

CLARIFYING AGENTS AND 3-SULFANYLHEXANOL PRECURSORS IN GRAPE JUICE

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ABSTRACT

We evaluated the impact of a number of clarifying agents on the concentration of S-3-(hexan-1-ol)-L-cysteine (Cys-3SH) and S-3-(hexan-1-ol)-L-glutathione (GSH-3SH). 19 clear grape juices were spiked with a grape skin tannin rich in Cys-3SH and GSH-3SH. Juices were then treated with Na-bentonite, PVPP or charcoal (1 g/L) and cold settled. The concentration of precursors was measured and compared to the corresponding untreated control juices in the devatted samples. Cys-3SH and GSH-3SH were analysed using UHPLC-MS/MS and accuracy was guaranteed with deuterated internal standards. Only charcoal caused a statistically significant depletion of both precursors, quantitatively limited even at the highest dose adopted. Technologically, the clarifiers used in juice affected the thiol precursors in a marginal manner.

Keywords: bentonite, charcoal, grape juice, polyvinylpolypyrrolidone, varietal thiols

1. INTRODUCTION

S-3-(hexan-1-ol)-l-cysteine (Cys-3SH) and S-3-(hexan-1-ol)-l-glutathione (GSH-3SH) are precursors, present in grapes and/or formed in juice (TOMINAGA and DUBOURDIEU, 2000; PEYROT DES GACHONS *et al.*, 2002; SCHNEIDER *et al.*, 2006; FEDRIZZI *et al.*, 2009; ROLAND *et al.*, 2011a), of 3-sulfanylhexanol (3SH), responsible - together with its acetate - for the tropical and grapefruit-like fruity notes produced during fermentation by some yeast strains having lyase activity (RONCORONI *et al.*, 2011; WINTER *et al.*, 2011). The grape variety, as well as the processing conditions of grape, pomace and juice, are very important in saving/producing a high level of precursors (ROLAND *et al.*, 2011a; CERRETI *et al.*, 2015; ROMÁN VILLEGAS *et al.* 2016). Technologically speaking, for example, 3SH precursors increase with longer skin-contact and stronger pressing conditions (MATTIVI *et al.*, 2012) and GSH-3SH in particular increases when oxidative pre-fermentative maceration is adopted (LARCHER *et al.*, 2013a). Nevertheless, the effects of the main clarifying agents on the content of the precursors cited are little known to date. For this reason, the aim of the experiment reported in this paper was to investigate whether certain common clarifiers used in juice can modify the concentration of Cys-3SH and GSH-3SH.

2. MATERIALS AND METHODS

2.1. Juice preparation

Nineteen lots of must (20 L) were produced from sound white and red grapes coming from different varieties and plots in Trentino (Northern Italy), selected in order to include a wide compositional variability. Grape lots (300 Kg) were destemmed, crushed and pressed (3.5 bar; press mod. UP600, Willmes, Lorsch, Germany) on a semi-industrial scale at the Edmund Mach Foundation experimental winery (San Michele all'Adige, Italy). To ensure a high concentration of thiol precursors, only the juice fraction over 65% w/v yield was used in the experiment (MAGGU *et al.*, 2007; ALLEN *et al.*, 2011; ROLLAND *et al.*, 2011b). Moreover, the juices were indirectly enriched randomly with 500-1000 mg/L of grape skin tannin containing 224.2 mg/kg GSH-3SH and 25.5 mg/kg Cys-3SH, quantified according to LARCHER *et al.*, (2013b), and supplemented with a volume of 15-25% of Sauvignon Blanc juice, a variety well-known for its richness in thiol precursors (CAPONE *et al.*, 2010; LARCHER *et al.*, 2013a). After sulfiting (20 mg/L SO₂), all juices were cold settled (< 20 nephelometric turbidity units), well beyond normal winemaking practice, in order to minimise the effect of solids suspended in the turbid juice. Clear juices were then devatted, divided into 4 fractions of 5 L each and supplemented with activated Na bentonite (Pentagel, 1 g/L; Perdomini-IOC S.p.A., S. Martino Buon Albergo, Italy), charcoal (Eno Anticromos, 1 g/L; Dal Cin S.p.A., Concorezzo, Italy) or polyvinylpyrrolidone (PVPP V, 1 g/L; Perdomini-IOC) in comparison with the unspiked fraction respectively. After treatment, all samples were cold settled again for 48h at 4°C.

