REVIEW

Various roles of β -glucan in invertebrates

V Vetvicka¹, P Sima²

¹University of Louisville, Department of Pathology, 511 S. Floyd, Louisville, KY 40202, USA ²Institute of Microbiology, Czech Academy of Sciences, Videnska 1083, 142 20 Prague 4, Czech Republic

Accepted November 15, 2017

Abstract

Glucans have a long history as immunomodulators; their effects confirmed in every species tested-from bees to humans. In invertebrates, glucan binding receptors are involved mainly in starting of the prophenoloxidase system, representing one of the first defense mechanisms that evolved in phylogeny. Our review summarizes the current knowledge of the glucan and lipopolysaccharide-binding proteins in invertebrates and offers a new possibility of using these proteins in human medicine.

Key Words: invertebrates; glucan, receptors; GBP: lipopolysaccharide

Introduction

Various glucans have a long history as immunomodulators. However, for several decades, the focus of investigations was almost exclusively oriented towards vertebrates. Glucans, and most of all their β -1,3 configurations, are structurally complex homopolymers of glucose, isolated from yeast, grain, seaweed, and fungus.

More than 20,000 scientific papers describing the biological activities of glucans exist. Strong immunostimulating effects of β -glucans have been demonstrated in all tested animal species including earthworms (Beschin *et al.*, 1998), bees, (Mazzei *et al.*, 2016) shrimp (Duvic *et al.*, 1992), fish (Anderson 1992), mice, rats (Feletti *et al.*, 1992), guinea pigs (Ferencik *et al.*, 1986), sheep, pigs (Benkova *et al.*, 1992), cattle (Buddle *et al.*, 1988), and humans. Currently, there are at least 80 clinical trials underway in numerous countries. Clearly, glucan belongs the oldest molecules with significant immunomodulating processes, and its activity found throughout the evolutionary scale.

From an evolutionary point of view, three main principles (*i.e.*, recognition, processing, and elimination) are common in all invertebrates, despite the fact that they might be based on varying molecular or biochemical backgrounds. Defense strategies of invertebrates lacking lymphocytes and antibodies are based entirely on innate immunity. The major defensive reactions involve phagocytosis, wound healing, graft rejection, production of various factors (e.g., agglutinins, precipitins, opsonins, clotting factors, lysozyme and many others), and humoral defense (Sima *et al.*, 1990). One of the major defensive mechanism of invertebrates is the use of distinctive molecular patterns, including β -glucans.

Prophenoloxidase system

Invertebrates lack immunoglobulins and their immune system relies on the innate ability initiated by pattern recognition receptors and pattern recognition proteins. These receptors are involved in microbial recognition and initiate protein-ligand interaction. These proteins are able to bind to a variety of microbial cell wall components.

In crustaceans alone, at least 15 different types of pattern recognition receptors are known. In general, arthropods, molluscs, and deuterostomian tunicates recognize the microbial surface determinants that are conserved and ubiquitous among microorganisms but not present in the eukarvotic host. These structures-mainly lipopolysaccharide, peptidoglycan, mannan, β-1,3 glucan, Gram-negative-binding protein, and C-type lectin-are recognized by means of a group of germline encoded receptors, usually termed pattern recognition receptors. Lipopolysaccharide and glucan-binding protein (LGBP) genes involved in the activation of prophenoloxidase (proPO) system were identified in almost every invertebrate species studied. The expression of proPO and LGBP genes is different in individual cell hemocyte types (Yang et al., 2015a). These pattern recognition proteins participate in both humoral and cellular aspects of defense reactions by facilitating pathogenic

Corresponding author Vaclav Vetvicka University of Louisville Department of Pathology 511 S. Floyd, Louisville, KY 40202, USA E-mail: vaclav.vetvicka@louisville.edu

recognition of pathogen-associated molecular patterns (PAMP) and by subsequent triggering of a cascade of reactions such as phagocytosis, production of antibacterial peptides, activation of clotting, and proPO cascade (Lai *et al.*, 2011).

