



NUTRITIONAL CHARACTERIZATION OF THE PERIPHERAL LAYERS OF

BARLEY GRAINS CULTIVATED IN ALGERIA

Rawda BENGUELLA¹, *Samira MEZIANI¹, Hayet MEHIDA¹, Yamina BELKESSAM¹, Kouider SENNOUS², Abassia DEMMOUCHE¹

 ¹Laboratory of Biotoxicology, Faculty of of Life Sciences and nature sciences, Department of Biology, University of Sidi-Bel-Abbes, Algeria, meziani_samira@ yahoo.fr
²Laboratory of Ecodevelopment of Spaces, department of environmental science, faculty of natural and life sciences, Djilali Liabes University of Sidi Bel Abbès, Algeria *Corresponding author: meziani_samira@yahoo.fr
Received 22th September 2021, accepted 30th December 2021

Abstract: The purpose of this paper is to stady the different analyzes were carried out in this work on the peripheral layers (PL) of barley grains of the Saida variety. Barley is the most important cereal rich in primary metabolite than secondary, which are beneficial for human health. Barley flour is based on peripheral layers which are generally intended for cattle feed, for their nutritional values linked to the supply of a food rich in vitamins, proteins, fibers etc. The analysis of polyphenols by liquid chromatography method at high pressure (RP- HPLC-LC 2030C 3D), allowing the detection of a phenolic acid (Syringic acid), the source of brut fibers with a gram of PL is 0.8%. Mineralcontents assessed by the spectrophotometric technique of atomic absorption, reveals that calcium and magnesium are the most abundant with values of 2.8 (mg / kg) and 2.1 (mg / kg) respectively.

Keywords: Wheat, peripheral layer, proteins, fibers, RP-HPLC- LC-2030C 3D.

1. Introduction

Cereals are plants cultivated mainly for its seeds, used in human and animal nutrition, often ground in the form of refined flour or more or less complete, but also in whole grains, we find such as wheat, barley, rye or dicotyledons such as sesame, quinoa, etc. Particularly barley has very interesting biological properties, due to its richness in secondary metabolites which are beneficial to human health. Studies have shown that barley contains phenolic compounds in free form and bound from the outer layer of the bran fraction [1] which have the potential to lower blood cholesterol and glucose levels and contribute to the intestinal microbial balance [2] [3] [4]. However, the peripheral layers of barley kernels (PL) have nutritional potential and contain most of the micronutrients such as fiber [5] and minerals which can greatly contribute to the increase of nutritional quality [6] and reduces of numerous diseases such as colon cancer, diabetes, obesity and cardiovascular disease [7] [8]. Barley is rich in protein, complex carbohydrates and fat-soluble vitamin E (tocols) [2] [9] Phenolic groups are the most abundant phenolic compounds in the walls of barley grains in cell-bound form, the main phenolic acids are ferulic acid, vanillic acid, syringic acid and p-coumaric acid [10] [11]. Several studies have focused on milling barley in order to obtain products other than standard flour from cell walls rich in fiber and bioactive phytochemicals [10] [12]. The ain of this study is to conduct research on the peripheral layers of barley grains of the Saida variety from the Sidi Bel Abbes Algeria region [13] through a more indepth study. An analysis of the polyphenols with the HPLC method was

DOI: https://doi.org/10.4316/fens.2021.042

carried out, determination of the raw fibers, as well as the determination of some minerals by atomic absorption pectrophotometer.

2. Matherials and methods

Plant material and extraction

The objective of this research is to highlight the consumption of the peripheral layers of barley grains (Hordeum Vulgare) of the Saïda variety cultivated in Algeria. A manual dissection method under the binocular lens was adopted to obtain the peripheral layers (PL) at the mature stage; they were kept until analysis to be powdered. A quantity of 10 g of the PL was extracted twice with 200 ml of methanol by the infusion method.

Characterization of Phenolic Compounds by RP-HPLC-LC-2030C **3D** Analysis

The phenolic investigation for the crude extract of barley variety (Saida) was carried out using a high performance liquid chromatography coupled with UV-vis RP-HPLC-LC-2030C detector 3D (Shimadzu Prominence-i; LC-2030C 3D). The analytical were evaluated using a processing data system. The separation was achived on a Supelco C18 column (25cm x4.6mm, 5 μm) at room temperature. The mobile phase consisted of water / acetic acid (pH = 3), phase B consisted of methanol / acetic acid (pH =3). The gradient elution system was as follows at the minute 0.01 min the concentration of solvent B is 5% until the 2nd minute, from the 2nd minute up to the 40th minute the gradient will successively change from 5% to reach 100% of solvent B, then will remain 100% until the 45th and the length wave was 280 nm. Phytochemicals were identified from a combination of retention time and spectral matching.

