, by affecting ACE2 protein outside the cell, and by inhibition of other anti-inflammatory functions, all of which are shown in the figure. (1) increased miR-200c expression that SARs-CoV-2 inhibit ACE2 indirectly by regulating miR-200c and directly inhibiting ACE2 expression, (2) by affecting the ACE2 gene, (3) ACE2 protein outside the cell, and (4) by inhibiting other anti-inflammatory functions, all of which are shown in the figure. In addition, miR-200c can also reduce ace2 expression, thereby reducing ACE2 expression and reducing its function. According to research results, the reduction in disease severity in COVID-19 patients associates with the correlation between low expression of ACE2 and high levels of miR-200c-3p in the lungs and the upper respiratory tract (114,115).



Fig. 1 - MiR-200c and ACE2 mechanism of function in the pathogenesis of COVID-19. SARS-CoV2 induces inflammation and severe ARDS through four mechanisms: (1) virus indirectly leads to ACE2 downregulation by enhancing miR-200c expression. (2) Virus directly inhibits ACE2 gene expression. (3) SARS-CoV2 inhibits binding of ACE2 protein to its receptor on the lung cells. (4) SARS-CoV2 inhibits the anti-inflammatory effects of ACE2. ACE2 = angiotensin-converting enzyme-2; ARDS = acute respiratory distress syndrome; COVID = coronavirus; SARS-CoV = severe acute respiratory syndromerelated coronavirus.

Recent studies about the entrance of SARS-CoV-2 to the host cells imply that some miRNAs can actually control the expression of ACE2 and TMPRSS2, which are potentially of high effect in SARS-COV-2 pathogenesis (116).

Several pathways have been studied about the effect of epigenetics on the regulation of ACE2/TMPRSS2 expression levels in respiratory diseases. The epigenetic repression of miRNA transcription can control their regulatory regions. For instance, Lysine-specific demethylase 5B (JARID1B, encoded by the KDM5B gene) was displayed to suppress the transcription of miR-200 family including miR-141, miR-200a, miR-200b, miR-200c, and miR-429. Hsa-miR-125a/hsa-let-7e miRNAs inhibit the transcription of miR-200 family through stimulating H3K4me3 histone, which demethylases the miRNAs of this family. Therefore, hsa-miR-125a-5p via binding to miR-200 family pursues 3'-UTR of ACE2 mRNA and results in the enhancement of ACE2 gene expression while 3'-UTR of the TMPRSS2 is targeted by hsa-let-7e-5p. Concludingly, JARID1B epigenetic activity doesn't directly regulate the expression of ACE2 and TMPRSS2 (116). Scientists have investigated if promoting H3K4me3 demethylation is caused by repression of the transcription of the let-7e and miR-125a via JARID1B gene (117); for example, the upregulation of JARID1B in lung cancer cell line A549 concluded threefold depression of miR-200a and miR-200c expression, while JARID1B knockdown enhanced 1.5-fold their conserved and stable levels (118).

The experimental data show the presence of controlling network containing miR-125a/let-7e/miR-200 families, ACE2/TMPRSS2 as well as histone demethylase JARID1B, and further point a new way for signaling pathway for ACE2 expression. In one report, the single-cell RNA sequencing data analysis sharply indicated that in the majority of human cells ACE2 and TMPRSS2 are not expressed without JARID1B. So, for better understanding, the viral infection pathogenesis needs to be investigated in the regulatory network related to the expression of JARID1B, ACE2, and TMPRSS2 in human respiratory epithelial cells (116).

According to cellular ontologies research on 24 miRNAs, for evaluating the miRNAs targeting SARS-CoV-2 host cell receptor ACE2, it was revealed that miR-429, miR-200a-3p, miR-210-3p, miR-200b-3p, and miR-200c-3p were highly expressed in the respiratory epithelial cells and miR-200c-3p exists abundantly in the cells including endo-epithelial cell, epithelial cells, respiratory epithelial cells, leukocytes, hematopoietic cells, and myeloid leukocytes. Also, miR-200b

and miR-200c were discovered to be extremely conserved (119).

