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# CAPILLARY ELECTROPHORETIC ANALYSIS OF EXHALED BREATH CONDENSATE IN THE DIAGNOSIS OF GASTROESOPHAGEAL REFLUX DISEASE

PETR KUBÁŇ \*¹, PAVOL ĎURȹ, JÚLIA LAČNÁ¹, MICHAL GREGUй, FRANTIŠEK FORET¹, JIŘÍ DOLINA², ŠTEFAN KONEČNݲ, MARTINA DOUBKOVÁ³, DAGMAR KINDLOVÁ³, EVA POKOJOVÁ³, AND JANA SKŘIČKOVÁ³

In this work, capillary electrophoresis with contactless conductometric detection (CCD) was used for the analysis of the ionic content of exhaled breath condensate (EBC) to differentiate between healthy individuals and patients with gastroe-sophageal reflux disease (GERD). The exhaled breath condensate was collected using a miniature sample collection device and the content analyzed using a separation electrolyte composed of 20 mM 2-(N-morpholino)ethanesulfonic acid, 20 mM L-histidine, 2 mM 18-Crown-6 and 30  $\mu$ M cetyltrimethylammonium bromide. The separation of anions took less than 2.5 minutes, while the cations were separated in less than 1.5 minutes. The most significantly elevated ions in the group of patients suffering from gastroesophageal reflux disease were chloride, nitrate, propionate and butyrate. Although the number of subjects was too small to draw definite conclusions with regard to the discriminatory power of these ions, the pilot data are promising for EBC as a useful non-invasive alternative for other methods used in the diagnosis of gastroesophageal reflux disease.

**Keywords:** capillary electrophoresis, exhaled breath condensate, ionic analysis, diagnosis, gastroesophageal reflux disease

#### 1. Introduction

Capillary electrophoresis is an attractive separation technique that is particularly suitable for the analysis of biological samples. Non-invasive sampling and the analysis of alternative biofluids are gaining scientific attention as these samples are acquired easily, can be obtained repeatedly without any particular stress to the patient, and often are formed of a simple matrix. One such sample is exhaled breath condensate (EBC).

EBC is obtained by cooling exhaled breath using suitable pieces of cooling equipment and apparatus. EBC is the aqueous part of the exhaled breath that mainly contains condensed water from breath and volatile compounds that are soluble in water, in addition to small respiratory droplets that provide information concerning the condition, inflammation and oxidative/nitrosative stress of the lungs. EBC was first used as a diagnostic sample by Sidorenko in the 1980s [1] and since then numerous articles on EBC have been published, including some re-

\*Correspondence: petr.kuban@gmail.com

cent reviews [2, 3]. EBC is attractive not only in terms of studying the condition of the lungs [4, 5], but possibly in the identification of other applications, for instance, in the assessment of inflammatory conditions of the trachea and esophagus. The latter is of particular importance because gastroesophageal reflux disease (GERD) is one of the most common diseases in the western world. Between 20 and 30% of the general population in Europe and USA will suffer from GERD [6] and about 60% of the adult population will experience some type of GERD during their lifespan. GERD is often diagnosed using invasive instruments such as 24-hour multichannel intraluminal impedance and pH monitoring (MII-pH) [7], during which the patient is required to continuously wear a narrow catheter in the esophagus. This device measures the spread and volume of gases, liquids as well as solids through the esophagus and evaluates the composition of refluxate. The data are eventually used to evaluate whether the person is suffering from the disease based on the number of recorded reflux episodes and the De-Meester score [8] which describes the severity of reflux.

Department of Bioanalytical Instrumentation, CEITEC Masaryk University, Veveří 97, 602 00 Brno, CZECH REPUBLIC

<sup>&</sup>lt;sup>2</sup>Department of Internal Gastroenterology, Faculty Hospital Brno, Faculty of Medicine, Masaryk University, Jihlavská 340/20, 625 00 Brno, CZECH REPUBLIC

<sup>&</sup>lt;sup>3</sup>Department of Pulmonary Diseases and Tuberculosis, Faculty Hospital Brno, Faculty of Medicine, Masaryk University, Jihlavská 340/20, 625 00 Brno, CZECH REPUBLIC

The use of MII-pH is, however, rather unpleasant and up to one third of subjects report discomfort. In this paper, the replacement of the above-mentioned MII-pH method with a significantly less invasive technique for analyzing EBC was attempted. EBC was collected using a specially designed miniature sampler [4, 9, 10] that was previously developed in our laboratory. EBC samples were collected from several healthy volunteers and a group of patients suffering from symptoms attributed to gastroesophageal reflux disease. Capillary electrophoresis was used to analyze the ionic content of EBC samples and differences in selected ion concentrations were identified between the groups.