Recognition of PAMP provides an essential step for the activation of the proPO cascade (Amparyup et al., 2008). Using a highly selective recognition process, signaling cascades are activated. These cascades regulate production of defense substances, agglutinins, opsonins, nonagglutinin factors, inducible or constitutive antibiotic peptides, and the components of the proPO complex in the host (Ratcliffe et al., 1979). Specific nonself-recognition mechanisms of the proPO system, as a basic part of immune defense of invertebrates, are involved during a row of hierarchized processes like cell cooperation and communication in the course of phagocytosis, nodule and capsule formation, melanin and sclerotization, hemocyte locomotion, and coagulation of blood (Vetvicka et al., 2004). Although the mechanism of the proPO system has been determined, the precise contribution of βglucan-binding protein (GBP) integration with glucan for the activation remains to be fully elucidated. Some studies show that the GBP can be specifically degraded following the activation of proPO with glucan, suggesting that the variations in the GBP levels after specific challenge are an important regulation mechanisms to immune response (Zhang et al., 2016).

Glucan-binding protein

Invertebrates are using innate immune mechanisms which are well conserved throughout the evolution of immunity. A heterologous group of hemolymph proteins serves as a surveillance mechanism by binding to the surface of invading microbes. One member of this group is GBP, which plays a critical role in triggering the innate immune response by detecting glucan found on the surface of microbes. GBP comprises of proteins that share sequence homology to β-glucanase of bacteria (Juncosa et al., 1994) and of sea urchins (Bachman et al., 1996). The first family of GBP was discovered in the hemolymph of Bombyx mori (Ochiai et al., 1988). Subsequently, proteins binding to the β glucan have been identified in numerous invertebrate species; their activity is usually the proPO stimulating activation cascade. Subsequent purification and identification revealed similar properties-proteins containing carboxylterminal glucanase-like domain without any enzymatic activity. Glucan is bound via less conserved amino-terminal domain. Older review studies of the GBP-protein binding are available (Vetvicka and Sima, 2004). Interestingly, in earthworms, the coelomic cytolytic factor shares strong homology with GBP isolated from numerous invertebrates (Bilej et al., 2000).

Glucan-binding protein isolated from mangrove crab *Episesarma tetragonum* and its interaction with pathogens was evaluated. Molecular recognition showed the specific binding affinity towards β glucan molecule. This bindings triggers the innate immunity inside the host (Sivakamavalli *et al.*, 2014). A molecular cloning of the LGBP isolated from Chinese mitten crab Eriocheir sinensis showed significant homology with the same protein in shrimp. Additional experiments showed that the highest level of expression is in hemocytes, but the protein is also expressed in hepatopancreas, muscles, gills, stomach, and intestines. The expression is upregulated after bacterial infection (Zhao et al., 2009). Subsequent study showed that the recombinant LGBP triggers the whole hemolymph-dependent melanization and stimulates the proPO cascade (Zhang et al., 2016). Additional study of GBP isolated from river crab Paratelhupsa confirmed anti-inflammatory, hydrodromus antioxidant, and antibiofim properties (Iswarya et al., 2017a).

In shrimp, GBP plays the vital role in the recognition mechanisms against PAMP found in the membrane of fungi. Recognition of PAMP leads to the widespread innate immune activation including cellular and humoral components of defense (Sritunyalucksana et al., 2000). Formation of GBPglucan complex induces degranulation of hemocytes and activation of the proPO system. As the gene encoding GBP is abundant in white spot syndrome virus (WSSV)-resistant shrimp, it can be hypothesized that GBP is involved in antiviral response. Purification, characterization, and functional analysis of GBP from green tiger shrimp Penaeus semisulcatus showed it has a bifunctional role in proPO-enhancing activity and agglutinating activity. This GBP not only recognizes PAMP, but also induces intracellular signaling (Sivakamavalli et al., 2013).