In clinical trials, miR-200 and its correlation with ACE2 can be used as early prognostic and diagnostic markers. Its location on the upstream of ARDS signaling pathways may reduce the morbidity and mortality rates of COVID-19 via epigenetic procedures, which can be so beneficial for human survival.

# Conclusion

At present, there is no exact treatment for COVID-19. Due to the importance of miRNAs in pulmonary diseases, mainly the infectious viral diseases as well as SARS-COV-2, they can be potential candidates of targeted therapy in SARS-COV-2 in order to reduce the morbidity and mortality rates of this disease as miR-200c and its correlation with ACE2 can be used as early prognostic and diagnostic markers. However, further research must be carried out to reveal the exact effect of miR-200c in the pathogenesis of COVID-19 in order to be used clinically.

# Authors' contributions

HS was responsible for the largest share in writing the article. SA and SG-GA conceptualized and wrote the article and article design. However, SA share has been higher. SA contributed in review and editing of final submitted version. MHKA and RA contributed in Methodology, Data validation and Writing original draft of this article.

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# References

- Egger G, Liang G, Aparicio A, Jones PA. Epigenetics in human disease and prospects for epigenetic therapy. Nature. 2004; 429(6990):457-463. <u>CrossRef PubMed</u>
- 2. Jones PA, Baylin SB. The fundamental role of epigenetic events in cancer. Nat Rev Genet. 2002;3(6):415-428. <u>CrossRef PubMed</u>
- Klose RJ, Bird AP. Genomic DNA methylation: the mark and its mediators. Trends Biochem Sci. 2006;31(2):89-97. <u>CrossRef</u> <u>PubMed</u>
- Baylin SB, Ohm JE. Epigenetic gene silencing in cancer a mechanism for early oncogenic pathway addiction? Nat Rev Cancer. 2006;6(2):107-116. <u>CrossRef PubMed</u>
- 5. Jones PA, Laird PW. Cancer epigenetics comes of age. Nat Genet. 1999;21(2):163-167. <u>CrossRef PubMed</u>
- Ambros V. The functions of animal microRNAs. Nature. 2004;431(7006):350-355. <u>CrossRef PubMed</u>
- 7. Jadideslam G, Ansarin K, Sakhinia E, Alipour S, Pouremamali F, Khabbazi A. The microRNA-326: autoimmune diseases,

diagnostic biomarker, and therapeutic target. J Cell Physiol. 2018;233(12):9209-9222. CrossRef PubMed

