



EXTRACTION TEMPERATURE AND pH AS DECISIVE FACTORS FOR THE YIELD AND PURITY OF GRAPE POMACE PECTIN

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Abstract: The purpose of this paper is the use of grape pomaces, obtained by processing two different Vitis vinifera varieties (Rară Neagră and Fetească Neagră) from 2019 harvest, cultivated in the Bugeac area, Republic of Moldova, as a source of unconventional pectin. Pectin was extracted under the following conditions, citric acid used as extractant for 3 h of extraction, 125-200 μ m of grape pomace particle size and solid to liquid ratio 1:10. The variables which were varied were temperature (70, 80 and 90 °C) and pH (1, 2 and 3). So, pectin yield and galacturonic acid content were determined under the influence of extraction temperature and pH. The obtained results demonstrated that the extraction conditions (temperature and pH) have a significant impact on the yield and purity of pectin from grape pomace. The highest influence on pectin yield was determined by pH 1 (15.54% and 9.96% for Fetească and Rară Neagră grape pomace pectin, respectively) and temperature of 90 °C (7.34% and 7.56% for Fetească and Rară Neagră grape pomace pectin, respectively). The same tendency is observed for galacturonic acid content.

Keywords: *grape pomace, galacturonic acid, pectin recovery, citric acid extraction.*

1. Introduction

Pectin is the third group of complex polysaccharides which constitute a part of plant wall characterized higher bv relatively extractability using different type of acids (mineral or organic) or chelators (ammonium oxalate, sodium citrate etc.) and a high content of galacturonic acid [1– 3]. Pectin is found in the middle lamella of the cell wall (CW), with a gradual reduction from the primary CW to the plasma membrane [4]. The structure and firmness of the plant CW depend on the mechanical properties, orientation and link between cellulose and pectic substances [5]. Although, some pectin molecules are linked to xyloglucan chains of the CW [6]. accomplishes Pectin two different functions, as a thickening element on the CW and as a "cementing" component in the middle lamella of plant CW [7]. Pectin is extracted from fruits, vegetables and

their processing by-products (peel, pomace or seeds) [8,9]. Presently, the main sources of commercially pectin are citrus peels and apple pomace which are suitable for specific applications in food industry [4.10.11]. Due to source of pectin. extraction conditions (time. pH. temperature, solid to liquid ratio, acid type etc.) and technique (conventional and unconventional extraction), the pectin has different ability to form gels [8]. Chen et [12] established that extraction al. parameters influence the pectin structure and properties. Moreover, the temperature is a decisive factor [12]. The low temperature kept the pectin structure more intact, remaining more neutral sugars (arabinose, fucose, glucose, galactose, mannose etc.) and possibly more close to the initial molecule in the CW [12]. The pH is considered as one of the most sensitive parameter to note while extracting pectin [13,14]. Commonly, low

acidic pH is essential for the hydrolysis of protopectin [14,15]. A compromise is often made between having a poor quality of pectin, but with higher yield at a lower pH and having a better quality of extracted pectin, but with low yield at a high pH; this trade-off is made between temperature and extraction time [14]. Mainly, the reported values for pH range from 1 to 3, but optimal pH value used for pectin extraction is 2 [4].

Due to the lack of information about how extraction factors influence grape pomace pectin yield and structure, we aimed to study the impact of pH and temperature on the yield and purity of pectin.

2. Materials and methods

2.1. Materials

Grape pomace was obtained by processing two different Vitis vinifera varieties (Rară Neagră and Fetească Neagră) from 2019 harvest, cultivated in the Bugeac area, Republic of Moldova. The grape pomace was dried at 50 °C until constant weight was achieved. The moisture content of dried grape pomace varied between 0.11% and 0.35%. Then, dried pomace was powdered and separated it on the particle size interval of 125-200 µm using an analytical sieve shaker Retsch AS 200 Basic (Retsch GmbH, Haan, Germany).