LGBP from shrimp Fenneropenaeus chinensis was cloned. Sequence analysis and comparison revealed a high identity of 94 %, 90 %, 87 %, and 72 % with Paneaus monodon β -1,3-glucan binding proteins. Litopenaeus stvlirostris LGBP. Marsupenaeu japonicus β-1,3-glucan bindina proteins, and Homarus gammarus β-1,3-glucan binding proteins, respectively (Liu et al., 2009). In all cases, the transcription of LGBP increased at 6hrs postinfection, suggesting that this gene is not only a constitutive expression gene, but and an inducible acute-phase expression gene, the product of which is necessary to amplify the activity of proPO. Subsequent study named this peptide FcβGBP-HDL and showed that the full-length cDNA of 6713 bp has an open reading frame with two glucanase-like motifs and one arginylglycylaspartic acid motif. The expression was upregulated upon infection, but the level of changes was different in each tissue, suggesting that this protein performs its role differently in different tissues (Lai et al., 2011). Similar homology between individual prawn species between LGBP was found genes from Macrobranchium nipponense and M. rosenbergii (89 identity), M. japonicus (76 %). % and Fenneropaneus chinensis (74 %). Again, expression LGBP mRNA was elevated the of in hepatopancreas (Xiu et al., 2014). Similar data were found for LGBP gene from Indian white shrimp F. inducus (Valli et al., 2012) and giant freshwater prawn M. rosenbergii (Yeh et al., 2009). A recent study not ony cloned and characterized LGBP from Fenneropeaeus merguiensis, but showed its wide

specificity towards both Gram-positive and Gramnegative bacteria and yeast (Chaosomboon *et al.*, 2017). Similar studies performed in *F. chinensis*, however, showed binding activity towards Gramnegative bacteria only and expression of LGBP mRNA in hemocytes only (Du *et al.*, 2007).

In crayfish *Astacus leptodactylus*, there exists three apolipoproteins, all translated as a large precursor. Cleavage at the furin-type side results in high density LGBP (Stieb *et al.*, 2014), which is directly involved in innate immunity of Crustaceans (Schmidt *et al.*, 2010).

A GBP isolated from hemocytes of blue swimmer crab *Portunus pelagicus* was shown to have a multifunctional role in defence reactions including agglutination, proPO-enhancing activity, phagocytosis, and encapsulation. In addition, GBP reaction product exhibited antibacterial and antibiofilm activity against both Gram-positive and Gram-negative bacteria (Anjugam *et al.*, 2016).

Molecular cloning and characterization of GBP from *Plutella xylostella* showed significant similarities with β -glucan recognition proteins of other insects. The transcription levels were found upregulated by microbial challenges in all life stages; tissue distribution was mainly expressed in fat body (Huang *et al.*, 2015).

Rather different effects of GBP were found in experiments using ZnO nanoparticles coated by the crustacean GBP. These particles had significant antibacterial activities and showed cytotoxicity against HepG2 cancer cells (Iswarya *et al.*, 2017b), but the mechanisms remain unclear. Rather similar and more detailed data were found using peptides derived from GBP isolated from Pacific abalone *Haliotis discus hannai* (Nam *et al.*, 2016).

Lectins and opsonins were routinely found in molluscs, but only limited information about GBP is available. The first study found and isolated a GBP from the plasma of marine mussel *Perna viridis*. Further characterization showed a 510 kDa protein with ability to activate the proPO cascade via inherent serine protease activity (Jayaraj *et al.*, 2008).

In scallop Chlamys farreri, the LGBP has significant polymorphism, enhancing the binding activity of lipopolysaccharide and glucan, This protein has a direct association with disease resistance (Siva et al., 2012). In Zhikong scallop Chlamys farreri, mRNA expression of LGPB in hemocytes was strongly upregulated by stimulation of lipopolysaccharide and β-glucan and moderately stimulated by peptidoglycan. A recombinant LGBP showed strong agglutination activity towards Escherichia coli, Bacillus subtilis, and Pochia pastoris (Yang et al., 2010). These results suggest that LGBP plays not only a significant part in the response against Gram-negative bacteria as in other invertebrates (Lee et al., 1996), but also against infection with Gram-positive bacteria. LGBP with vastly diverse specificities might function as a nonclonal effector for animal immune system. An excellent review of LGBP molecules in bivalves was published in 2015 (Allam et al., 2015). One group managed chromosomal localization and molecular marker development of the LGBP gene in the Zhikong scallop Chlamys farreri (Huan et al., 2010).

Molecular characterization and gene expression analysis of LGBP was also successfully achieved in the hard clam, *Meretrix meretrix*. The expression was observed in six different tissues, the highest levels in the gill and digestive gland tissue (Liu *et al.*, 2014).