- Kolahi S, Farajzadeh M-J, Alipour S, et al. Determination of mir-155 and mir-146a expression rates and its association with expression level of TNF-α and CTLA4 genes in patients with Behcet's disease. Immunol Lett. 2018;204:55-59. <u>CrossRef</u> <u>PubMed</u>
- Shahriar A, Ghaleh-Aziz Shiva G, Ghader B, Farhad J, Hosein A, Parsa H. The dual role of mir-146a in metastasis and disease progression. Biomed Pharmacother. 2020;126:110099. <u>CrossRef PubMed</u>
- 10. Rajewsky N. microRNA target predictions in animals. Nat Genet. 2006;38(6)(suppl):S8-S13. <u>CrossRef PubMed</u>
- Cheng AM, Byrom MW, Shelton J, Ford LP. Antisense inhibition of human miRNAs and indications for an involvement of miRNA in cell growth and apoptosis. Nucleic Acids Res. 2005;33(4):1290-1297. <u>CrossRef PubMed</u>
- 12. Karp X, Ambros V. Developmental biology. Encountering microR-NAs in cell fate signaling. Science. 2005;310(5752):1288-1289. CrossRef PubMed
- Miska EA. How microRNAs control cell division, differentiation and death. Curr Opin Genet Dev. 2005;15(5):563-568. <u>CrossRef PubMed</u>
- 14. Xu P, Guo M, Hay BA. MicroRNAs and the regulation of cell death. Trends Genet. 2004;20(12):617-624. <u>CrossRef PubMed</u>
- Alshalalfa M, Alhajj R. Using context-specific effect of miR-NAs to identify functional associations between miRNAs and gene signatures. BMC Bioinformatics. 2013;14(12)(suppl 12): S1. <u>CrossRef PubMed</u>
- Bartel DP. MicroRNAs: target recognition and regulatory functions. Cell. 2009;136(2):215-233. <u>CrossRef PubMed</u>
- Cui Q, Yu Z, Purisima EO, Wang E. Principles of microRNA regulation of a human cellular signaling network. Mol Syst Biol. 2006;2(1):46. <u>CrossRef PubMed</u>
- Barbu MG, Condrat CE, Thompson DC, et al. MicroRNA involvement in signaling pathways during viral infection. Front Cell Dev Biol. 2020;8:143. <u>CrossRef PubMed</u>
- Yang N, Ekanem NR, Sakyi CA, Ray SD. Hepatocellular carcinoma and microRNA: new perspectives on therapeutics and diagnostics. Adv Drug Deliv Rev. 2015;81:62-74. <u>CrossRef PubMed</u>
- 20. Nana-Sinkam SP, Geraci MW. MicroRNA in lung cancer. J Thorac Oncol. 2006;1(9):929-931. <u>CrossRef PubMed</u>
- Yang N, Coukos G, Zhang L. MicroRNA epigenetic alterations in human cancer: one step forward in diagnosis and treatment. Int J Cancer. 2008;122(5):963-968. <u>CrossRef PubMed</u>
- Blenkiron C, Miska EA. miRNAs in cancer: approaches, aetiology, diagnostics and therapy. Hum Mol Genet. 2007;16(Spec No 1):R106-R113. <u>CrossRef PubMed</u>
- Divakaran V, Mann DL. The emerging role of microRNAs in cardiac remodeling and heart failure. Circ Res. 2008;103(10): 1072-1083. <u>CrossRef PubMed</u>
- 24. Rottiers V, Näär AM. MicroRNAs in metabolism and metabolic disorders. Nat Rev Mol Cell Biol. 2012;13(4):239-250. CrossRef PubMed
- 25. Sessa R, Hata A. Role of microRNAs in lung development and pulmonary diseases. Pulm Circ. 2013;3(2):315-328. CrossRef PubMed
- Qiu Y-Y, Zhang Y-W, Qian X-F, Bian T. miR-371, miR-138, miR-544, miR-145, and miR-214 could modulate Th1/Th2 balance in asthma through the combinatorial regulation of Runx3. Am J Transl Res. 2017;9(7):3184-3199. <u>PubMed</u>
- 27. Yu Y, Wang L, Gu GJCCA. The correlation between Runx3 and bronchial asthma. Clin Chim Acta. 2018;487:75-79. <u>CrossRef</u>
- 28. Haj-Salem I, Fakhfakh R, Bérubé JC, et al. MicroRNA-19a enhances proliferation of bronchial epithelial cells by

20

targeting TGF  $\beta R2$  gene in severe asthma. Allergy. 2015;70(2): 212-219.

