

## TOWARDS BIOCONTROL OF POST-HARVEST ANTHRACNOSE BY ANTAGONISTIC BACTERIA AND YEAST ISOLATED FROM FERMENTED MANGO (*MANGIFERA INDICA*, VAR KENT)

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Abstract: In Côte d'Ivoire, mango fruit plays an important economic role as the third major exported fruit after banana and pineapple. Despite this economic importance, fungal diseases are one of the main issues in the mango value chain with Colletotrichum gloeosporioides causing total postharvest losses estimated up to 20%. The aim of this study was to evaluate the antifungal activity of yeasts, lactic acid bacteria and Bacillus strains isolated from fermented mango fruits as potential biocontrol agents of Colletotrichum gloeosporioides. First, C. gloeosporioides was isolated from contaminated mangoes on potato agar medium, while fermented mangoes were used to isolate yeasts, Bacillus and lactic acid bacteria on DRBC, nutrient and MRS agar, respectively. Afterwards, the in vitro antifungal activity of yeasts, Bacillus and lactic acid bacteria against C. gloeosporioides was tested using dual culture and diffusion methods. The total isolates bateria and yeasts from fermented mango were: 238 yeasts, 214 Bacillus and 252 lactic acid bacteria. The results of antifungal activity of isolated bacteria and yeasts against C. gloeosporioides were as follows: 78 yeasts isolates (33%) were able to inhibit C. gloeosporioides with 11% inhibition more than 40% mycelial growth. The Bacillus isolates were less active (28% inhibition) of C. gloeosporioides with 2% inhibition greater than 60%. As for lactic acid bacteria, they were more active since 58% of isolates showed antifungal activity with 23% of which showed inhibition greater than 60%. Based on the results lactic acid bacteria isolated from fermented mangoes can be used as valuable starters for anthracnose biocontrol.

Keywords: antagonistic microorganisms, biocontrol, Colletotrichum gloeosporioides, mango

#### 1. Introduction

Mango (*Mangifera indica L.*) is one of the most important and commonly eaten fruits in the tropical and subtropical areas. In Côte d'Ivoire, mango fruit plays an important economic role as the third major exported fruit after banana and pineapple. The main producing regions (Korhogo, Ferkessedougou, Sinematiali, Boundiali, Odienne) of mango fruits are in Northern part of Côte d'Ivoire and the most cultivated varieties are Kent, Keitt and Amélie. Despite the economic importance of mango in Côte d'Ivoire, fungal diseases are one of the main issues in the mango chain, occurring during value the production and handling stages [1]. Fungal diseases are known as responsible for postharvest losses in mango production and they also constitute a health risk for consumers due to mycotoxins [2]. Among the fungal diseases, anthracnose caused by gloeosporioides Colletotrichum is considered as one of the most important that negatively impacts mango production causing total losses estimated up to 20% [3.4].

To fight against fungal diseases, chemical fungicides are widely used as treatments to reduce fruit losses from anthracnose [5]. Nevertheless, long-term, and excessive use of chemicals has negative consequences, such as long period of degradation, development of fungicide resistance, harmful effects on human health and the environment [6]. Hence, researchers are working to develop alternative and less strategies for the control of toxic phytopathogens. Among the different methods, biological control appears a promising option for the control of origin fungal diseases of [7.8]. Furthermore, biological control the efficacy of microbial agents has been demonstrated against several postharvest diseases of fruit and vegetables [2].

Biocontrol is essentially based on the use of living microorganisms and/or their metabolites to inhibit the growth or metabolism of phytopathogenic Compared microorganisms [9]. to chemical agents, the use of antagonistic microorganisms has several advantages including specificity of action, absence of and environmentally toxic harmful residues. safer application and more economical production [10]. In addition, biocontrol is known as efficient solution in the short, medium and long term [11]. On the other hand, microbial agents with

antagonistic activity used for the control of fungal plant pathogens are from several taxonomic groups such as bacteria, yeasts and filamentous fungi [12]. However, lactic acid bacteria, Bacillus and yeasts are often used for food biopreservation [13]. The aim of this study was to evaluate the antifungal activity of yeasts, lactic acid bacteria and Bacillus strains isolated from fermented mango fruits as potential of Colletotrichum biocontrol agents gloeosporioides.