Mineral content by atomic absorption spectrophotometer

Regarding the mineral assay protocol [14] we used a brand atomic absorption spectrophotometer (SHIMADZU model AA7000). The samples were first mixed with 65% nitric acid, then 30% of hydrogen peroxide H2O2 was added, followed by heating at 90 to 120°C to remove all the solvent, after cooling, hydrolysis followed by filtration were carried out The samples were atomized by a flame (Acetylene-air) and the reading was made at lamps and wavelengths specific to each chemical element, results were expressed as mg / kg of DW.

Raw fabers content (Weende)

The method is based on the solubilization of non-cellulose compounds in solutions of sulfuric acid and potassium hydroxide. 1g of each sample was added to 150 ml of 1.25 % sulfuric acid as well as 3-5 drops of the anti-foaming agent n-octanol, they were boiled for 30 min then the crucibles were washed 3 times with hot distilled water. Then the same steps were repeated three times, but with 1.25% potassium hydroxide except that of the last wash which was with cold deionized water and 25 ml of acetone. The crucibles were removed and then weighed after drying at 105 ° for 1 hour, the content represents the crude fibers plus the ash content (F1), to remove the ash, the crucibles were placed in a muffle furnace at 550° for 3 hours and reweighed after cooling in a desiccator (F2) and the results were expressed in%.

% Crude fiber = $(F1 - F2 / F0) \times 100$ Where: F1: Gross fiber weight + ash F2: Ash weight F0: Weight after grinding

3. Results and discussion

Phenolic **Results** Compounds of Characterization bv **RP-HPLC-LC-**2030C 3D Analysis

The analysis of total polyphenols of crude extract from the peripheral layers of barley grains (Saida) was carried out by the RP-HPLC-LC-2030C 3D method using 4 modes identification. of The chromatogram obtained (Figure 1) showed a peak of syringic acid (4-hydroxy-3,5dimethoxybenzoic acid) within 280 nm. Based on studies by López-Perea et al., 2019 [15] on the quantification of phenolic acids in barley husks showed that ferulic acid and benzoic acid showed a higher concentration with 50% acetone; however, benzoic acid was observed at a high concentration with 80% methanol which is consistent with our results by detection of svringic acid.

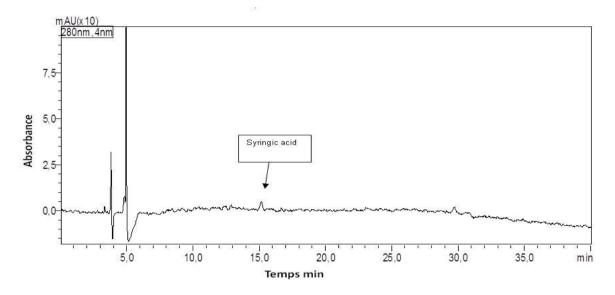


Fig.1. HPLC chromatogram of the crude extracts of peripheral layer of barley (Saida) variety

Results of minerals

The microelement content of the peripheral layers of the wheat grains obtained (Figure 2) shows that calcium is the most abundant mineral with the value of 2.79 mg / kg compared with 2.09 mg / kg of Mg, 0.82 mg / kg of iron, 0.23 mg / kg of Mn and 0.04 mg / kg of Cu. Afify et al., (2016) (16), asserted that sprouted barley have the highest levels of Mg (0.83 \pm 0.03 and 0.34 ± 0.01 mg 100g-1) which confirms our results, as well as the K and Fe contents decreased by the germination of barley and these results are in agreement with that of Hubner et al. (2010) (17), which are decreased from 3.7 to 2.8 mg 100g-1) after germination.

Svetlan and Musa, (2016) [18] found that the Cr and Ni content of malted barley grains at 1.4 mg / kg, while the Cu and Fe content varieds between 0.7-6.8 mg / kg and 57.7-79.9 mg / kg respectively and they observed a strong increase in mineral and metallic elements during malting. Platel et al., (2010) (19) showed that the iron (Fe) and zinc (Zn) content was higher in malted barley than in barley. Youssef et al., 2013 [20], proved that the mineral content in barley was distributed as follows: Ca = 120-160 mg / 100g, Mg = 130-180 mg / 100g, iron = 3.27-39.9 mg / 100g Cu = 0.550-0.985 mg / 100g and Mn= 5.75-13.85 mg / 100g.