2.2. Sampling

The settled juice was sampled (25 mL), supplemented with methanol (25 mL, -20°C) and stored at -20°C until analysis. The methanol solution was spiked with *d*₅-GSH-3SH and *d*₃-Cys-3SH as labelled internal standards and filtered through a 0.22 µm filter (Millex-GV, Millipore, Ireland) before analysis.

2.3. Chemical analysis

The juice composition was analysed using a WineScan FT 120 Type 77310 (Foss, Hillerød, Denmark), accurately aligned according to the official methods (OIV 2012).

An UPLC Acquity system coupled with a Xevo TQ MS mass spectrometer (Waters Corporation, Milford, USA) was used for LC-MS/MS quantification of thiol precursors. A 5 µL sample was injected into an Acquity UPLC HSS T3 C18 column (1.8 µm film thickness, 2.1 mm × 100 mm; Waters) set with a flow rate of 0.45 mL/min and a temperature of 40°C. MS isotopic dilution analysis was performed in positive ion mode (capillary voltage, 2.5 kV), using argon (0.20 mL/min) and nitrogen (1,000 L/h) as collision and desolvation gas respectively. Other characteristics of the method are specified in LARCHER *et al.* (2013a).

2.4. Statistical analysis

Anova (main effects: juice, clarifier) and Tukey's HSD test were carried out using STATISTICA v. 8.0 (StatSoft Inc., Tulsa, OK).

3. RESULTS AND CONCLUSIONS

The clarifiers were chosen because they are extensively used in winemaking during prefermentation manipulation of white grape must, due to their depletion features in relation to specific classes of compounds (bentonite vs. proteins; PVPP vs. polyphenols) or to their high but non-selective adsorption capacity (charcoal). To our knowledge, there are no reports that specifically link fining agents and thiol precursor content, while their depletion capacity has been previously reported in relation to free and bound primary aromas (MOÏO *et al.*, 2004) and other odour active compounds in juice (LAMBRI *et al.*, 2010).

The juices were chemically characterised by their base composition (mean ± st. dev.; min - max) for total soluble solids (21.2±2.0 °Brix; 18.6-26.0), pH (3.23±0.11; 3.01-3.45), titratable acidity (6.63±1.23 g/L; 4.70-10.00), tartaric acid (5.76±0.69 g/L; 4.93-7.62), malic acid (3.25±0.76 g/L; 1.93-4.68), potassium (1348±158 mg/L; 1086-1618). These data highlight the considerable compositional variability used to ensure the robustness of the results, since the grape cultivar and ripeness not only affect the precursor content (KOBAYASHI *et al.*, 2010; CERRETI *et al.*, 2015) but also influence either the composition (Pirie and Mullins, 1977; POCOCK *et al.*, 2000) or the haze (MESQUITA *et al.*, 2001) of the most usual target molecules for these clarifiers and hence the clarifying activity.

The ranges obtained for GSH-3SH (min-max: 240 - 564 µg/L) and Cys-3SH (36,5 - 244 µg/L) in the control juices match the literature (PEÑA GALLEGO *et al.*, 2012; LARCHER *et al.*, 2013a). Comparison of the results of the corresponding control juices and treated samples showed that bentonite and PVPP had a limited and not statistically significant effect (Table 1) on the concentration of GSH-3SH and Cys-3SH. On the contrary, charcoal treatment significantly reduced ($p<0.05$) the two thiol precursors, however this reduction was limited, being roughly 20% for GSH-3SH and 10% for Cys-3SH.

The significance of these results is not limited to winemaking, but could also be of interest for the grape juice industry, where there is the possibility of using hybrid varieties, resistant to mold diseases and consequently with lower operating costs. Precursors are also present in their juices (LARCHER *et al.*, 2014), and the release of 3SH through specific commercial enzymes could contribute to overall aroma.

Table 1: Thiol precursors in juice in relation to the clarifying agent used. (Values with the same letter are not statistically different in Tukey's HSD test, $p < 0.05$; n.s. = non significant).