2.2. Extraction and purification of pectin

Initially, a sample of 10 g grape pomace powder was mixed with 100 mL of solvent (solid-liquid ratio of 1:10, w/v) acquired by adding citric acid to ultrapure (Milli-O) water until a pH 1, 2 and 3 were achieved. Then, the mixtures were kept in a water bath Precisdig (JP Selecta, Barcelona, Spain) at the different temperature (70, 80 and 90 °C) for 3 h.

After extraction, the mixtures were cooled to room temperature, around 20-22 °C. Firstly, the pectin was segregated by centrifugation at 3500 rpm for 35 min. Then, the obtained supernatants were got through clean strainer, plased into the neck of a Duran® laboratory glass bottle with pouring ring and screw cap. Afterwards, ethyl alcohol (>96%, v/v) was added to supernatants in order to achieve 1:1 ratio (v/v). The mixtures were kept at 4-6 °C for 12 h to accomplish the precipitation. The precipitated pectin was separated by centrifugation at 4000 rpm for 30 min. The pectin was washed 3 times with ethyl alcohol (>96%, v/v) and dried in an oven with air circulation Zhicheng ZRD-A5055 (Zhicheng, Shanghai, China) at 50 °C until constant weight was achieved.

2.3. Pectin yield

Pectin yield was calculated using Eq. (1):

Yield (%) =
$$\frac{m_0}{m} \times 100\%$$
 (1)

where: m_0 – weight of dried pectin (g), m - weight of dried grape pomace powder (g) [16,17].

2.4. Galacturonic acid content

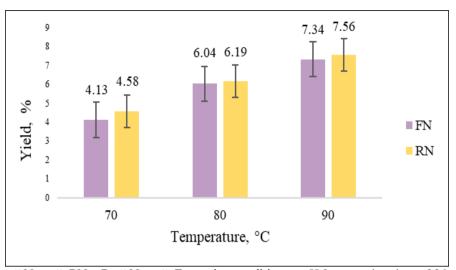
The galacturonic acid content (GalA) of pectin was estimated using the sulfamate/m-hydroxydiphenyl method developed by Filisetti-Cozzi & Carpita [18,19]. Sample preparation was made according to Miceli-Garcia [20].

3. Results and discussion

3.1. Effect of temperature on the pectin vield and galacturonic acid content

In agreement with the results in Fig. 1, yield of pectin was significantly influenced by extraction temperature. It is possible to

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* FN – Fetească Neagră, RN – Rară Neagră. Extraction conditions – pH 2, extraction time of 3 h and 125-200 um of particle size.

Fig. 1 Effect of the temperature on the pectin yield

observe that yield varied between 4.13% and 7.34% for Fetească Neagră (FN) grape pomace pectin and between 4.58% and 7.56% for Rară Neagră (RN) grape pomace pectin depending on the extraction temperature. The obtained results showed that temperature has a major impact on the yield of pectin. A rising temperature from 50 to 95 °C utilizing citric acid at pH 2.5 or 4.0 significantly enhanced the yield of cocoa husks pectin, from 3.58% to 5.66% (extraction temperature of 50 °C and 95 °C, respectively at pH 2.5) and from 3.72% to (extraction temperature of 50 °C and 95 °C, respectively at pH 4.0) [21]. Also, Vriesmann et al. [22], Méndez et al. [23] and Gutöhrlein [24] demonstrated that temperature increasing significantly enhanced the pectin yield from cocoa husks by citric acid, watermelon rind waste by hydrochloric acid and pea hulls by citric acid, respectively. The heated acid solution facilitated to solubilize the pectin and other pectic compounds retained in the CW of plants (protopectin), thus increased of the pectin yield [21,25]. Furthermore, a low value of temperature may be inadequate to allow the protopectin hydrolysis (waterinsoluble pectic substance) by different

acids, thereby achieving a lower pectin yield [21].