# 2. Material and methods

## 2.1. Sampling of mangoes

Plant material consists of mangoes (Mangifera indica var Kent), collected in three (03) producing regions of Northern Côte d'Ivoire: Korhogo (9° 25' 0.001" N 5° 37' 0.001" W), Ferkéssédougou (9° 35' 60" N 5° 12' 0" W), Sinématiali (9° 34' 59.999" N 5° 22' 59.999" W). Mangoes were randomly collected in orchards during harvesting period (mid-May 2022). Two groups of mangoes were collected: healthy mature mangoes for the first group and mangoes with symptoms of anthracnose for the second group (Figure 1). The collected mangoes were put in separated labeled, and transported boxes, to Biotechnology Laboratory, Félix Houphouët-Boigny University) for further analysis.



Fig. 1. Photography of healthy mature Kent mango (A) and contaminated Kent mango with symptoms of anthracnose (B)

#### Isolation morphological 2.2. and Colletotrichum identification of gloeosporioides

Isolation Colletotrichum of gloeosporioides was carried out according to Pitt and Hocking [14]. Mangoes with symptoms of anthracnose (translucent, brown and blackish lesions spots) were washed with sterile distilled water and dried at ambient temperature. After drying, anthracnose lesions on mangoes surfaces were removed using a sterile scalpel and placed on potato dextrose agar (PDA) supplemented medium with 0.1% chloramphenicol. The Petri dishes were incubated at 30 °C for 3 to 5 days. The fungal isolates were subcultured on PDA medium until pure cultures were obtained. Then, pure cultures were incubated for 7 days at 30 °C and stored at 4 °C on agar slant for identification. Macroscopic and microscopic characters of pure isolates were described by using identification keys and Hunter of Barnett [15] after observation under optical microscope at ×40 magnification (ZEISS, Germany).

### **2.3. Fermentation of mangoes**

Fifteen (15) ripe and healthy mangoes were cut into slices of small pieces of about 5 cm. The slices were crushed in a blender (Moulinex, France) briefly for 5 to 10 seconds and the paste was fermented in sterile and hermetically sealed glass jars for 48 h at laboratory temperature (25 °C).

#### 2.4. Isolation and biochemical identification of lactic acid bacteria and **Bacillus** from fermented mangoes

Twenty-five (25 g) grams of fermented mixture from mangoes slices were mixed in 225 mL sterile buffered peptone water. The whole mixture was stirred for 3 to 5 min for homogenization. Afterwards, serial decimal dilutions  $(10^{-2} \text{ to } 10^{-5})$  were carried out by using tryptone salt buffer as

diluent. For lactic acid bacteria, an inoculum of 100 µL of each dilution was spread on MRS agar supplemented with 100 ppm of nystatin into Petri dishes. Afterwards, the plates were incubated at 30 °C for 24-72 h. Isolation of Bacillus sp. was done by inoculation of 100 µL of each dilution on nutient agar [16]. The plates were incubated at 30 °C for 24 to 72 h. Biochemical identification of isolates of lactic acid bacteria and *Bacillus* was based on Gram staining, catalase and oxidase tests.

#### 2.5. Isolation and morphological identification of yeasts from fermented mangoes

The isolation of yeasts from fermented mangoes was carried out by spreading 100  $\mu$ L of each dilution (10<sup>-3</sup> to 10<sup>-5</sup>) on Dichloran Rose Bengal Chloramphenicol (DRBC) agar medium and the Petri dishes were incubated at 30 °C for 24 h. The morphological identification of yeasts isolates was done by observation under optical microscope at ×40 magnification.