Direct effects of glucan in invertebrates

In earthworms injected with glucan, an increase in celomic cytolytic factor and lysozyme-like activity was reported. Is seems that this action is caused by direct binding of glucan to hemocytes (Kohlerova *et al.*, 2004; Vetvicka and Sima 2004).

Diet supplementation of sea cucumber Apostichopus japonicus with glucan resulted in strong enrichment of intestinal-dominant classes increased and in proliferation of the and Verrucomicrobiaceae Rhodobacteraceae families. In addition, glucan addition had significant impact on immune response of the intestine via NFκB signaling pathway (Yang et al., 2015b). In scallops, glucan treatment increased the expression of cf/TEP, leading to higher survival of those infected by Vibrio (Xue et al., 2017).

One of the most common targets of glucan are shrimp, as the negative impact of WSSV can be commercially significant. A detailed study revealed how the pattern recognition protein binds to glucan and subsequently activates the proPO system (Amparyup et al., 2012). Besides general immunostimulating activity, various glucans were found to offer protection in Penaeus monodon post larvae against WSSV infection by changing the immune gene expression (Wilson et al., 2015). A study (Bae et al., 2012) of flesh shrimp F. chinensis showed that glucan supplementation in absence of pathogen challenge increased total hemocyte counts. Single administration of glucan prior to the WSSV challenge resulted in strong activation of the proPO cascade and reduced shrimp mortality up to 50 % (Thitamadee et al., 2014). However, the second application of glucan led to a significant increase in mortality, most probably through the combination of WSSV infection and overproduction of reactive oxygen species. These data suggest that with prolonged application of glucan in shrimp aquaculture, caution should be prudent as more is not always better (Wang et al., 2013).

Conclusions

Many of the genes for GBP and LGBP are highly expressed in the digestive glands, particularly in bivalves, suggesting that the filter-feeding life might show biased pattern recognition towards digestive system as a first line of defense (Allam and Raftos, 2015). Clearly, the importance of these binding molecules in invertebrates is extremely high, with some authors even suggesting that the functional diversity of these molecules is comparable to antibodies in vertebrates (Fisher *et al.*, 1991).

However, the current rush to clone and characterize the LGBP in various invertebrates is reminiscent of the golden age of competitive immunology, when the scientists happily evaluated phagocytosis or other immune attributes in one species after another. A plethora of reports on LGBP in invertebrates concludes that this protein plays an important role against infection in crustaceans. Although the detailed characterization of this protein in individual species adds to the mosaic of our knowledge of the immune reaction in invertebrates, the rush to be first to describe it in an additional species is counterproductive to the real needs in this field.

Anticancer activities of either GBP or GBPderived peptides offer a new window for GBP research. Particularly important may be the peptides, as they can be produced in a costeffective manner. If more data confirm these results, it might potentially offer a new way how to obtain anticancer drugs.

In addition to the use of GBP, externally added glucan was also found to significantly improve immune reactions of invertebrates. However, due to the limited available data, our knowledge of possible mechanisms is still lacking.

References

- Allam B, Raftos D. Immune responses to infectious diseases in bivalves. J. Invertebr. Pathol. 131: 121-136, 2015.
- Amparyup P, Kondo H, Hirono I, Aoki T, Tassanakajon A. Molecular cloning, genomic organization and recombinant expression of a crustin-like antimicrobial peptide from black tiger shrimp *Penaeus monodon*. Mol. Immunol. 45: 1085-1093, 2008.
- Amparyup P, Sutthangkul J, Charoensapsri W, Tassanakajon A. Pattern recognition protein binds to lipopolysaccharide and beta-1,3-glucan and activates shrimp prophenoloxidase system. J. Biol. Chem. 287: 10060-10069, 2012.
- Anderson DP. Immunostimulants, adjuvants, and vaccine carriers in fish: Applications to aquaculture. Ann. Rev. Fish Dis. 2: 281-307, 1992.
- Anjugam M, Iswarya A, Vaseeharan B. Multifunctional role of beta-1, 3 glucan binding protein purified from the haemocytes of blue swimmer crab *Portunus pelagicus* and *in vitro* antibacterial activity of its reaction product. Fish Shellfish Immunol. 48: 196-205, 2016.
- Bachman ES, McClay DR. Molecular cloning of the first metazoan beta-1,3 glucanase from eggs of the sea urchin *Strongylocentrotus purpuratus*. Proc. Natl. Acad. Sci. USA 93: 6808-6813, 1996.
- Bae SH, Kim BR, Kang BJ, Tsutsui N, Okutsu T, Shinji J, et al. Molecular cloning of prophenoloxidase and the effects of dietary beta-glucan and rutin on immune response in hemocytes of the fleshy shrimp, *Fenneropenaeus chinensis*. Fish Shellfish Immunol. 33: 597-604, 2012.
- Benkova M, Boroskova Z, Soltys J. [Immunostimulatory effects of certain substances in experimental ascaridiasis in pigs]. Vet. Med. (Praha) 36: 717-724, 1992.
- Beschin A, Bilej M, Hanssens F, Raymakers J, Van Dyck E, Revets H, et al. Identification and cloning of a glucan- and lipopolysaccharidebinding protein from *Eisenia foetida* earthworm