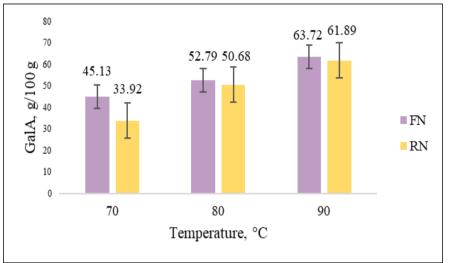
The influence of the temperature on the galacturonic acid content (GalA) is presented in Fig. 2. Therefore, temperature influenced the GalA of grape pomace pectin from both grape pomace varities (FN and RN). The GalA varied from 45.13 g/100 g to 63.72 g/100 g and from 33.92 g/100 g to 61.89 g/100 g for pectin of FN and RN grape pomace, respectively. This is in accordance with the research of Chan & Choo [21] who stated that the uronic acid content of cocoa husks pectin where higher at 95 °C than at 50 °C when was extracted with citric acid at different pH (2.5 or 4.0). Also, Garna et al. [26] demonstrated that 90 °C contributes to a higher GalA obtained from apple pectin then 80 °C, with a sulphuric acid solution at pH 1.5 or 2.0. These results may be ascribed to the increased chemical hydrolysis of pectin sugars when temperature significantly enhances [27-29]. Chaharbaghi et al. [30], who analyzed the optimization of pectin extraction from pistachio hull, noted that the pectin yield

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was enhanced with an increase in extraction temperature.

Thus, they obtained the highest yield of pectin from pistachio hull (23.42%), under

the following conditions: pH 0.5, extraction time of 30 min, temperature of 90 $^{\circ}$ C and a solid to liquid ratio of 50 v/w [30].



* FN – Fetească Neagră, RN – Rară Neagră, GalA – galacturonic acid content. Extraction conditions – pH 2, extraction time of 3 h and 125-200 μm of particle size.

Fig. 2 Effect of the temperature on the galacturonic acid content

A higher temperature increased the yield of pectin due to diffusion of solvent into the structure of CW and so, this movement caused the increase of the mass of polysaccharides releasing from the CW's particles into the acid solution [30]. Further, these conclusions are related to the research data obtained from waste durian rinds and sour orange peel pectin by Maran [31] and Hosseini et al. [32], respectively.

3.2. Effect of pH on the pectin yield and galacturonic acid content

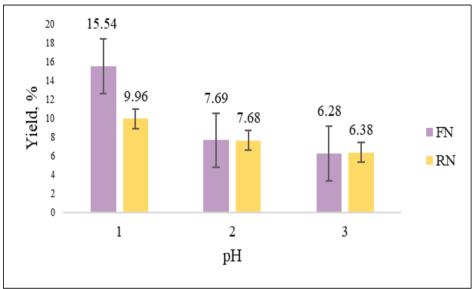
Stronger acid conditions (pH of about 2) play an essential role in influencing the structure and properties of pectin [33]. Fig. 3 presents the influence of pH on the pectin yield, and Fig. 4 illustrates its impact on the galacturonic acid content of pectin. The yield varied between 6.28% and 15.54% for FN grape pomace pectin and between 6.38% and 9.96% for RN

grape pomace pectin. Thus, the highest influence on pectin yield was determined

by pH 1 (15.54% and 9.96% for FN and RN grape pomace pectin, respectively). These results also are in line with the research of Chaharbaghi et al. [30], who studied the extraction optimization of pectin from pistachio hull and obtained the highest pectin yield (23.42%) at pH 0.5. Furthermore, Ma et al. [34] examined the influence of extraction factors on the sugar beet pulp pectin, and noted that decrease in pH increased the pectin yield. This is possibly as a result of CW's lysis in strong acidic solutions and thus, dissolve and release the pectin in acid extract [34]. Also, Raji et al. [35] obtained the maximum extraction yield of pectin from melon peel (29.48%) under pН 1. temperature of 90 °C after 200 min of citric acid extraction.