### 2.6. Screening for antifungal activity of **Bacillus** sp. and yeasts isolates

The *in vitro* antifungal assay for *Bacillus* and yeasts isolates was carried out based on the dual-culture method [16]. Fungal specie (*Colletotrichum gloeosporioides*) plugs of  $6 \times 6$  mm diameter were placed at the center of PDA medium and 5 µL of 24 hours Bacillus and yeasts grown in nutrient broth were placed on the opposite four sides of the plates at 1.5 cm away from the fungal disc. Plates containing the fungal plugs without Bacillus or yeasts inoculation were used as control. Petri dishes were incubated at 30 °C for 5 days and the growth diameter of *Colletotrichum* gloeosporioides was determined. The percentage of inhibition of the fungal

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growth was calculated using the following formula:

$$I = (R - r) \times 100/R$$

With I - inhibition percentage: r - mycelial ray of Colletotrichum gloeosporioides in the control plate (mm); R \_ mycelial ray of Colletotrichum gloeosporioides in the plate with Bacillus sp. or yeasts (mm)

#### 2.7. Screening for antifungal activity of lactic acid bacteria isolates

Lactic acid bacteria isolates were tested for their antifungal activity against Colletotrichum gloeosporioides according to agar diffusion method [17]. For this, overnight grown of lactic acid bacteria strains were inoculated on MRS agar by using spot technique. The plates were incubated at 30 °C for 48 hours, and then overlaid with 10 mL of PDA soft agar (PDB and 0.7% agar) containing  $10^6$ of spores/mL Colletotrichum gloeosporioides. After the PDA medium was added, the plates were incubated at 30 °C for 3-4 days. The zones of inhibition around the colonies of lactic acid bacteria were recorded and the percentage of

inhibition of the fungal growth was calculated using the following formula:

### I = d x 100/D

With d - diameter of halo around the colonies and D diameter of growth of the control.

## 2.8. Statistical analysis

Excel 2016 (Microsoft Corporation) was used for statistical analysis.

### 3. Results and discussion

#### 3.1. Morphological characteristics of isolated Colletotrichum gloeosporioides

Two macroscopic characteristics of Colletotrichum gloeosporioides were obtained after isolation on potato dextrose agar (PDA) medium (Fig. 2). The first culture showed mycelia with a woolly texture with white color both for face and reverse (Fig. 2.A and 2.B). The second culture highlighted mycelia with a cottony texture, with gray color both for face and reverse (Fig. 2.C and 2.D). In addition, the cultures showed orange conidial mass in the center of plates.

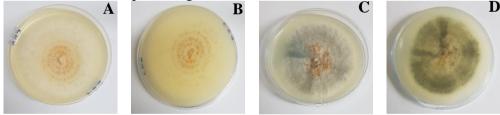


Fig. 2. Photography of macroscopic characteristics of Collectotrichum gloeosporioides on potato dextrose agar (PDA) medium.

Microbial spoilage is one of the most important factors in post-harvest fruits losses. Mangoes, like most commercial fruits, are particularly susceptible to many microbial diseases [18]. Among these diseases, anthracnose is one of the most widespread in tropical Africa [19]. The results of our study were in agreement with those of several authors [20,21]. These authors have demonstrated that Colletotrichum is the main fungi responsible for anthracnose contamination mango fruits. Anthracnose in contamination is characterized by the development of deep, black rot spots or linear necrotic lesions on the fruit [1]. Particularly, the specie *C. gloeosporioides* is the main causative agent of anthracnose in mango [22].

#### 3.2. Morphological and biochemical characteristics of isolated bacteria and veasts from fermented mango

The total isolates bateria and yeasts from fermented mango were: 238 yeasts, 214 Bacillus and 252 lactic acid bacteria. The lactic acid bacteria isolates were Grampositive with a diversity of shape (cocci, coccobacillus, or bacillus) and arrangement mode (clump or chain), catalase negative and oxidase negative. As for Bacillus strains, they were Grampositive and exclusively rod-shaped. The veasts showed a pink color on Dichloran-Rose-Bengal-Chloramphenicol (DRBC) agar medium. The isolation of fermentative microorganisms from mango showed a diversity of microbial groups with occurence of lactic acid bacteria and yeasts. These results were in agreement with those obtained by authors who showed that the surface of mango fruits is

colonized by microorganisms mainly composed of Gram-positive and Gramnegative bacteria and yeast [23].

