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ACTIVATING THE GLUTATHIONE SYSTEM OF *EISENIA FETIDA* DURING EXPOSURE TO CONTAMINATED SILT SLUDGE

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Abstract. In order to identify effective and inexpensive method of purifying wastewater sludge, we tested the hypothesis about the ability of earthworm Eisenia fetida to detoxify the silt complex of different chemical components through the activation of glutathione system. In the laboratory conditions, earthworms were exposed to silt from the sedimentation tanks during 112 days in the boxes, while the control earthworms were on a sunflower husk substrate. Glutathione S-transferase (GST) activity and reduced glutathione (GSH) content in coelomic fluid of earthworms were measured spectrophotometrically. The results revealed biomass of the experimental E. fetida worms was declined by 69% compared to control after the exposure. Heavy metals content in the silt decreased in the range from 13% (for zinc and lead) to almost 100% in the case of cadmium compared to beginning of the experiment. The GST activity increased significantly (87% above the control level), as well as GSH content (36% above the control). Induction of 4 new proteins biosynthesis was found by gel-electrophoresis exploration of coelomic fluid proteins of the worms exposed to contaminated silt. Study results indicate a high level of toxicant-induced activation of the earthworm glutathione-dependent system, and confirmed the efficiency of E. fetida using for detoxification of silt from the sedimentation tanks in the case of complex contamination.

Key words: earthworm, wastewater sludge, heavy metals, detoxification, glutathione S-transferase, glutathione.

1. Introduction

In Ukraine, the annual accumulation of compacted sewage sludge (CSS) in the sedimentation tanks reaches 40 million tons, implying a need to occupy 120 hectares of natural land for storage silt on the drying beds and burning [1]. Such actions ultimately lead to pollution of atmosphere, surface and ground water, soil, and vegetation, and thereby deteriorate the quality of human life. Silt wastewater sludge utilization is also actual problem in many countries [2], [3], where the CSS application includes the addition to agricultural and other soils. Silt wastewater sludge contains a

amount of various high organic compounds [4], heavy metals and trace elements [5], so the pretreatment is required for the successful application of CSS [3]. One of the affordable and effective methods of silt sludge cleaning might be ensured by vermiculture, in particular Eisenia fetida culture. The final product of processing is the vermicompost (or biohumus), which can be used as fertilizer, and for the soils restoration and cleaning from radionuclides, heavy metals and pesticide residues as well. It has been found [6] that earthworms can bio-

accumulate, biodegrade bioor transform the heavy metals and a wider range of other xenobiotics, such as herbicides [7] and veterinary drugs [8]. It was established [9], [10], that resistance of E. fetida to the toxins action has always been associated with activation of the metabolic defense including the glutathione-dependent system. In particular, the increase in glutathione-S-transferase (GST) activity was found towards the pesticides [7], [11] and heavy metals [12], [13]. On the other hand, the high inhibition of E. fetida GST activity was observed due to the action of carbamate pesticide carbaryl [14]. The goals of our work were (1) to evaluate the detoxifying ability of E. fetida in the case of integrated substrate contamination and (2) to identify the role of earthworm glutathione-dependent system in the substrate cleaning processes.

2. Materials and methods

The object of the study (vermiculture of earthworm *Eisenia fetida*) was provided by the Research Institute "Biotechnology" State University, Dnipropetrovsk; specification 3336406.00 2–95.

Substrates for vermiculture were prepared on the basis of modified sunflower husk (SH, control); mixed husk of sunflower, rice and buckwheat (MH), and compacted sewage sludge (CSS) from the sedimentation tanks. Each substrate was ground, and the fraction 200 - 500 microns was subjected to fermentation in the clamps 50 - 60 cm high, while maintaining 70 - 80% humidity. Process of substrates fermentation provided by microflora activity, lasted 14 days at a daily stirring. The fermented substrates were used for vermiculture, and the E. fetida worms were placed in substrate with layer thickness 30 cm at a density of 5 – 10 thousand of the earthworms per square meter. The following conditions been complied during the experiment: pH level of substrate was in the range 6.5 - 7.5, humidity reached 70 - 80%, and the temperature fluctuation was within $20 - 25^{\circ}$ C. For each substrate, three replicates were prepared. The worms was selected at 112 day and placed on wet filter paper to remove the gut content. After 3 days with daily replacement of filter paper coelomic fluid was extracted, frozen, and stored at -20° C.