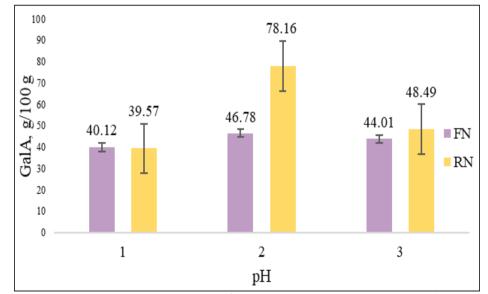
The GalA is a great indicator of pectin purity, thus that a direct relation between this factor and purity of extracted pectin was occured [30,36]. Moreover, according to FAO/WHO the GalA of pectin should not be less than 65% [37]. In current study, the data achieved from used citric acid extraction of pectin showed a GalA range of 40.12-46.78 g/100 g and 39.57-78.16 g/100 g for FN and RN grape pomace pectin, respectively (Fig. 4). The highest

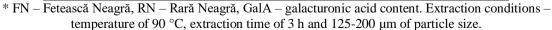
value of GalA (78.16 g/100 g) was achieved for RN grape pomace pectin under the following conditions: pH 2, 90 °C, extraction time of 3 h and 125-200 µm of particle size. Also, this result is nearly related to data acquired from sugar beet pulp (78.8 g/100 g) at pH 2 of citric acid solution, for 2 h of extraction by Ma et al. [34].



* FN - Fetească Neagră, RN - Rară Neagră. Extraction conditions - temperature of 90 °C, extraction time of 3 h and 125-200 µm of particle size.

Fig. 3 Effect of the pH on the pectin yield





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Fig. 4 Effect of the pH on the galacturonic acid content

Furthermore, Yapo [38] presented similar results for GalA (70.6%, w/w) extracted from pectin of yellow passion fruit rind under citric acid extraction at pH 1.8, for 60 min at temperature of 75 °C.

The obtained data confirmed that pH as well as temperature can influence the yield and GalA of extracted pectin from grape pomace.

4. Conclusion

The extraction conditions (temperature and pH) have a significant impact on the yield and purity of pectin from grape pomace. Considering the high yield of pectin and its purity, it can be assured that grape pomace is an unconventional source of pectin.

The highest influence on pectin yield was determined by pH 1 (15.54% and 9.96% for FN and RN grape pomace pectin, respectively) and temperature of 90 °C (7.34% and 7.56% for FN and RN grape pomace pectin, respectively). Moreover, these results are in accordance with previous studies based on unconventional sources of pectin extraction. Additional study about grape pomace pectin should be conducted to the influence of other extraction conditions with application of different extraction techniques.

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6. References

[1]. HARHOLT, J., SUTTANGKAKUL, A., & VIBE SCHELLER, H., Biosynthesis of pectin. Plant Physiology, 153, 384-395, (2010).

[2]. WUSIGALE, L., L., & LUO, Y., Casein and pectin: Structures, interactions, and applications. Trends in Food Science and Technology, 97, 391-403, (2020).

[3]. ZHANG, X., LIN, J., PI, F., ZHANG, T., AI, C., & YU, S., Rheological characterization of RG-I chicory root pectin extracted by hot alkali and chelators. International Journal of Biological Macromolecules, 164, 759-770, (2020).

[4]. PICOT-ALLAIN, M. C. N., RAMASAWMY, & EMMAMBUX, M. N., Extraction, B... characterisation, and application of pectin from tropical and sub-tropical fruits: a review. Food Reviews International, 1–31, (2020).

[5]. ANEES, M., GAO, L., UMER, M. J., YUAN, P., ZHU, H., LU, X., HE, N., GONG, C., KASEB, M. O., ZHAO, S., ... & WENGE, L., Identification of key gene networks associated with cell wall components leading to flesh firmness in watermelon. Frontiers in Plant Science, 12, (2021). [6]. FULLERTON, C. G., PRAKASH, R., NINAN, A. S., ATKINSON, R.G.; SCHAFFER, R.J.; HALLETT, I. C., & SCHRÖDER, R., Fruit from two kiwifruit genotypes with contrasting softening Rates show differences in the xyloglucan and pectin domains of the cell wall. Frontiers in Plant Science, 11. (2020).

[7]. INDURU, J., Pectin-based nanomaterials in drug delivery applications. In Biopolymer-Based Nanomaterials in Drug Delivery and Biomedical Applications, Elsevier, pp. 87-117, (2021).