Treatment	GSH-3SH ($\mu\text{g/L}$)			Cys-3SH ($\mu\text{g/L}$)		
	Mean (n=19)	S.D.	sign.	Mean (n=19)	S.D.	sign.
Control	344	73	a	80	53	a
Bentonite (1 g/L)	336	75	a	79	53	a
Charcoal (1 g/L)	276	66	b	73	46	b
PVPP (1 g/L)	344	74	a	80	53	a

In conclusion, of the clarifying agents used in this experiment, only charcoal proved able to significantly reduce 3-sulfanylhexanol precursors in juice. Nevertheless, in the light of the usually lower doses of these products adopted for juice in modern white winemaking, the low conversion ratios of the precursors to the corresponding free thiols (ROLAND *et al.*, 2011a), and the limited percentage changes observed in this experiment, it can be deduced that the clarifying agents used affect the content of thiol precursors in a technologically and sensorially negligible manner, despite the low sensory threshold of the relative derivatives in free and acetate form.

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REFERENCES

- Allen T., Herbst-Johnstone M., Girault M., Butler P., Logan G., Jouanneau S. and Kilmartin P.A. 2011. Influence of grape-harvesting steps on varietal thiol aromas in Sauvignon blanc wines. *Journal of Agricultural and Food Chemistry* 59:10641.
- Cerreti M., Esti M., Benucci I., Liburdi K., de Simone C. and Ferranti P. 2015. Evolution of S-cysteinylated and S-glutathionylated thiol precursors during grape ripening of *Vitis vinifera* L. cvs. Grechetto, Malvasia del Lazio and Sauvignon Blanc. *Australian Journal of Grape and Wine Research* 21:411.
- Capone D.L., Sefton M.A., Hayasaka Y. and Jeffery D.W. 2010. Analysis of precursors to wine odorant 3-mercaptohexan-1-ol using HPLC-MS/MS: resolution and quantitation of diastereomers of 3-S-cysteinylohexan-1-ol and 3-S-glutathionylhexan-1-ol. *Journal of Agricultural and Food Chemistry* 58:1390.
- Fedrizzi B., Pardon K.H., Sefton M.A., Elsey G.M. and Jeffery D.W. 2009. First identification of 4-S-glutathionyl-4-methylpentan-2-one, a potential precursor of 4-mercapto-4-methylpentan-2-one, in Sauvignon Blanc juice. *Journal of Agricultural and Food Chemistry* 57:991.
- Kobayashi H., Takase H., Kaneko K., Tanzawa F., Takata R., Suzuki S. and Konno T. 2010. Analysis of S-3-(hexan-1-ol)-glutathione and S-3-(hexan-1-ol)-L-cysteine in *Vitis vinifera* L. cv. Koshu for aromatic wines. *American Journal of Enology and Viticulture* 61:176.
- Lambri M., Dordoni R., Silva A. and De Faveri D.M. 2010. Effect of bentonite fining on odor-active compounds in two different white wine styles. *American Journal of Enology and Viticulture* 61:225.
- Larcher R., Nicolini G., Tonidandel L., Román Villegas T., Malacarne M. and Fedrizzi B. 2013a. Influence of oxygen availability during skin-contact maceration on the formation of precursors of 3-mercaptohexan-1-ol in Müller-Thurgau and Sauvignon Blanc grapes. *Australian Journal of Grape and Wine Research* 19:342.
- Larcher R., Tonidandel L., Nicolini G. and Fedrizzi B. 2013b. First evidence of the presence of S-cysteinylated and S-glutathionylated precursors in tannins. *Food Chemistry* 141:1196.