- Ezzie ME, Crawford M, Cho J-H, et al. Gene expression networks in COPD: microRNA and mRNA regulation. Thorax. 2012;67(2):122-131. <u>CrossRef</u>
- Sato T, Liu X, Nelson A, et al. Reduced miR-146a increases prostaglandin E2 in chronic obstructive pulmonary disease fibroblasts. Am J Respir Crit Care Med. 2010;182(8):1020-1029.
- 31. Nouws J, Wan F, Finnemore E, et al. MicroRNA miR-24-3p reduces DNA damage responses, apoptosis, and susceptibility to chronic obstructive pulmonary disease. JCI Insight. 2021;6(2). <u>CrossRef</u>
- 32. Dang X, Qu X, Wang W, et al. Bioinformatic analysis of microRNA and mRNA Regulation in peripheral blood mononuclear cells of patients with chronic obstructive pulmonary disease. Respir Res. 2017;18(1):1-13. <u>CrossRef</u>
- Oglesby IK, Bray IM, Chotirmall SH, et al. miR-126 is downregulated in cystic fibrosis airway epithelial cells and regulates TOM1 expression. J Immunol. 2010;184(4):1702-1709.
- 34. Gillen AE, Gosalia N, Leir S-H, Harris A. MicroRNA regulation of expression of the cystic fibrosis transmembrane conductance regulator gene. Biochem J. 2011;438(1):25-32. <u>CrossRef</u>
- 35. Sonneville F, Ruffin M, Coraux C, et al. MicroRNA-9 downregulates the ANO1 chloride channel and contributes to cystic fibrosis lung pathology. Nat Commun. 2017;8(1):1-11.
- Pandit KV, Corcoran D, Yousef H, et al. Inhibition and role of let-7d in idiopathic pulmonary fibrosis. Am J Respir Crit Care Med. 2010;182(2):220-229. <u>CrossRef</u>
- Liu G, Friggeri A, Yang Y, et al. miR-21 mediates fibrogenic activation of pulmonary fibroblasts and lung fibrosis. J Exp Med. 2010;207(8):1589-1597.
- Yang S, Banerjee S, de Freitas A, et al. Participation of miR-200 in pulmonary fibrosis. Am J Pathol. 2012;180(2):484-493. <u>CrossRef</u>
- Yang G, Yang L, Wang W, Wang J, Wang J, Xu Z. Discovery and validation of extracellular/circulating microRNAs during idiopathic pulmonary fibrosis disease progression. Gene. 2015; 562(1):138-144.
- 40. Zhang B, Liu T, Wu T, Wang Z, Rao Z, Gao J. microRNA-137 functions as a tumor suppressor in human non-small cell lung cancer by targeting SLC22A18. Int J Biol Macromol. 2015;74:111-118.
- Ma Z-L, Hou P-P, Li Y-L, et al. MicroRNA-34a inhibits the proliferation and promotes the apoptosis of non-small cell lung cancer H1299 cell line by targeting TGFβR2. Tumour Biol. 2015; 36(4):2481-2490.
- Ren X-S, Yin M-H, Zhang X, et al. Tumor-suppressive microRNA-449a induces growth arrest and senescence by targeting E2F3 in human lung cancer cells. Cancer Lett. 2014;344(2): 195-203. <u>CrossRef</u>
- 43. Chen J, Hu C, Pan P. Extracellular vesicle microRNA transfer in lung diseases. Front Physiol. 2017;8:1028. <u>CrossRef PubMed</u>
- Aliotta JM, Pereira M, Wen S, et al. Exosomes induce and reverse monocrotaline-induced pulmonary hypertension in mice. Cardiovasc Res. 2016;110(3):319-330. <u>CrossRef PubMed</u>
- Fujita Y, Araya J, Ito S, et al. Suppression of autophagy by extracellular vesicles promotes myofibroblast differentiation in COPD pathogenesis. J Extracell Vesicles. 2015;4(1):28388. <u>CrossRef</u> <u>PubMed</u>
- Lee C, Mitsialis SA, Aslam M, et al. Exosomes mediate the cytoprotective action of mesenchymal stromal cells on hypoxiainduced pulmonary hypertension. Circulation. 2012;126(22): 2601-2611. <u>CrossRef PubMed</u>
- 47. Fanta CH. Asthma. N Engl J Med. 2009;360(10):1002-1014. CrossRef PubMed
- Martinez-Nunez RT, Louafi F, Sanchez-Elsner T. The interleukin 13 (IL-13) pathway in human macrophages is

modulated by microRNA-155 via direct targeting of interleukin 13 receptor  $\alpha 1$  (IL13Ralpha1). J Biol Chem. 2011;286(3): 1786-1794. CrossRef PubMed

