### PSEUDOMONAS SP. AND AEROMONAS SP. SELECTION FOR TREATMENT OF GOOSE DOWN WASHING WASTEWATER

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**Abstract:** 10 bacterial strains, isolated from municipal wastewater and active sludge that belong to Aeromonas sp. (3) and Pseudomonas sp. (7), were investigated. Strains were analyzed for their ability to treat pre-centrifuged and diluted in different ratios goose down washing wastewater. It is shown that Aeromonas sp. exhibit higher purification degree of goose down washing wastewater compared to the strains of Pseudomonas sp. only at a dilution of 1:1, and the highest purification degree is determined for strains Aeromonas 2 and Aeromonas 2AS with 64,94% each. At a dilution of goose down washing wastewater 1:5 the highest purification degree is at a strain Pseudomonas 1 – 86,49%, and at a dilution of goose down washing wastewater 1:10 Pseudomonas 3AS – 87,27 %.

Keywords: Aeromonas sp., Pseudomonas sp., goose down washing wastewater, purification degree

### **1. Introduction**

From poultry processing as by-product large quantities feathers are produced - 7% of live weight [1]. Utilization of feathers include using of various technologies composting, in food industry after hydrolysis of keratin [2, 3, 4, 5, 6], as adsorbent of heavy metals [7], filling material for clothes and bags [8], in composite wood MDF [9]. Obtaining quality feather include its preliminary treatment with NaCl and HCl solutions, followed by washing it with detergents 2.Materials and methods

### 2.1.Microorganisms

In this work are used 10 strains of bacteria. Bacterial strains *Aeromonas sp. Aeromonas* 2 are isolated from municipal wastewater, while *Aeromonas* 1AS, *Aeromonas* 2AS – from activated sludge. Bacterial strains *Pseudomonas sp.*  and, if necessary - discoloration. As a result large quantities of wastewater with high pollution levels are produced.

Biological methods have application in wastewater treatment from food industry, and many of them are based on the specific action of particularly selected bacterial strains.

The aim of this work is the selection of *Aeromonas sp.* and *Pseudomonas sp.* strains for treatment of goose down washing wastewater.

*Pseudomonas* 1, *Pseudomonas* 3, *Pseudomonas* 4 were isolated from municipal wastewater, while *Pseudomonas* 1AS, *Pseudomonas* 2AS, *Pseudomonas* 3AS, *Pseudomonas* 4AS – from activated sludge.

### 2.2.Nutrient mediums

2.2.1.Luria – Bertany glucose agar medium (LBG) with composition (g/dm<sup>3</sup>): triptone (Difco) – 10 g, yeast extract – 5 g, NaCl – 10 g, glucose (Scharlau) – 10 g; agar – 20 g. pH=7,5. The medium is sterilized for 25 minutes at 121° C.

2.2.2.Glutamate-starch-phenol red agar medium (GSP) with composition (g/dm<sup>3</sup>): sodium glutamate - 10 g, starch (soluble) -20 g, KH<sub>2</sub>PO<sub>4</sub> - 2 g, MgSO<sub>4</sub>, Phenol - red -0,36 g, agar - 12 g. pH 7,2  $\pm$  0,2. All ingredients are dissolved and medium is sterilized for 15 min at 121° C. Cool to 45-50° C and aseptically add to it 100 IU/ml penicillin G, and if necessary, 10 µg/ml pymaricyn. The resulting composition is mixed thoroughly and dispensed in sterile petri dishes.

**2.3. Wastewater:** In the experiments goose down washing wastewater to slaughterhouse is used. Wastewater is precentrifuged at 3000 min<sup>-1</sup> for 10 min to remove insoluble substances and fat and diluted with non-sterile distilled water in ratios 1:1, 1:5 and 1:10.

# 2.4.Cultivation and storage of microorganisms.

Isolated strains are grown in LBG agar medium at 30° C in thermostat for 48 h and stored in a refrigerator at 4°  $\pm$  2° C for 2 weeks.

## 2.5.Analytical methods.

## 2.5.1.Isolation of Aeromonas sp. u Pseudomonas sp. strains.

Isolation of strains of *Aeromonas sp.* and *Pseudomonas sp.* was carried out by seeding samples of municipal wastewater, and active sludge on GSP agar medium and their subsequent development at  $30^{\circ} \pm 2^{\circ}$  C in a thermostat for 24 h. Colored in yellow colonies belong to *Aeromonas sp.*, while red relate to *Pseudomonas sp.* 

**2.5.2.** Development of isolated strains of bacteria in the diluted wastewater. With biomass from developed on LBG agar medium at  $30^\circ \pm 2^\circ$  C for 48 h bacterial strains 5 cm<sup>3</sup> of diluted wastewater is inoculated. Wastewater samples with investigated cultures are cultivated for 72 h at  $30^\circ \pm 2^\circ$  C.