#### 3.3. In vitro antifungal activity of isolated bacteria and veasts against Colletotrichum gloeosporioides

The results of antifungal activity of isolated bacteria and yeasts against C. gloeosporioides are shown in Table 1 and Figure 3. For yeasts, 78 isolates (33%) were able to inhibit C. gloeosporioides with 11% inhibition more than 40% mycelial growth. The Bacillus isolates were less active (28% inhibition) against C. gloeosporioides with 2% inhibition greater than 60%. As for lactic acid bacteria, they were more active since 58% of isolates showed antifungal activity, and with 23% of this showed inhibition greater than 60%.

Table 1.

Isolates	No inhbition against C. gloeosporioides	Inhibition against C. gloeosporioides		
		I < 40%	40% < I < 60%	I > 60%
Yeasts	160 (67%)	52 (22%)	26 (11%)	0 (0%)
Bacillus sp.	154 (72%)	37 (17%)	19 (9%)	4 (2%)
Lactic acid bacteria	106 (42%)	10 (4%)	78 (31%)	58 (23%)

Inhibition percentage of isolated bacteria and yeasts against C. gloeosporioides

Post-harvest diseases are usually controlled by the application of synthetic fungicides [12]. However, the use of synthetic products leads to various issues such as environmental pollution, toxicity, as well development as the of pathogens resistance. Therefeore, it is essential to adopt alternative approaches as biocontrol for plant diseases management [24].

Based on the antagonism tests, the microbial isolates presented inhibition percentage greater than 25%. Moreover, 50% of lactic acid bacteria isolates inhibited C. gloeosporioides. This result is higher than that (18%) found by some [25]. This result could be authors explained by the fact that tested isolates were from the ecological same environment as C. gloeosporioides.

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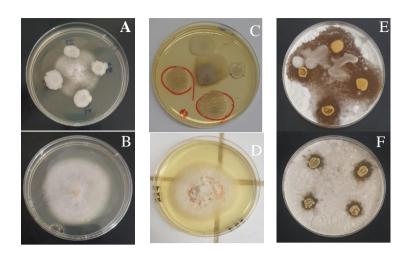


Fig. 3. Photography of *in vitro* inhibition of *C. gloeosporioides* by yeast (A and B), Bacillus (C and D), and lactic acid bacteria (E and F) isolates

Indeed, according to some authors [26], antagonistic microorganisms used control plant diseases should be taken from ecological environment similar to those where the target disease is present.

#### 4. Conclusion

The aim of this study was to evaluate the antifungal activity of yeast, lactic acid bacteria and Bacillus strains isolated from mango fruits as potential fermented agents biocontrol of Colletotrichum gloeosporioides. The results of the present study showed that yeasts, Bacillus and lactic acid bacteria isolates presented inhibition percentage greater than 25% on gloeosporioides. Colletotrichum Thus. yeasts, Bacillus and lactic acid bacteria isolated from fermented mangoes can be used as valuable starters for anthracnose biocontrol. Before their use as starters, molecular identification of yeasts, Bacillus and lactic acid bacteria isolates must be performed.