- ElbehidyRM, YoussefDM, El-ShalAS, etal. MicroRNA-21asanovel biomarker in diagnosis and response to therapy in asthmatic children. Mol Immunol. 2016;71:107-114. <u>CrossRef PubMed</u>
- Chiba Y, Tanabe M, Goto K, Sakai H, Misawa M. Downregulation of miR-133a contributes to up-regulation of Rhoa in bronchial smooth muscle cells. Am J Respir Crit Care Med. 2009;180(8):713-719. <u>CrossRef PubMed</u>
- Collison A, Mattes J, Plank M, Foster PS. Inhibition of house dust mite–induced allergic airways disease by antagonism of microRNA-145 is comparable to glucocorticoid treatment. J Allergy Clin Immunol. 2011;128(1):160-167. e4. <u>CrossRef</u>
- Wu K-L, Tsai Y-M, Lien C-T, Kuo P-L, Hung AJ. The roles of MicroRNAinlung cancer. Int J Mol Sci. 2019;20(7):1611. <u>CrossRef</u> <u>PubMed</u>
- 53. Lin PY, Yu SL, Yang PC. MicroRNA in lung cancer. Br J Cancer. 2010;103(8):1144-1148. CrossRef PubMed
- 54. He L, Thomson JM, Hemann MT, et al. A microRNA polycistron as a potential human oncogene. Nature. 2005;435(7043):828-833.
- Chen L, Gibbons DL, Goswami S, et al. Metastasis is regulated via microRNA-200/ZEB1 axis control of tumour cell PD-L1 expression and intratumoral immunosuppression. Nat Commun. 2014;5(1):5241. <u>CrossRef PubMed</u>
- Osei ET, Florez-Sampedro L, Timens W, Postma DS, Heijink IH, Brandsma C-A. Unravelling the complexity of COPD by microRNAs: it's a small world after all. Eur Respir J. 2015;46(3): 807-818. <u>CrossRef PubMed</u>
- 57. Hobbs BD, Tantisira KG. MicroRNAs in COPD: small molecules with big potential. Eur Respir J. 2019;53:1900515.
- Wang Y, Lyu X, Wu X, Yu L, Hu K. Long non-coding RNA PVT1, a novel biomarker for chronic obstructive pulmonary disease progression surveillance and acute exacerbation prediction potentially through interaction with microRNA-146a. J Clin Lab Anal. 2020;34(8):e23346. <u>CrossRef PubMed</u>
- Riordan JR, Rommens JM, Kerem B, et al. Identification of the cystic fibrosis gene: cloning and characterization of complementary DNA. Science. 1989;245(4922):1066-1073. <u>CrossRef</u> <u>PubMed</u>
- Oglesby IK, Chotirmall SH, McElvaney NG, Greene CM. Regulation of cystic fibrosis transmembrane conductance regulator by microRNA-145, -223, and -494 is altered in ΔF508 cystic fibrosis airway epithelium. J Immunol. 2013;190(7): 3354-3362. <u>CrossRef PubMed</u>
- 61. Ramachandran S, Karp PH, Jiang P, et al. A microRNA network regulates expression and biosynthesis of wild-type and DeltaF508 mutant cystic fibrosis transmembrane conductance regulator. Proc Natl Acad Sci USA. 2012;109(33): 13362-13367. CrossRef PubMed
- Poy MN, Eliasson L, Krutzfeldt J, et al. A pancreatic isletspecific microRNA regulates insulin secretion. Nature. 2004; 432(7014):226-230. <u>CrossRef PubMed</u>
- 63. Moschos SA, Williams AE, Perry MM, Birrell MA, Belvisi MG, Lindsay MA. Expression profiling in vivo demonstrates rapid changes in lung microRNA levels following lipopolysaccharide-induced inflammation but not in the anti-inflammatory action of glucocorticoids. BMC Genomics. 2007;8(1):240. <u>CrossRef PubMed</u>
- Buggele WA, Johnson KE, Horvath CM. Influenza A virus infection of human respiratory cells induces primary microRNA expression. J Biol Chem. 2012;287(37):31027-31040. <u>CrossRef</u> <u>PubMed</u>
- 65. Lam W-Y, Yeung AC-M, Ngai KL-K, et al. Effect of avian influenza A H5N1 infection on the expression of microRNA-141

in human respiratory epithelial cells. BMC Microbiol. 2013;13(1):104. <u>CrossRef PubMed</u>