# **2.5.3.***Purification degree wastewater determination using the permanganate oxidizability method.*

Developed for 72 h at  $30^{\circ} \pm 2^{\circ}$  C in wastewater cultures are centrifuged at 3000 min<sup>-1</sup> for 10 min. The resulting supernatant is analyzed for permanganate oxidizability in accordance with standard BS 17.1.4.16-79.

100 cm<sup>3</sup> sample or lower volume brought to 100 cm<sup>3</sup> with distilled water is placed in a 250-300  $\text{cm}^3$  flask, which has added pumice. 5 cm<sup>3</sup> H<sub>2</sub>SO<sub>4</sub> (1+2) and 20 cm<sup>3</sup> 0,01 N KMnO<sub>4</sub> solution are added. The composition is heated so as to boil for no more than 5 min and boiling for 10 min. To the hot solution  $20 \text{ cm}^3 0.01 \text{ N}$  solution HOOC-COOH are added. of Hot discoloured solution is titrated with 0,01 N KMnO<sub>4</sub> solution. The temperature of the solution during titration should not be below 80° C. If the boiling solution is discolored or brown precipitates are formed, the determination is repeated with a smaller sample size.

During titration should not spend more than  $12 \text{ cm}^3$ , and not less than  $4 \text{ cm}^3$  of KMnO<sub>4</sub> solution for diluted sample. Similarly, a blank is done with wastewater. Oxidizability (X) in mg/dm<sup>3</sup> oxygen is given by the formula:

$$X = \frac{(a-b).N.8000}{V},$$

where

 $a - V_{KMnO4}$ , spent for titration of the sample,  $cm^3$ ;

b –  $V_{KMnO4}$ , spent for titration of blank, cm<sup>3</sup>;

 $N\,-\,exact$  normality of  $KMnO_4$  solution;

V - volume of sample taken for analysis,  $cm^3$ .

Similarly oxidizability of a control sample of analyzed wastewater at the corresponding dilution of 1:1, 1:5 and 1:10 is determined.

Purification degree PD, in %, is determined using the formula:

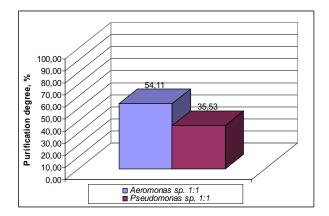
 $PD = \frac{Xc - Xs}{Xs} \cdot 100,\%$ , where

Xc – oxidizability of control sample, mg/dm<sup>3</sup> oxygen;

Xs – oxidizability of sample, mg/dm<sup>3</sup> oxygen.

### 3. Results and discussion.

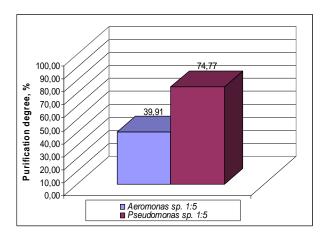
In a set of experiments the ability of isolated *Aeromonas sp.* and *Pseudomonas sp.* strains to break down pollutants in washing goose down wastewater, diluted with distilled water in different ratios, is investigated. Summarized results are presented in Fig. 1, Fig. 2 and Fig. 3. Experimental data show an average level of treatment for all strains of *Aeromonas sp.* and *Pseudomonas sp.* at wastewater dilution 1:1, 1:5 and 1:10, respectively.

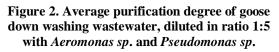


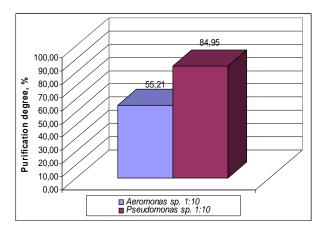
### Figure 1. Average purification degree of goose down washing wastewater, diluted in ratio 1:1 with *Aeromonas sp.* and *Pseudomonas sp.*

It is noteworthy that with increasing dilution increases the degree of purification as in *Aeromonas sp.*, and in *Pseudomonas sp.* Exceptions are strains *Aeromonas sp.* with the degree of dilution 1:5 (Fig. 2).