involved in the activation of prophenoloxidase cascade. J. Biol. Chem. 273: 24948-24954, 1998.

- Bilej M, De Baetselier P, Beschin A. Antimicrobial defense of the earthworm. Folia Microbiol. 45: 283, 2000.
- Buddle BM, Pulford HD, Ralston M. Protective effect of glucan against experimentally induced staphylococcal mastitis in ewes. Vet. Microbiol. 16: 67-76, 1988.
- Chaosomboon A, Phupet B, Rattanaporn O, Runsaeng P, Utarabhand P. Lipopolysaccharideand beta-1,3-glucan-binding protein from *Fenneropenaeus merguiensis* functions as a pattern recognition receptor with a broad specificity for diverse pathogens in the defense against microorganisms. Dev. Comp. Immunol. 67: 434-444, 2017.
- Du XJ, Zhao XF, Wang JX. Molecular cloning and characterization of a lipopolysaccharide and beta-1,3-glucan binding protein from fleshy prawn (*Fenneropenaeus chinensis*). Mol. Immunol. 44: 1085-1094, 2007.
- Duvic B, Soderhall K. Purification and partial characterization of a beta-1,3-glucan-bindingprotein membrane receptor from blood cells of the crayfish *Pacifastacus leniusculus*. Eur. J. Biochem. 207: 223-228, 1992.
- Feletti F, De Bernardi di Valserra M, Contos S, Mattaboni P, Germogli R. Chronic toxicity study on a new glucan extracted from *Candida albicans* in rats. Arzneimittelforschung 42: 1363-1367, 1992.
- Ferencik M, Kotulova D, Masler L, Bergendi L, Sandula J, Stefanovic J. Modulatory effect of glucans on the functional and biochemical activities of guinea-pig macrophages. Methods Find. Exp. Clin. Pharmacol. 8: 163-166, 1986.
- Fisher WS, DiNuzzo AR. Agglutination of bacteria and erythrocytes by serum from six species of marine molluscs. J. Invertebr. Pathol. 57: 380-394, 1991.
- Huan P, Zhang X, Li F, Zhang Y, Zhao C, Xiang J. Chromosomal localization and molecular marker development of the lipopolysaccharide and beta-1,3-glucan binding protein gene in the Zhikong scallop *Chlamys farreri* (Jones et Preston) (*Pectinoida, Pectinidae*). Genet. Mol. Biol. 33: 36-43, 2010.
- Huang W, Xu X, Freed S, Zheng Z, Wang S, Ren S, et al. Molecular cloning and characterization of a β -1,3-glucan recognition protein from *Plutella xylostella* (L). New Biotechnol. 32: 290-299, 2015.
- Iswarya A, Anjugam M, Vaseeharan B. Role of purified beta-1,3 glucan binding protein (beta-GBP) from *Paratelphusa hydrodromus* and their anti-inflammatory, antioxidant and antibiofilm properties. Fish Shellfish Immunol. 68: 54-64, 2017a.
- Iswarya A, Vaseeharan B, Anjugam M, Ashokkumar B, Govindarajan M, Alharbi NS, *et al.* Multipurpose efficacy of ZnO nanoparticles coated by the crustacean immune molecule β-1,3-glucan binding protein: Toxicity on HepG2 liver cancer cells and bacterial pathogens. Colloids Surf. B Biointerfaces 158: 257-269, 2017b.