- Li Y, Li J, Belisle S, Baskin CR, Tumpey TM, Katze MG. Differential microRNA expression and virulence of avian, 1918 reassortant, and reconstructed 1918 influenza A viruses. Virology. 2011;421(2):105-113. <u>CrossRef PubMed</u>
- Baulina NM, Kulakova OG, Favorova OO. MicroRNAs: the role in autoimmune inflammation. Acta Nat (Engl Ed). 2016;8(1):21-33. <u>PubMed</u>
- Birkhaug IM, Inchley CS, Aamodt G, Ånestad G, Nystad W, Nakstad B. Infectious burden of respiratory syncytial virus in relation to time of birth modifies the risk of lower respiratory tract infection in infancy: the Norwegian Mother and Child Cohort. Pediatr Infect Dis J. 2013;32(6):e235-e241. <u>CrossRef PubMed</u>
- Rossi GA, Silvestri M, Colin AA. Respiratory syncytial virus infection of airway cells: role of microRNAs. Pediatr Pulmonol. 2015;50(7):727-732. <u>CrossRef PubMed</u>
- Głobińska A, Pawełczyk M, Kowalski ML. MicroRNAs and the immune response to respiratory virus infections. Expert Rev Clin Immunol. 2014;10(7):963-971. <u>CrossRef PubMed</u>
- 71. Leon-Icaza SA, Zeng M, Rosas-Taraco AG. microRNAs in viral acute respiratory infections: immune regulation, biomarkers, therapy, and vaccines. ExRNA. 2019;1(1):1-7. <u>CrossRef PubMed</u>
- 72. Gralinski LE, Baric RS. Molecular pathology of emerging coronavirus infections. J Pathol. 2015;235(2):185-195. <u>CrossRef PubMed</u>
- Lu D, Chatterjee S, Xiao K, et al. MicroRNAs targeting the SARS-CoV-2 entry receptor ACE2 in cardiomyocytes. J Mol Cell Cardiol. 2020;148:46-49. <u>CrossRef PubMed</u>
- Othumpangat S, Noti JD, Beezhold DH. Lung epithelial cells resist influenza A infection by inducing the expression of cytochrome c oxidase VIc which is modulated by miRNA 4276. Virology. 2014;468-470:256-264. <u>CrossRef PubMed</u>
- Song L, Liu H, Gao S, Jiang W, Huang WJ. Cellular microRNAs inhibit replication of the H1N1 influenza A virus in infected cells. J Virol. 2010;84(17):8849-8860.
- 76. Ma YJ, Yang J, Fan XL, et al. Cellular microRNA let-7c inhibits M1 protein expression of the H1N1 influenza A virus in infected human lung epithelial cells. J Cell Mol Med. 2012;16(10):2539-2546. <u>CrossRef PubMed</u>
- Deng Y, Yan Y, Tan KS, et al. MicroRNA-146a induction during influenza H3N2 virus infection targets and regulates TRAF6 levels in human nasal epithelial cells (hNECs). Exp Cell Res. 2017;352(2):184-192.
- 78. Tambyah PA, Sepramaniam S, Ali JM, et al. microRNAs in circulation are altered in response to influenza A virus infection in humans. PLoS One. 2013;8(10):e76811.
- 79. Xia B, Lu J, Wang R, et al. miR-21-3p regulates influenza A virus replication by targeting histone deacetylase-8. Front Cell Infect Microbiol. 2018;8:175.
- Bondanese VP, Francisco-Garcia A, Bedke N, Davies DE, Sanchez-Elsner T. Identification of host miRNAs that may limit human rhinovirus replication. World J Biol Chem. 2014;5(4):437.
- Ouda R, Onomoto K, Takahasi K, et al. Retinoic acid-inducible gene I-inducible miR-23b inhibits infections by minor group rhinoviruses through down-regulation of the very low density lipoprotein receptor. J Biol Chem. 2011;286(29): 26210-26219. <u>CrossRef</u>
- Bakre A, Mitchell P, Coleman JK, et al. Respiratory syncytial virus modifies microRNAs regulating host genes that affect virus replication. J Gen Virol. 2012;93(Pt 11):2346-2356. <u>CrossRef</u>
- Thornburg NJ, Hayward SL, Crowe Jr. Respiratory syncytial virus regulates human microRNAs by using mechanisms involving beta interferon and NF-κB. mBio. 2012;3(6):e00220-12. <u>CrossRef</u>
- Chen X-M, Splinter PL, O'Hara SP, LaRusso NF. A cellular micro-RNA, let-7i, regulates Toll-like receptor 4 expression

and contributes to cholangiocyte immune responses against Cryptosporidium parvum infection. J Biol Chem. 2007;282(39): 28929-28938.