Strains Aeromonas sp. purify wastewater better than the strains *Pseudomonas sp.* when diluted 1:1 with 18,58% (Fig. 1), while at 1:5 and 1:10 dilutions of the strains *Pseudomonas sp.* show a higher level of treatment - with 34,86% (Fig. 2) and 29,73% (Fig. 3).







#### Figure 3. Average purification degree of goose down washing wastewater, diluted in ratio 1:10 with *Aeromonas sp.* and *Pseudomonas sp.*

A comparative characteristic between strains of *Aeromonas sp.* and *Pseudomonas sp.* for the purification degree of goose down wastewater at different dilution ratios is done. Results from these studies are presented in Figure 4, Figure 5 and Figure 6. The experimental data represented in Figure 4 show that with purification degree over 50% are strains Aeromonas 2, Aeromonas 2AS, Pseudomonas 3, Pseudomonas 2AS and the highest is the degree at Aeromonas 2 and Aeromonas 2AS – 64,94%. Strain *Pseudomonas* 3 exceeds the other three *Pseudomonas sp.* with the purification degree 59, 09 %.

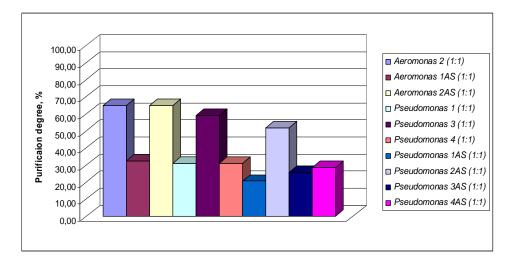


Figure 4. Purification degree of goose down washing wastewater, diluted in ratio 1:1 with *Aeromonas sp.* and *Pseudomonas sp.* 

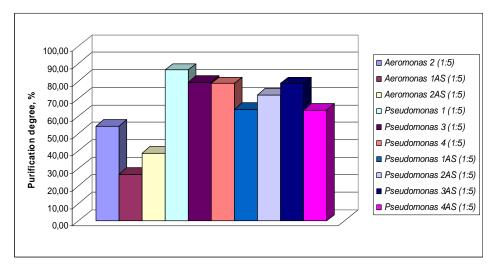


Figure 5. Purification degree of goose down washing wastewater, diluted in ratio 1:5 with *Aeromonas sp.* and *Pseudomonas sp.* 

At a dilution ratio 1:5 (Figure 5) strain *Pseudomonas* 1 shows the highest degree of purification – 86,49%, followed by strain *Pseudomonas* 3 – 79,48 %. Third place is shared by strains *Pseudomonas* 4 and *Pseudomonas* 3AS - with average value 78,70 %. From *Aeromonas sp.* with the highest purification degree is strain *Aeromonas* 2 with 54,23%.

At 1:10 dilution ratio (Figure 6) the highest purification degree has strain Pseudomonas 3AS – 87,27%, followed by Pseudomonas 3 and Pseudomonas 1 with 87.19% and 87.01%. respectively. Purification degree of Aeromonas sp. strains reached only 57,95% of Aeromonas 2AS, followed by Aeromonas 2 and Aeromonas 1AS with 56,92% and 50,77%, respectively.

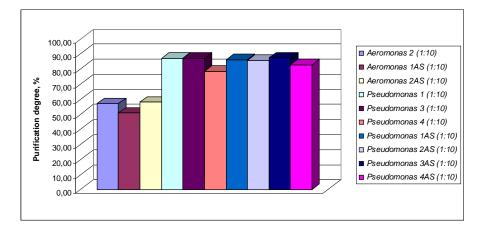


Figure 6. Purification degree of goose down washing wastewater, diluted in ratio 1:10 with *Aeromonas sp.* and *Pseudomonas sp.* 

### 4. Conclusion.

As a result of investigated analyses on goose down washing wastewater treatment the following more important conclusions can be made:

1. The more dilution of goose down washing wastewater is done, the greater is its purification degree. Exception is made for strains *Aeromonas sp.* with dilution factor of the effluent 1:5;

2.Strains *Aeromonas sp.* exhibit higher purification degree of goose down washing wastewater compared to *Pseudomonas sp.* strains only at a dilution factor 1:1.

3.At a dilution factor of goose down washing wastewater 1:1 the highest purification degree is determined at strains *Aeromonas* 2 and *Aeromonas* 2AS with 64,94%, at dilution ratio 1:5 – at strain *Pseudomonas* 1 – 86,49% and at dilution factor 1:10 strain *Pseudomonas* 3AS – 87,27%.

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