[8]. CUI, J., ZHAO, C., FENG, L., HAN, Y., DU, H., XIAO, H., & ZHENG, J., Pectins from fruits: Relationships between extraction methods. structural characteristics, and functional properties. Trends in Food Science and Technology, 110, 39-54. (2021).

[9]. FIERASCU, R. C., SIENIAWSKA, E., ORTAN, A., FIERASCU, I., & XIAO, J., Fruits by-products – a source of valuable active principles. A short review. Frontiers in Bioengineering and Biotechnology, 8, (2020).

[10]. MOSLEMI, M. Reviewing the recent advances in application of pectin for technical and health promotion purposes: From laboratory to market. Carbohydrate Polymers, 254, 117324, (2021).

[11]. MARQUES, C, SOTILES, A. R., FARIAS, F. O., OLIVEIRA, G., MITTERER-DALTOÉ, M. L.,

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& MASSON, M. L., Full physicochemical characterization of malic acid: Emphasis in the potential as food ingredient and application in pectin gels. Arabian Journal of Chemistry, 13, 9118–9129, (2020).

[12]. CHEN, J., CHENG, H.; ZHI, Z., ZHANG, H., LINHARDT, R. J., ZHANG, F., CHEN, S., & YE, X., Extraction temperature is a decisive factor for the properties of pectin. Food Hydrocolloids, 112, 106160, (2021).

[13]. LIEW, S. Q., CHIN, N. L., & YUSOF, Y. A., Extraction and characterization of pectin from passion fruit peels. Agriculture and Agricultural Science Procedia, 2, 231-236, (2014).

[14]. ADETUNJI, L. R., ADEKUNLE, A., ORSAT, V., & RAGHAVAN, V. Advances in the pectin production process using novel extraction techniques: A review. Food Hydrocolloids, 62, 239-250, (2017).

[15]. KRAVTCHENKO, T. P., ARNOULD, I., VORAGEN, A. G. J., & PILNIK, W., Improvement of the selective depolymerization of pectic substances by chemical β-elimination in aqueous solution. Carbohydrate Polymers, 19, 237-242, (1992).

[16]. DRANCA, F., & OROIAN, M., Extraction, purification and characterization of pectin from alternative sources with potential technological applications. Food Research International, 113, 327-350, (2018).

[17]. LIEW, S. Q., NGOH, G. C., YUSOFF, R., & TEOH, W. H., Sequential ultrasound-microwave assisted acid extraction (UMAE) of pectin from pomelo peels. International Journal of Biological Macromolecules, 93, 426-435, (2016).

[18]. FILISETTI-COZZI, T. M. C. C., & CARPITA, N. C., Measurement of uronic acids without interference from neutral sugars. Analytical Biochemistry, 197, 157-162, (1991).

[19]. MELTON, L. D., & SMITH, B. G., Determination of the uronic acid content of plant cell walls using a colorimetric assay. Current Protocols in Food Analytical Chemistry, 00, (2001).

[20]. MICELI-GARCIA, L. G., Pectinfrom apple pomace: extraction, characterization, and utilization encapsulating alpha-tocopherol in acetate, University of Nebraska-Lincoln, (2014).

[21]. CHAN, S.-Y., & CHOO, W.-S., Effect of extraction conditions on the yield and chemical properties of pectin from cocoa husks. Food Chemistry, 141, 3752-3758, (2013).

[22]. VRIESMANN, L. C., TEÓFILO, R. F., & DE OLIVEIRA PETKOWICZ, C. L., Extraction and characterization of pectin from cacao pod husks (Theobroma cacao L.) with citric acid. LWT – Food Science and Technology, 49, 108–116, (2012).

[23]. MÉNDEZ, D. A., FABRA, M. J., GÓMEZ-MASCARAQUE, L., LÓPEZ-RUBIO, A., & MARTINEZ-ABAD, A., Modelling the extraction of pectin towards the valorisation of watermelon rind waste. Foods, 10, 738, (2021).

[24]. GUTÖHRLEIN, F., DRUSCH, S., & SCHALOW, S., Extraction of low methoxylated pectin from pea hulls via RSM. Food Hydrocolloids, 102, 105609, (2020).