- Vergoulis T, Vlachos IS, Alexiou P, et al. TarBase 6.0: capturing the exponential growth of miRNA targets with experimental support. Nucleic Acids Res. 2012;40(D1):D222-D229.
- Othumpangat S, Walton C, Piedimonte G. MicroRNA-221 modulates RSV replication in human bronchial epithelium by targeting NGF expression. PLoS One. 2012;7(1):e30030.
- Inchley CS, Sonerud T, Fjærli HO, Nakstad B. Nasal mucosal microRNA expression in children with respiratory syncytial virus infection. BMC Infect Dis. 2015;15(1):1-11.
- Lai FW, Stephenson KB, Mahony J, Lichty BD. Human coronavirus OC43 nucleocapsid protein binds microRNA 9 and potentiates NF-κB activation. J Virol. 2014;88(1):54-65.
- Mallick B, Ghosh Z, Chakrabarti J. MicroRNome analysis unravels the molecular basis of SARS infection in bronchoalveolar stem cells. PLoS One. 2009;4(11):e7837.
- Tahamtan A, Inchley CS, Marzban M, et al. The role of microR-NAs in respiratory viral infection: friend or foe? Rev Med Virol. 2016;26(6):389-407. <u>CrossRef</u>
- Zhang X, Chu H, Wen L, et al. Competing endogenous RNA network profiling reveals novel host dependency factors required for MERS-CoV propagation. Emerg Microbes Infect. 2020;9(1):733-746. <u>CrossRef</u>
- Hasan MM, Akter R, Ullah M, Abedin M, Ullah G, Hossain M. A computational approach for predicting role of human microRNAs in MERS-CoV genome. Adv Bioinformatics. 2014; 2014:967946.
- Sabbatinelli J, Giuliani A, Matacchione G, et al. Decreased serum levels of the inflammaging marker miR-146a are associated with clinical non-response to tocilizumab in COVID-19 patients. Mech Ageing Dev. 2021;193:111413. <u>CrossRef</u>
- Chow JT-S, Salmena LJG. Prediction and analysis of SARS-CoV-2-targeting microRNA in human lung epithelium. Genes (Basel). 2020;11(9):1002. <u>CrossRef</u>
- 95. Mukhopadhyay D, Mussa BM. Identification of novel hypothalamic microRNAs as promising therapeutics for SARS-CoV-2 by regulating ACE2 and TMPRSS2 expression: an in silico analysis. Brain Sci. 2020;10(10):666.
- Mutlu M, Raza U, Saatci Ö. miR-200c: a versatile watchdog in cancer progression, EMT, and drug resistance. J Mol Med (Berl). 2016;94(6):629-644. <u>CrossRef PubMed</u>
- Filios SR, Xu G, Chen J, Hong K, Jing G, Shalev A. MicroRNA-200 is induced by thioredoxin-interacting protein and regulates Zeb1 protein signaling and beta cell apoptosis. J Biol Chem. 2014;289(52):36275-36283. <u>CrossRef PubMed</u>
- Shao X-L, Chen Y, Gao L. MiR-200c suppresses the migration of retinoblastoma cells by reversing epithelial mesenchymal transition. Int J Ophthalmol. 2017;10(8):1195-1202. <u>CrossRef PubMed</u>
- 99. Pimenta R, Viana NI, Dos Santos GA, et al. MiR-200c-3p expression may be associated with worsening of the clinical course of patients with COVID-19. Mol Biol Res Commun. 2021;10(3):141-147. <u>PubMed</u>
- Liu Q, Du J, Yu X, et al. miRNA-200c-3p is crucial in acute respiratory distress syndrome. Cell Discov. 2017;3(1):17021. <u>CrossRef</u> <u>PubMed</u>
- 101. Zou Z, Yan Y, Shu Y, et al. Angiotensin-converting enzyme 2 protects from lethal avian influenza A H5N1 infections. Nat Commun. 2014;5(1):3594. <u>CrossRef PubMed</u>
- 102. Imai Y, Kuba K, Rao S, et al. Angiotensin-converting enzyme 2 protects from severe acute lung failure. Nature. 2005;436 (7047):112-116. <u>CrossRef PubMed</u>
- 103. Vickers C, Hales P, Kaushik V, et al. Hydrolysis of biological peptides by human angiotensin-converting enzyme-related