- Jayaraj SS, Thiagarajan R, Arumugam M, Mullainadhan P. Isolation, purification and characterization of beta-1,3-glucan binding protein from the plasma of marine mussel *Perna viridis*. Fish Shellfish Immunol. 24: 715-725, 2008.
- Juncosa M, Pons J, Dot T, Querol E, Planas A. Identification of active site carboxylic residues in *Bacillus licheniformis* 1,3-1,4-beta-D-glucan 4-glucanohydrolase by site-directed mutagenesis. J. Biol. Chem. 269: 14530-14535, 1994.
- Kohlerova P, Beschin A, Silerova M, De Baetselier P, Bilej M. Effect of experimental microbial challenge on the expression of defense molecules in *Eisenia foetida* earthworm. Dev. Comp. Immunol. 28: 701-711, 2004.
- Lai X, Kong J, Wang Q, Wang W, Meng X. Cloning and characterization of a beta-1,3-glucanbinding protein from shrimp *Fenneropenaeus chinensis*. Mol. Biol. Rep. 38: 4527-4535, 2011.
- Lee WJ, Lee JD, Kravchenko VV, Ulevitch RJ, Brey PT. Purification and molecular cloning of an inducible gram-negative bacteria-binding protein from the silkworm, *Bombyx mori.* Proc. Natl. Acad. Sci. USA 93: 7888-7893, 1996.
- Liu F, Li F, Dong B, Wang X, Xiang J. Molecular cloning and characterisation of a pattern recognition protein, lipopolysaccharide and β-1,3-glucan binding protein (LGBP) from Chinese shrimp *Fenneropenaeus chinensis*. Mol. Biol. Rep. 36: 471-477, 2009.
- Liu SX, Qi ZH, Zhang JJ, He CB, Gao XG, Li HJ. Lipopolysaccharide and beta-1,3-glucan binding protein in the hard clam (*Meretrix meretrix*): molecular characterization and expression analysis. Genet. Mol. Res. 13: 4956-4966, 2014.
- Mazzei M, Fronte B, Sagona S, Carrozza ML, Forzan M, Pizzurro F, *et al.* Effect of 1,3-1,6 beta-glucan on natural and experimental deformed wing virus infection in newly emerged honeybees (*Apis mellifera ligustica*). PLOS ONE 11: e0166297, 2016.
- Nam BH, Moon JY, Park EH, Kong HJ, Kim YO, Kim DG, *et al.* Antimicrobial and antitumor activities of novel peptides derived from the lipopolysaccharide- and beta-1,3-glucan binding protein of the Pacific abalone *Haliotis discus hannai.* Mar. Drugs 14, 2016.
- Ochiai M, Ashida M. Purification of a beta-1,3glucan recognition protein in the prophenoloxidase activating system from hemolymph of the silkworm, *Bombyx mori.* J. Biol. Chem. 263: 12056-12062, 1988.
- Ratcliffe NA, Rowley AF. Role of hemocytes in defense against biological agents. In: Gupta AP (ed.), Insect Hemocytes: Development, Forms, Functions and Techniques. Cambridge University Press, Cambridge, pp 331-414, 1979.
- Schmidt O, Soderhall K, Theopold U, Faye I. Role of adhesion in arthropod immune recognition. Annu. Rev. Entomol. 55: 485-504, 2010.
- Sima P, Vetvicka V. Evolution of Immune Reactions, CRC Press, Boca Raton, 1990.