[25]. GREVE, L. C., MCARDLE, R. N., GOHLKE, J. R., & LABAVITCH, J. M., Impact of heating on carrot firmness: changes in cell wall components. Journal of Agricultural and Food Chemistry, 42, 2900-2906 (1994).

[26]. GARNA, H., MABON, N., ROBERT, C., CORNET, C., NOTT, K., LEGROS, H., WATHELET, B., & PAQUOT, M., Effect of extraction conditions on the yield and purity of apple pomace pectin precipitated but not washed by alcohol. Journal of Food Science, 72, C001-C009, (2007).

[27]. FRAEYE, I., DEROECK, A., DUVETTER, VERLENT, I., HENDRICKX, M., & Т., VANLOEY, A., Influence of pectin properties and processing conditions on thermal pectin degradation. Food Chemistry, 105, 555-563, (2007).

[28]. MAO, G., WU, D., WEI, C., TAO, W., YE, X., LINHARDT, R. J., ORFILA, C., & CHEN, S., conventional and Reconsidering innovative methods for pectin extraction from fruit and vegetable waste: Targeting rhamnogalacturonan I. Trends in Food Science and Technology 94, 65–78, (2019).

[29]. MAO, Y., LEI, R., RYAN, J., ARRUTIA RODRIGUEZ, F., RASTALL, В., CHATZIFRAGKOU, A., WINKWORTH-SMITH, C., HARDING, S. E., IBBETT, R., & BINNER, E., Understanding the influence of processing conditions on the extraction of rhamnogalacturonan-I "hairy" pectin from sugar beet pulp. Food Chemistry X, 2, 100026, (2019).

[30]. CHAHARBAGHI, E., KHODAIYAN, F., & HOSSEINI, S. S., Optimization of pectin extraction from pistachio green hull as a new source. Carbohydrate Polymers, 173, 107–113, (2017).

[31]. MARAN, J. P., Statistical optimization of aqueous extraction of pectin from waste durian rinds. International Journal of Biological Macromolecules, 73, 92–98, (2015).

[32]. HOSSEINI, S. S., KHODAIYAN, F., & YARMAND, M. S., Aqueous extraction of pectin from sour orange peel and its preliminary physicochemical properties. International Journal of Biological Macromolecules, 82, 920–926, (2016).

Mariana SPINEI, Mircea OROIAN, Extraction temperature and pH as decisive factors for the yield and purity of grape pomace pectin, Food and Environment Safety, Volume XX, Issue 4 - 2021, pag. 314 - 321320

[33]. BAI, L., ZHU, P., WANG, W., & WANG, M., The influence of extraction pH on the chemical compositions, macromolecular characteristics, and rheological properties of polysaccharide: The case of okra polysaccharide. Food Hydrocolloids, 102, 105586, (2020).

[34]. Ma, S., Yu, S., Zheng, X., Wang, X., Bao, Q., & Guo, X., Extraction, characterization and spontaneous emulsifying properties of pectin from sugar beet pulp. Carbohydrate Polymers, 98, 750-753, (2013).

[35]. RAJI, Z., KHODAIYAN, F., REZAEI, K., KIANI, H., & HOSSEINI, S. S., Extraction optimization and physicochemical properties of pectin from melon peel. International Journal of Biological Macromolecules, 98, 709–716, (2017).

[36]. LIANG, R., CHEN, J., LIU, W., LIU, C., YU, W., YUAN, M., & ZHOU, X., Extraction, characterization and spontaneous gel-forming property of pectin from creeping fig (Ficus pumila Linn.) seeds. Carbohydrate Polymers, 87, 76-83, (2012).

[37]. Residue Monograph prepared by the meeting of the Joint FAO/WHO Expert Committee on Food Additives (JECFA), 82nd meeting 2016. Pectins, (2016).

[38]. YAPO, B. M., Biochemical characteristics and gelling capacity of pectin from yellow passion fruit rind as affected by acid extractant nature. Journal of Agricultural and Food Chemistry, 57, 1572-1578, (2009).