# 2. Experimental

#### 2.1 Instrumentation

An in-house-built CE instrument was used for all electrophoretic separations at  $\pm$  15 kV using a high voltage power supply unit (Spellman CZE2000R Start Spellman, Pulborough, UK). Two Pt wires (with an outer diameter (OD) of 0.5 mm, 3 cm in length, Advent Research Materials Ltd., Eynsham, UK) were inserted into the electrolyte vials to serve as electrodes. Fused-silica (FS) capillaries (with an inner diameter (ID) of 50 µm, OD of 375 μm, 39 cm in total length, Polymicro Technologies, Phoenix, AZ, USA) were used for the separation. All CE experiments were performed at ambient temperature. A custom-made contactless conductometric detector (CCD) (Version 5.06, ADMET s.r.o., Prague, Czech Republic) operating at a frequency of 1.8432 MHz and a voltage of  $50 V_{\rm p-p}$  was used for the detection of the separated analytes. Data were collected using an Orca 2800 24-bit A/D converter and ECOMAC software ver. 0.254 (ECOM spol s.r.o., Prague, Czech Republic). In all experiments, hydrodynamic injection was applied to the samples, consisting of the elevation of the sample vial to a height of 10 cm for 20 s.

The sampler for the collection of EBC has been developed in our group previously [9]. Briefly, it was constructed from a 2 ml syringe (B. Braun Melsungen AG, Melsungen, Germany) cooled by a 5 cm-long hollow aluminum cylinder with an OD of 2.5 cm and an ID identical to the OD of the syringe (see Fig. 1).

The cylinder was kept in dry ice at -80 °C prior to the collection of EBC and an insulation sleeve was used during its collection. A straw with an OD of 6 mm, and a wall thickness of 0.2 mm (purchased in a local store) was used to exhale the air through the sampler. The end of the syringe was enclosed with a parafilm septum to avoid loss of EBC.

# 2.2 Chemicals

All chemicals were of reagent grade (Sigma-Aldrich, Steinheim, Germany) and deionized (DI) water (Purite,

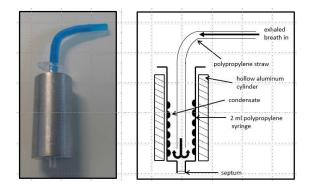


Figure 1: The photograph and scheme of the EBC sampler used in this work.

Neptune, Watrex, Prague, Czech Republic) was used for the preparation of stock solutions and dilutions. Background electrolytes (BGEs) for CE measurements were prepared daily by diluting 100 mM of stock solutions of 2-(N-morpholino)ethanesulfonic acid (MES), L-histidine (L-His) and 18-Crown-6 to the required concentrations. Cetyltrimethylammonium bromide (CTAB) was prepared as a 10 mM stock solution in 5 % acetonitrile.

The separation electrolyte that was used for separation and quantitation of inorganic anions, cations and organic acids consisted of 20 mM MES, 20 mM L-His, 2 mM 18-Crown-6 and 30  $\mu$ M CTAB.

# 2.3 Capillary-conditioning procedure

Prior to its first use, the separation capillary was preconditioned by flushing it with 0.1 M NaOH for 30 mins., then with DI water for 30 mins. and finally with a solution of BGEs for 10 mins. Between two successive injections, the capillary was flushed with the BGE solution for 1 min. At the end of a working day, the capillaries were washed with DI water for 15 mins., followed by the application of a vacuum for 5 mins. before being stored dry overnight.

### 2.4 Sample-collection procedure

The EBC was collected in the morning, the subjects did not eat or drink beforehand. The EBC was collected using the previously described EBC-sampler. The subject was asked to exhale deeply 3-5 times into the sampler, but depending on the lung capacity of each, the number of exhalations was modified to collect approximately  $100~\mu l$  of their EBC. After the sampling of EBC was completed, the condensate from the walls of the syringe was forced to the tip of the syringe by the plunger and transferred to the sample vial for CE analysis. Part of the EBC sample was also used in another experiment published elsewhere, in which the pH was measured using a microelectrode.

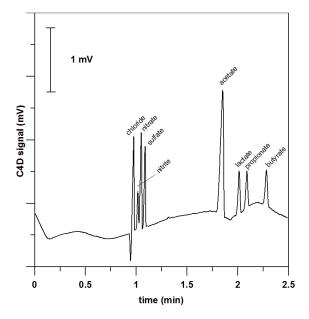


Figure 2: Separation of a model mixture consisting of 4 inorganic anions and 4 organic acids. BGE: 20 mM MES/L-His, 2 mM 18-Crown-6, 30  $\mu$ M CTAB. HV: positive +15 kV, C4D detection. Ion concentrations in the parentheses in  $\mu$ M: acetate (200); chloride, nitrate, lactate, propionate, butyrate (25); nitrite, sulfate (10).