- Siva VS, Yang C, Yang J, Wang L, Wang L, Zhou Z, et al. Association of CfLGBP gene polymorphism with disease susceptibility/resistance of Zhikong scallop (*Chlamys farreri*) to *Listonella anguillarum*. Fish Shellfish Immunol. 32: 1117-1123, 2012.
- Sivakamavalli J, Selvaraj C, Singh SK, Vaseeharan B. Interaction investigations of crustacean β-GBP recognition toward pathogenic microbial cell membrane and stimulate upon prophenoloxidase activation. J. Mol. Recognit. 27: 173-183, 2014.
- Sivakamavalli J, Vaseeharan B. Purification, characterization and functional analysis of a novel beta-1, 3-glucan binding protein from green tiger shrimp *Penaeus semisulcatus*. Fish Shellfish Immunol. 35: 689-696, 2013.
- Sritunyalucksana K, Söderhäll K. The proPO and clotting system in crustaceans. Aquaculture 191: 53-69, 2000.
- Stieb S, Roth Z, Dal Magro C, Fischer S, Butz E, Sagi A, *et al.* One precursor, three apolipoproteins: The relationship between two crustacean lipoproteins, the large discoidal lipoprotein and the high density lipoprotein/βglucan binding protein. Biochim. Biophys. Acta 1841: 1700-1708, 2014.
- Thitamadee S, Srisala J. Taengchaiyaphum S, Sritunyalucksana K. Double-dose beta-glucan treatment in WSSV-challenged shrimp reduces viral replication but causes mortality possibly due to excessive ROS production. Fish Shellfish Immunol. 40: 478-484, 2014.
- Valli JS, Vaseeharan B. cDNA cloning, characterization and expression of lipopolysaccharide and beta-1,3-glucan binding protein (LGBP) gene from the Indian white shrimp *Fenneropenaeus indicus*. Comp. Biochem. Physiol. 163A: 74-81, 2012.
- Vetvicka V, Sima P. β-Glucan in invertebrates. Inv. Surv. J. 1: 60-65, 2004.
- Wang YC, Chang CF, Chen HY. The role of glucans in protection of shrimp against disease. In: Vetvicka V, Novak M (eds), Biology and Chemistry of Beta Glucan, Vol. 2. Bentham Science Publishers, pp 173-194, 2013.
- Wilson W, Lowman D, Antony SP, Puthumana J, Bright Singh IS, Philip R. Immune gene expression profile of *Penaeus monodon* in response to marine yeast glucan application and white spot syndrome virus challenge. Fish Shellfish Immunol. 43: 346-356, 2015.
- Xiu Y, Wu T, Liu P, Huang Y, Ren Q, Gu W, *et al.* Molecular cloning and characterization of the lipopolysaccharide and beta-1,3-glucan binding protein from oriental river prawn, Macrobrachium nipponense. Mol. Biol. Rep. 41: 3935-3944, 2014.
- Xue Z, Wang L, Liu Z, Wang W, Liu C, Song X, *et al.* The fragmentation mechanism and immuneprotective effect of CfTEP in the scallop *Chlamys farreri.* Dev. Comp. Immunol. 76: 220-228, 2017.
- Yang CC, Lu CL, Chen S, Liao WL, Chen SN. Immune gene expression for diverse haemocytes derived from pacific white shrimp,

Litopenaeus vannamei. Fish Shellfish Immunol. 44: 265-271, 2015a.

- Yang G, Xu Z, Tian X, Dong S, Peng M. Intestinal microbiota and immune related genes in sea cucumber (*Apostichopus japonicus*) response to dietary beta-glucan supplementation. Biochem. Biophys. Res. Commun. 458: 98-103, 2015b.
- Yang J, Qiu L, Wang L, Wei X, Zhang H, Zhang Y, et al. CfLGBP, a pattern recognition receptor in *Chlamys farreri* involved in the immune response against various bacteria. Fish Shellfish Immunol. 29: 825-831, 2010.
- Yeh MS, Chang CC, Cheng W. Molecular cloning and characterization of lipopolysaccharide- and beta-1,3-glucan-binding protein from the giant

freshwater prawn *Macrobrachium rosenbergii* and its transcription in relation to foreign material injection and the molt stage. Fish Shellfish Immunol. 27: 701-706, 2009.

- Zhang X, Zhu YT, Li XJ, Wang SC, Li D, Li WW, et al. Lipopolysaccharide and beta-1, 3-glucan binding protein (LGBP) stimulates prophenoloxidase activating system in Chinese mitten crab (*Eriocheir sinensis*). Dev. Comp. Immunol. 61: 70-79, 2016.
- Zhao D, Chen L, Qin C, Zhang H, Wu P, Li E, *et al.* Molecular cloning and characterization of the lipopolysaccharide and β -1,3-glucan binding protein in Chinese mitten crab (*Eriocheir sinensis*). Comp. Biochem. Physiol. 154B: 17-24, 2009.