- Larcher R., Tonidandel L., Nicolini L., Román Villegas T., Gardiman M. and Flamini R. 2014. Precursori di aromi solforati fruttato-tropicali in uve da ibridi. Book of Abstracts 37th World Congress of Vine and Wine, J.M. Aurand (ed.), 9-14 November, Mendoza (Arg), p. 696.
- Maggi M., Winz R., Kilmartin P.A., Trought M.C. and Nicolau L. 2007. Effect of skin contact and pressure on the composition of Sauvignon Blanc must. *Journal of Agricultural and Food Chemistry* 55:10281.
- Mattivi F., Fedrizzi B., Zenato A., Tiefenthaler P., Tempesta S., Perenzoni D., Cantarella P., Simeoni F. and Vrhovsek U. 2012. Development of reliable analytical tools for evaluating the influence of reductive winemaking on the quality of Lugana wines. *Analytica Chimica Acta* 732:194.
- Mesquita P.R., Piçarra-Pereira M.A., Monteiro S., Loureiro V.B., Teixeira A.R. and Ferreira R.B. 2001. Effect of wine composition on protein stability. *American Journal of Enology and Viticulture* 52:324.
- Moio L., Ugliano M., Gambuti A., Genovese A., and Piombino P. 2004. Influence of clarification treatment on concentrations of selected free varietal aroma compounds and glycoconjugates in Falanghina (*Vitis vinifera* L.) must and wine. *American Journal of Enology and Viticulture* 55:7.
- OIV 2012. Compendium of international methods of analysis of wines and musts. Organisation Internationale de la Vigne et du Vin, Paris, France.
- Peña-Gallego A., Hernández-Orte P., Cacho J. and Ferreira V. 2012. S-Cysteinylated and S-glutathionylated thiol precursors in grapes. A review. *Food Chemistry*, 131:1.
- Peyrot des Gachons C., Tominaga T. and Dubourdiou D. 2002. Sulfur aroma precursor present in S-glutathione conjugate form: identification of S-3-(hexan-1-ol)-glutathione in must from *Vitis vinifera* L. cv. Sauvignon blanc. *Journal of Agricultural and Food Chemistry* 50:4076.
- Pirie A. and M.G. Mullins. 1977. Interrelationships of sugars, anthocyanins, total phenols and dry weight in the skin of grape berries during ripening. *American Journal of Enology and Viticulture* 28:204.
- Pocock K.F., Hayasaka Y., McCarthy M.G. and Waters E.J. 2000. Thaumatin-like Proteins and Chitinases, the Haze-Forming Proteins of Wine, Accumulate during Ripening of Grape (*Vitis vinifera*) Berries and Drought Stress Does Not Affect the Final Levels per Berry at Maturity. *Journal of Agricultural and Food Chemistry*, 48:1637.
- Roland A., Schneider R., Razungles A. and Cavelier F. 2011a. Varietal thiols in wine: discovery, analysis and applications. *Chemical Reviews* 111:7355.
- Roland A., Schneider R., Charrier F., Cavelier F., Rossignol M., and Razungles A. 2011b. Distribution of varietal thiol precursors in the skin and the pulp of Melon B. and Sauvignon Blanc grapes. *Food Chemistry* 125:139.
- Román Villegas T., Tonidandel L., Fedrizzi B., Larcher R. and Nicolini G. 2016. Novel technological strategies to enhance tropical thiol precursors in winemaking by-products. *Food Chemistry* 207:16.
- Roncoroni M., Santiago M., Hooks D.O., Moroney S., Harsch M.J., Lee S.A., Richards K.D., Nicolau L. and Gardner R.C. 2011. The yeast IRC7 gene encodes a β -lyase responsible for production of the varietal thiol 4-mercapto-4-methylpentan-2-one in wine. *Food Microbiology* 28:926.
- Schneider R., Charrier F., Razungles A. and Baumes R. 2006. Evidence for an alternative biogenetic pathway leading to 3-mercaptohexanol and 4-mercapto-4-methylpentan-2-one in wines. *Analytica Chimica Acta* 563:58.
- Tominaga T. and Dubourdiou D. 2000. Identification of cysteinylated aroma precursors of certain volatile thiols in passion fruit juice. *Journal of Agricultural and Food Chemistry* 48:2874.
- Winter G., van der Westhuizen T., Higgins V. J., Curtin C. and Ugliano M. 2011. Contribution of cysteine and glutathione conjugates to the formation of the volatile thiols 3-mercaptohexan-1-ol (3MH) and 3-mercaptohexyl acetate (3MHA) during fermentation by *Saccharomyces cerevisiae*. *Australian Journal of Grape and Wine Research* 17:285.

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