# 2.5 Selection of patients and healthy individuals

In this initial screening, a group of healthy individuals and a group of patients diagnosed with asthma and pulmonary fibrosis were selected. The healthy individuals had no history of GERD or heartburn and were free from other related symptoms. The healthy individuals selected were from approximately the same age group as the patients, i.e., the average age of healthy subjects was 48 and that of patients was 56. The patients suffered from asthma and pulmonary fibrosis and were from the Department of Pulmonary Diseases and Tuberculosis at the Faculty Hospital Brno.

# 3. Results and Analysis

# 3.1 Electrolyte selection

To separate the ionic content of the EBC, the BGEs composed of MES and L-His was chosen as this electrolyte allows for the analysis of anions, organic acids, and cations with good degrees of sensitivity. MES and L-His fulfill the criteria for suitable coions and counterions in CE separation using contactless conductivity detection (C4D). The conductance of the respective BGE components is low, thus analytes including small organic acids (acetate, lactate, butyrate) will be detected as positive peaks. The addition of 18-Crown-6 to the separation electrolyte improves the resolution of  $K^+$  and  $NH_4^+$  cations

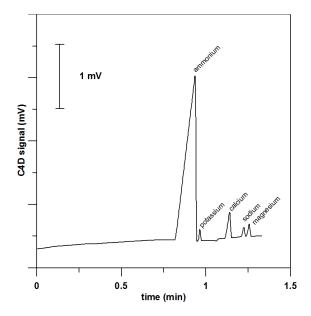


Figure 3: Separation of a model mixture of 5 inorganic cations. BGE: 20 mM MES/L-His, 2 mM 18-Crown-6, 30  $\mu$ M CTAB. HV: negative -15 kV, C4D detection. Ion concentrations in the parentheses in  $\mu$ M: ammonium (1000); calcium (25); potassium, sodium and magnesium (10).

and does not influence the selectivity of other measured cations to a great extent, nor does it influence the separation of anions and organic acids. CTAB is added to the separation electrolyte to decrease the electroosmotic flow (EOF).

It should be noted here that although this separation electrolyte allows for the simultaneous separation of anions and cations using dual-opposite end injection [11], in this work anions and cations were determined separately by switching the high-voltage polarity. The separation of a model mixture consisting of 8 anions in less than 2.5 mins. is shown in Fig. 2.

The ion concentrations were selected to be similar to those found in EBC samples. The separation of cations using the same conditions but using reverse-polarity voltage is shown in Fig. 3. The ammonium cation is usually present at a concentration 100 times greater than other cations in the EBC, its concentration was thus also increased to 1000  $\mu M$  to reflect the expected concentration. Note that even with this significant excess the separation of  $NH_4^+$  from  $K^+$  is sufficient and the BGE composition can be used for real sample analysis.

#### 3.2 Analysis of anions in EBC samples

From each subject, the EBC sample was collected as described previously. About 50  $\mu$ l of the sample was transferred into an Eppendorf tube and hydrodynamically injected into the CE system. The EBC samples were analyzed first for anions and organic acids using positive HV polarity and then for cations using negative HV polarity,

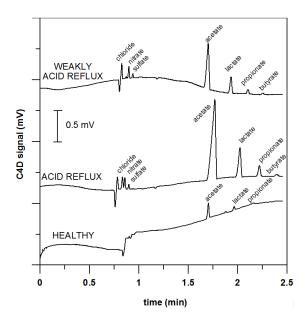


Figure 4: Separation of anions in EBC samples of healthy persons as well as patients with acid reflux or weakly acid reflux. BGE: 20 mM MES/L-His, 2 mM 18-Crown-6, 30  $\mu$ M CTAB. HV: positive +15 kV, C4D detection.

with the HV electrode placed in the detection vial. The concentrations of all 13 ions in the samples were evaluated using calibration curves measured with standard solutions. The parameters such as the calibration curve, regression coefficient  $(R^2)$  and concentration range are shown in Table 1. In this initial screening the analysis of several samples from the healthy group and patients that were diagnosed as having the acid reflux or weakly acid reflux was attempted and the ions that would be significantly different in these groups identified.