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#### 6. References

ARCHANA T., GOGOI R., KAUR C., [1]. VARGHESE E., SHARMA R., SRIVASTAV M., TOMAR M., KUMAR M., KUMAR A., Bacterial Volatile Mediated Suppression of Postharvest Anthracnose and Quality Enhancement in Mango, Postharvest Biology and Technology, 177, (2021) https://doi.org/10.1016/j.postharvbio.2021.111525 RANJITH F., MUHIALDIN B., ARROO [2]. R., YUSOF N., MOHAMMED N., HUSSIN A., Lacto-fermented Polypeptides Integrated with Edible Coatings for Mango (Mangifera indica L.) Bio-preservation, Food Control, 134, (2022), https://doi.org/10.1016/j.foodcont.2021.108708 PERUMAL A., NAMBIAR R., PERIYAR [3]. SELVAM SELLAMUTHU P., ROTIMI S., Use of Modified Atmosphere Packaging Combined with Essential Oils for Prolonging Post-Harvest Shelf Life of Mango (cv. Banganapalli and cv. Totapuri), LWT. 148, (2021),https://doi.org/10.1016/j.lwt.2021.111662. KRISHNAPILLAI N., WILSON [4]. WIJERATNAM W., Sap Volatile Components in Relation to Susceptibility of Anthracnose and Aspergillus Rot of Mangoes (Mangifera indica L.), The Journal of Horticultural Science and Biotechnology, 92, 206-213, (2016)https://doi.org/10.1080/14620316.2016.1249962

LIMA N., LIMA W., TOVAR-PEDRAZA [5]. et al. Comparative Epidemiology J. of Species from Colletotrichum Mango in Northeastern Brazil, Eur J Plant Pathol 141, 679-688 (2015). https://doi.org/10.1007/s10658-014-0570-v

ONS L., BYLEMANS D., THEVISSEN K., [6]. CAMMUE B., Combining Biocontrol Agents with Chemical Fungicides for Integrated Plant Fungal Disease Control. Microorganisms, 8, 19-30, (2020), https://doi.org/10.3390/microorganisms8121930

[7]. PU L., LI L., CHAO-AN L., Characterization of Competition for Nutrients in the Biocontrol of Penicillium italicum by Kloeckera apiculata, Biological Control, 67, 157-162, (2013), https://doi.org/10.1016/j.biocontrol.2013.07.011.

LAHLALI R., AKSISSOU W., LYOUSFI [8]. N., EZRARI S., BLENZAR A., TAHIRI A., ENNAHLI S., HRUSTIĆ J., MACLEAN D., AMIRI S., Biocontrol Activity and Putative Mechanism of Bacillus amyloliquefaciens (SF14 and SP10), Alcaligenes faecalis ACBC1, and Pantoea agglomerans ACBP1 against brown rot disease of fruit, Microbial Pathogenesis, 139, (2020),

https://doi.org/10.1016/j.micpath.2019.103914.

CARMONA-HERNANDEZ S., REYES-[9]. PÉREZ J.J., CHIOUITO-CONTRERAS R.G., RINCON-ENRIQUEZ G., CERDAN-CABRERA C., HERNANDEZ-MONTIEL L., Biocontrol of Postharvest Fruit Fungal Diseases by Bacterial Antagonists: A Review. Agronomy, 9, (2019), https://doi.org/10.3390/agronomy9030121

[10]. BONATERRA A., BADOSA E., DARANAS N., FRANCÉS J., ROSELLÓ G., MONTESINOS E., Bacteria as Biological Control Agents of Plant Diseases. Microorganisms, 2022, 10. 17-59 (2022),https://doi.org/10.3390/microorganisms10091759

[11]. CUTHBERT R., DICK J., CALLAGHAN A., DICKEY J., Biological Control Agent Selection Under Environmental Change Using Functional

Responses, Abundances and Fecundities; the Relative Control Potential (RCP) metric, Biological Control, 121, 50-57, (2018).

https://doi.org/10.1016/j.biocontrol.2018.02.008

SANGWAN [12]. DUKARE A., S., MAHESHWARI H., GURU P., KHADE Y., VISHWAKARMA R., Chapter 15 - Utilization of Antagonistic Microbes for the Eco-friendly Management of Fungal Diseases of the Harvested fruits during Postharvest Handling and Storage, Editor(s): Ajay Kumar, Samir Droby, Food Security Plant Disease Management, and Woodhead Publishing, 307-322, (2021),https://doi.org/10.1016/B978-0-12-821843-3.00015-5.