Rawda BENGUELLA, Samira MEZIANI, Hayet MEHIDA, Yamina BELKESSAM, Kouider SENNOUS, Abassia DEMMOUCHE, Nutritional characterization of the peripheral layers of barley grains cultivated in Algeria, Food and Environment Safety, Volume XX, Issue 4 – 2021, pag. 411 – 415

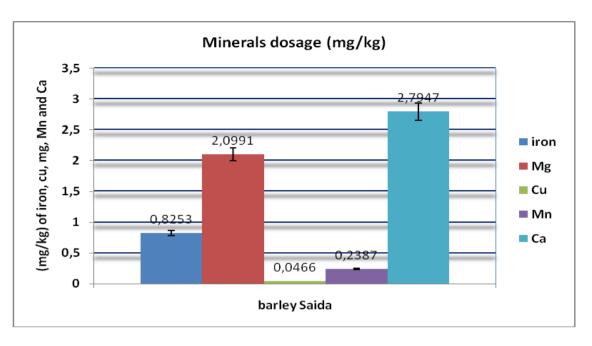


Fig.2. Mineral composition of the peripheral layers of barley grains (Saida)

Raw fiber content

According to the obtained result the crude fiber content is 0.8%, they agree with Montanuci et al., 2013 [21] who reported that the ash content of barley varieties varied from 1.73% to 2.47 % for the two varieties BRs 195 and BRs GRETA, as well as Afify et al., 2016 [16] who showed that the ash content varied from 2.54 \pm 0.07 to 1.90 \pm 0.01% for bare barley and demonstrated that the crude fiber contents before and after germination were; 5.21 \pm 0.11, $4.84 \pm 0.13\%$. Barley without shell is a cereal that can be considered as an energy source of food for livestock feed, according to Yaghoubfar et al., 2013[22], the amount of crude fiber is the lowest (1.4%) compared to the other lines. Sullivan et al, 2010 [18], confirmed that the amount of fiber, specifically insoluble fiber, in whole barley was significantly decreased by the peeling process.

4. Conclusion

In conclusion we can remark that the outer layers of barley kernels rich in fiber and minerals compared to wheat kernels, it

exhibit much nutritional may more properties as whole kernels or as an addition to flour. The establishment of a diet rich in barley bran could replace synthetic antioxidants because of its richness in phenolic compounds which can play an important role in the prevention of certain diseases and valued the basic raw material for recipes of traditional bakery or various pasta products. More studies are needed to get more in-depth results and understand the recommended uses across multiple areas.

5. Acknowledgments

The authors acknowledge the University of Sidi Bel Abbes, Biology Department at Sidi Bel Abbes for the financial support of this research

6. References

[1]. LIU Q.; YAO H., Antioxidant Activities of Barley Seeds Extracts. Food Chem., 102, 732-737. (2007) DOI: 10.1016/j. foodchem. [2]. MARTÍNEZ M., MOTILVA MJ., LÓPEZ H., MC ROMERO MP., VACULOVA K., LUDWIG IA., Phytochemical composition and β-glucan

Rawda BENGUELLA, Samira MEZIANI, Hayet MEHIDA, Yamina BELKESSAM, Kouider SENNOUS, Abassia DEMMOUCHE, Nutritional characterization of the peripheral layers of barley grains cultivated in Algeria, Food and Environment Safety, Volume XX, Issue 4 – 2021, pag. 411 – 415 414

content of barley genotypes from two different human health geographic originsfor food production. Food Chem, 245, 61–70, (2018)

[3]. KOMEILI HR., SHEIKHOLESLAMI Z., Replacement effect of wheat flour with barley flour and hull-less barleyflour on the bread porosity and color, Adv. Agric, Biol. 2, 39-43, (2014)

[4]. CARDONA F., ANDRÉS-LACUEVA C., TULIPANI S., TINAHONES F.J., OUEIPO-ORTUÑO M.I., Benefits of polyphenolson gut microbiota and implications in human health. Journal Nutr. Biochem, 24, 1415–1422, (2013)

[5] MORRISON I., M., & BRICE RE., The digestion of untreated and ammonia-treated barley straw in an artificial rumen. Animal Feed Science and Technology journal, 10(2-3), 229-238, (1984)

[6]. HEMERY Y., ROUAU X., LULLIEN-PELLERIN V., BARRON C., ABECASSIS J., Dry processes to develop wheat fractions and products with enhanced nutritional quality. Journal of Cereal Science, 46(3), 327-347, (2007)