carboxypeptidase. J Biol Chem. 2002;277(17):14838-14843. CrossRef PubMed

- 104. Michel MC, Brunner HR, Foster C, Huo Y. Angiotensin II type 1 receptor antagonists in animal models of vascular, cardiac, metabolic and renal disease. Pharmacol Ther. 2016;164:1-81. <u>CrossRef PubMed</u>
- Chow BS, Allen TJ. Angiotensin II type 2 receptor (AT2R) in renal and cardiovascular disease. Clin Sci (Lond). 2016;130(15): 1307-1326. <u>CrossRef PubMed</u>
- 106. Patel VB, Zhong J-C, Grant MB, Oudit GY. Role of the ACE2/ angiotensin 1–7 axis of the renin–angiotensin system in heart failure. Circ Res. 2016;118(8):1313-1326. <u>CrossRef PubMed</u>
- 107. Andersen KG, Rambaut A, Lipkin WI, Holmes EC, Garry RF. The proximal origin of SARS-CoV-2. Nat Med. 2020;26(4): 450-452. <u>CrossRef PubMed</u>
- Humphries B, Yang C. The microRNA-200 family: small molecules with novel roles in cancer development, progression and therapy. Oncotarget. 2015;6(9):6472-6498. <u>CrossRef PubMed</u>
- 109. Benlhabib H, Guo W, Pierce BM, Mendelson CR. The miR-200 family and its targets regulate type II cell differentiation in human fetal lung. J Biol Chem. 2015;290(37):22409-22422. CrossRef PubMed
- 110. Fosbøl EL, Butt JH, Østergaard L, et al. Association of angiotensin-converting enzyme inhibitor or angiotensin receptor blocker use with COVID-19 diagnosis and mortality. JAMA. 2020;324(2):168-177. <u>CrossRef PubMed</u>
- 111. Papannarao JB, Schwenke D, Manning PJ, Katare R. Upregulated miR-200c may increase the risk of obese individuals to severe COVID-19. medRxiv. 2021. <u>CrossRef</u>

- 112. Li W, Moore MJ, Vasilieva N, et al. Angiotensin-converting enzyme 2 is a functional receptor for the SARS coronavirus. Nature. 2003;426(6965):450-454. <u>CrossRef PubMed</u>
- 113. Qin C, Zhou L, Hu Z, et al. Dysregulation of immune response in patients with coronavirus 2019 (COVID-19) in Wuhan, China. Clin Infect Dis. 2020;71(15):762-768. CrossRef PubMed
- 114. Yang C, Li Y, Xiao S-Y. Differential expression of ACE2 in the respiratory tracts and its relationship to COVID-19 pathogenesis. EBioMedicine. 2020;60:103004. <u>CrossRef PubMed</u>
- 115. Soltani S, Zandi M. miR-200c-3p upregulation and ACE2 downregulation via bacterial LPS and LTA as nteresting aspects for COVID-19 treatment and immunity. Mol Biol Rep. 2021;48(7):5809-5810. <u>CrossRef</u>
- 116. Nersisyan S, Shkurnikov M, Turchinovich A, Knyazev E, Tonevitsky A. Integrative analysis of miRNA and mRNA sequencing data reveals potential regulatory mechanisms of ACE2 and TMPRSS2. PLoS One. 2020;15(7):e0235987-e. <u>CrossRef</u>
- 117. Mitra D, Das PM, Huynh FC, Jones FE. Jumonji/ARID1 B (JARID1B) protein promotes breast tumor cell cycle progression through epigenetic repression of microRNA let-7e. J Biol Chem. 2011;286(47):40531-40535. CrossRef PubMed
- 118. Enkhbaatar Z, Terashima M, Oktyabri D, et al. KDM5B histone demethylase controls epithelial-mesenchymal transition of cancer cells by regulating the expression of the microRNA-200 family. Cell Cycle. 2013;12(13):2100-2112. CrossRef PubMed
- 119. Bozgeyik I. Therapeutic potential of miRNAs targeting SARS-CoV-2 host cell receptor ACE2. Meta Gene. 2021;27:100831. CrossRef PubMed