[13]. SALAS M., MOUNIER J., VALENCE F., COTON M., THIERRY A., COTON E., Antifungal Microbial Agents for Food Biopreservation-A Review. Microorganisms, 2017, 8, 10.3390/microorganisms5030037. PMID: 28698479; PMCID: PMC5620628

[14]. PITT J., HOCKING A., Fungi and Food Spoilage. Springer, 3rd edition, New York, USA, 524p. (2009).

[15]. BARNETT H., HUNTER B., Illustrated Genera of Imperfect Fungi. Mineapolis: Burgress Publishing Company, Minneapolis MN, 241 p., (1972).

[16]. SUN P., CUI J., JIA X., WANG W., Isolation and Characterization of Bacillus Amyloliquefaciens L-1 for Biocontrol of Pear Ring Rot. Horticultural Plant Journal, 3, 183-189, (2017).

[17]. CHEONG E., SANDHU A., JAYABALAN J., ZWIELEHNER J., BANSAL N., TURNER M., Isolation of Lactic Acid Bacteria with Antifungal Activity Against the Common Cheese Spoilage Mold Penicillium Commune and their Potential as Biopreservatives in Cheese. Food Control, 46, 91-97, (2014).

[18]. ZHENG M., SHI J., WANG O., LI Y., Antimicrobial Effects of Volatiles Produced by two Antagonistic Bacillus strains on the Anthracnose Pathogen in Postharvest Mangoes. Biological Control, 65, 200-206, (2013).

[19]. ARAUZ L., Mango Anthracnose: Economic Impact and Current Options for Integrated Management. Plant Diseases, 84, 600-611, (2000).

[20]. DEMBELE D., KAMARA A., GRECHI I., SILUÉ N., N'GORAN N., YÉO Y., REY J., KONÉ D., Morphological Characteristics and Distribution of Colletotrichum isolates Morphotypes Infecting Mango (Mangifera indica L.) in the North of Côte d'Ivoire. African Journal of Food, Agriculture, Nutrition and Development, 20 :15837-15856, (2020).

[21]. LI Q., BU J., SHU J., YU Z., TANG L., HUANG S., GUO T., MO J., LUO S., SOLANGI G., Colletotrichum Species Associated with Mango in Southern China. Scientific reports, 9, (2019).

[22]. HERNANDEZ-MONTIEL L., ZULUETA-RODRIGUEZ R., ANGULO C., RUEDA-PUENTE Е., OUINONEZ-AGUILAR E.. GALICIA R., Marine Yeasts and Bacteria as Biological Control Agents Against Anthracnose on Mango. Journal of Phytopathology, 165, 833-840, (2017).

[23]. ABADIAS M., USALL J., ANGUERA M., SOLSONA C., VIÑAS I., Microbiological Quality Minimally-Processed of Fresh, Fruit and Vegetables, and Sprouts from Retail

Establishments. International Journal of Food Microbiology, 123, 121-129, (2008).

[24]. ROMANAZZI G., LICHTER A., GABLER F., SMILANICK J., Recent Advances on the Use of Natural and Safe Alternatives to Conventional Methods to Control Postharvest Gray Mold of Table Grapes. Postharvest Biology and Technology, 63, 141-147, (2012).

[25]. BARRIOS-ROBLERO C., ROSAS-QUIJANO R., SALVADOR-FIGUEROA M., GÁLVEZ-LÓPEZ D., VÁZQUEZ-OVANDO A., Antifungal Lactic Acid Bacteria Isolated from Fermented Beverages with Activity Against Colletotrichum gloeosporioides. Food Bioscience, 29, 47-54, (2019).

[26]. JANISIEWICZ W., KORSTEN L., Biological Control of Postharvest Diseases of Fruits. Annual Review of Phytopathology, 40, 411-441, (2002).