A series of electropherograms of anions and organic acids in a healthy individual, patient with acid reflux and patient with weakly acid reflux is shown in Fig. 4. One can clearly see that the concentrations of several anions, most notably chloride, nitrate and organic acids, are significantly higher in the group of patients than in that of the healthy volunteers. Some small inorganic anions such as nitrite and nitrate are often found in the EBC of patients with respiratory diseases as markers of nitrosative stress. On the other hand, the presence of an increased concentration of chloride and organic acids may be a significant step towards the diagnosis of GERD.

#### 3.3 Analysis of cations in EBC samples

The analysis of cations in the same samples can be seen in Fig. 5. Although higher concentrations of the ammonium cation were found in patients compared to healthy individuals, the content was not significantly different from that of healthy individuals, and this applied for other cations  $(K^+, Na^+, Ca^{2+}, Mg^{2+})$  as well.

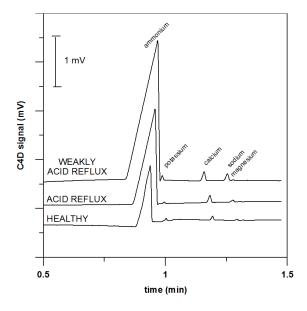


Figure 5: Separation of cations in EBC samples of healthy persons as well as those with acid reflux and weakly acid reflux. BGE: 20 mM MES/L-His, 2 mM 18-Crown-6, 30 μM CTAB. HV: negative -15 kV, C4D detection.

#### 4. Discussion

A limited number of subjects were tested and the determination of whether a simple EBC sample could be used as a surrogate for more invasive, expensive and tedious diagnostic methods attempted. The analysis of anions showed some promising results with regard to the small inorganic anions (chloride and nitrate) collected and the concentration of organic acids that were elevated in the patients

*Table 1:* Calibration equations of all separated anions, cations and organic acids. BGE: 20 mM MES/L-His, 2 mM 18-Crown-6, 30 μM CTAB. HV: positive +15 kV or negative -15 kV, C4D detection.

Ion	Calibration equation	$R^2$	range (μM)
Cl <sup>-</sup>	y = 0.0278x + 0.0258	0.9957	0.6-25
$\mathrm{NO}_2^-$	y = 0.0270x + 0.0023	0.9994	0.6-25
$NO_3^-$	y = 0.0315x + 0.0166	0.9987	0.6-25
$\mathrm{SO}_4^{2-}$	y = 0.0656x + 0.0143	0.9980	0.25-10
acetate	y = 0.0156x + 0.0200	0.9995	1-200
lactate	y = 0.0207x + 0.0097	0.9996	0.6-25
propionate	y = 0.0189x + 0.0024	0.9983	0.6-25
butyrate	y = 0.0188x - 0.0001	0.9990	0.6-25
$\mathrm{NH}_4^+$	y = 0.0211x - 0.1437	0.9993	25-1000
$K^+$	y = 0.0347x + 0.0013	0.9994	0.25-10
$Ca^{2+}$	y = 0.0528x + 0.0059	0.9995	0.6-25
Na <sup>+</sup>	y = 0.0356x + 0.0079	0.9996	0.25-10
$Mg^{2+}$	y = 0.0618x - 0.0054	0.9995	0.25-10

suffering from GERD symptoms, compared to healthy volunteers. Unfortunately, with regard to the analysis of cations, the differences were not significant and cations were deemed unsuitable.

#### 5. Conclusion

Capillary electrophoretic analysis of EBC was used for the first time in an attempt to distinguish the groups of patients with GERD from healthy individuals. CE with C4D was used for the analysis of small inorganic anions, cations and organic acids present in the EBC samples. Although it has been shown that selected samples had elevated concentrations of chloride, nitrate, butyrate and propionate, the number of subjects was too small to draw definite conclusions concerning the discriminatory power of these ions. It also seems that although there is a difference between groups of healthy individuals and sufferers of acid reflux/weakly acid reflux, the results with regard to acid reflux and weakly acid reflux are very similar and these two groups cannot be distinguished based on the current data. A larger scale clinical study, in which the EBC of patients and healthy individuals is scrutinized with regard to the ion content, pH and concentration of other biomarkers (for instance pepsin in saliva) to obtain statistically significant results for evaluation is currently being undertaken. Nevertheless, the initial data is somewhat promising for EBC as a useful non-invasive alternative for other methods used in GERD diagnostics.

# **Symbols**

BGE background electrolyte CE capillary electrophoresis

C4D contactless conductivity detection CTAB cetyltrimethylammonium bromide

DI deionized

EBC exhaled breath condensate
GERD gastroesophageal reflux disease

L-His L-histidine ID inner diameter

MES 2-(N-morpholino)ethanesulfonic acid

OD outer diameter

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