[7]. PRINSEN P., GUTIÉRREZ A., FAULDS C. B., DEL RÍO JC., Comprehensive study of valuable lipophilic phytochemicals in wheat bran. Journal of agricultural and food chemistry, 62(7), 1664-1673, (2014)

[8]. SANG S., ZHU Y., Bioactive phytochemicals in wheat bran for colon cancer prevention. In Wheat and Rice in Disease Prevention and Health, Academic Press, (pp. 121-129), (2014)

[9]. ABDEL-AAL E.SM., CHOO TM., Differences in compositional properties of a hulless barley cultivar grown in23 environments in eastern Canada. Canadien Plant Science Journal, 94, 807-815, (2014)

[10].Idehe E., Tang, Y., Sang S., Bioactive phytochemicals in barley. Food Drug Anal Journal, 25, 148-161, (2017)

[11] GAMEL TH., ABDEL-AAL, ES Phenolic acids and antioxidant properties of barley wholegrain and pearlingfractions. Agric. Food Science Journal, 21, 118–131, (2012)

[12]. IZYDORCZYK MS., MCMILLAN T., BAZIN S., KLETKE J., DUSHNICKY L., DEXTER J., CHEPURNA A, ROSSNAGEL B, Milling of Canadian oats and barley for functional food ingredients: Oat bran and barley fibre-rich fractions.Can. J. Plant Sci, 94, 573-586, (2014)

[13]. BENGUELLA R., MEZIANI S GHUEFFARI I., MENADI N., CHENNI FZ., LABGA L., &DEMMOUCHE, A., Bioactive Compounds in the Peripheral Layers of Barley and Triticale Species in the Mature Grain Cultivated in Algeria. Journal of Drug Delivery and Therapeutics, 10(4), 37-43, (2020)

[14]. MAHOOD HE., Estimation of essential elements and mineral in Catharanthus roseus and its biological importance as a medicinal plant. Plant Cell Biotechnology and Molecular Biology Journal, 22(25), 1-7, (2021)

[15]. LÓPEZ-PEREA P., GUZMÁN-ORTIZ F A., ROMÁN-GUTIÉRREZ AD., CASTRO-ROSAS J., GÓMEZ-ALDAPAC A., RODRÍGUEZ-MARÍN ML., TORRUCO-UCO JG, Bioactive compounds and antioxidant activity of wheat bran and barley extracts husk in the with different polarity. International Journal of Food Properties, 22(1), 646-658, (2019)

[16] AFIFY, A. E. M., ABBAS, M. S., ABD EL-ALI, A. M. Chemical, LATTEFI, B. M., rheological and physical properties of germinated wheat and naked barley. International Journal of ChemTech Research, 9(9), 521-531, (2016)

[17] HÜBNER, F., O'NEIL, T., CASHMAN, K. D., & ARENDT, E. K.. The influence of germination conditions on beta-glucan, dietary fibre and phytate during the germination of oats and barley. European FoodResearch and Technology, 231(1), 27-35, (2010)

[18] SULLIVAN P., O'FLAHERTY J., BRUNTON N., GEE VL, ARENDT E. ET GALLAGHER E., Composition chimique et microstructure des fractions d'orge broyées. Recherche et technologie alimentaires européenne sJournal, 230 (4), 579-595. (2010)

[19]. PLATEL, K., EIPESON, S. W., & SRINIVASAN, K. Bioaccessible mineral content of malted finger millet (Eleusine coracana), wheat (Triticum aestivum), and barley (Hordeum vulgare). Journal of Agricultural and Food Chemistry, 58(13), 8100-8103, (2010)

[20]. YOUSSEF MA., ELKHODARY ST., Utilization of airborne gamma ray spectrometric data for geological mapping, radioactive mineral exploration and environmental monitoring of southeastern Aswan city, South Eastern Desert, Egypt. Geophysical International Journal, 195(3), 1689-1700, (2013)

[21]. MONTANUCI FD., JORGE LMD., JORGE RMM., Kinetic, thermodynamic properties, and optimization of barley hydration. Food Science and Technology Journal, 33, 690-698, (2013)

[22]. YAGHOUBFAR, A., GHAFFARI, S., YOUSEFI, A. Determination nutritive value of hull-less barley cultivars used in poultry nutrition, 15-23, (2013)

Rawda BENGUELLA, Samira MEZIANI, Hayet MEHIDA, Yamina BELKESSAM, Kouider SENNOUS, Abassia DEMMOUCHE, Nutritional characterization of the peripheral layers of barley grains cultivated in Algeria, Food and Environment Safety, Volume XX, Issue 4 – 2021, pag. 411 – 415