The average biomass weight of *E. fetida* from each substrate was determined by weighing a group on analytical scales. Earthworm's weight measurement was carried out during the whole experiment, selecting samples every 14 days. Parameters of growth inhibition for the various exposure periods were computed according to [15].

The content of heavy metals was determined by atomic absorption method in the substrates before placing of earthworms and after their removal at the end of 112 days.

Activity of glutathione-S-transferase (GST, EC 2.5.1.18) in the coelomic fluid was measured according to the method described by Habig et al. [16] and Saint-Denis et al. [9] with 1-chloro-2,4-dinitro-benzene (CDNB) as а substrate. The assay mixture (100 µl of 0.1 M/L Tris buffer, pH 8.0, 100 µl GSH, and 200 µl sample) was incubated during 10 min at 30° C, and the reaction was initiated by addition of 100 μ l CDNB. Optical density change was detected at 340 nm during four minutes, and GST activity was expressed in nMol CDNB sec⁻¹ml⁻¹ (nkatal ml⁻¹).

The reduced glutathione (GSH) content determination in the coelomic fluid was conducted by the method of Anderson [17], based on spectrophotometric

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registration of the glutathione reaction with the Ellman's reagent (dithiobis-2nitrodenzoic acid). Optical density of assay mixture (110 μ l of 0.1 m/L Kphosphate buffer contained EDTA, and 300 μ l of sample) was detected at 412 nm after the addition of Ellman's reagent. GSH content was calculated by using calibration graph, and expressed in nMol GSH ml⁻¹.

Protein spectrum study of E. *fetida* coelomic fluid was carried out by a modified SDS-PAGE method of Laemmly [18] in 7.5 - 17.5% gradient acrylamide gel. Protein samples (50 µg ml⁻¹) were boiled for 5 min at 95° C and applied to the respective wells in the gel. Analysis was run at 20 mA in the electrophoresis unit VE-20 (Russia). Protein standards were run in parallel

with the samples. Bands were stained with 0.25% Coomassie brilliant R-250. All measurements were performed in triplicate at least. Data present mean values and standard deviation (M±SD). Significance of the differences between control and treated samples was estimated with Student's t-test (P < 0.05). Fisher's test was used for analysis of variances, $F_{crit.}(0.05; 4; 4) = 6.39$.

3. Results and discussion

The biomass growth of *E. fetida* exposed to the sunflower husk, as well as to the mixed husk, was a positive trend during the experiment, whereas decrease in weight observed in the case of worms exposed to the silt wastewater sludge (Figure 1).

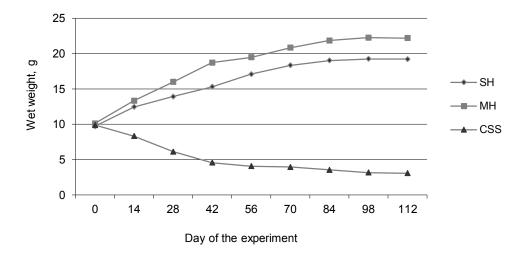


Fig 1. Wet weight of *E. fetida* earthworms exposed to different substrates: sunflower husk (SH, control), mixed husk (MH), and compacted sewage sludge (CSS)

The increase in *E. fetida* biomass at both substrates SH and MH occurred 98 days until the experiment and reached, respectively 198% and 220% compared to initial weight of the worms. At the same time, the wet weight of the earthworms exposed to substrate CSS diminished up to 112 days; weight loss was 16% of control already at 14 days of the experiment, and the final weight loss was 69% of baseline. The results obtained indicated, that growth of E. *fetida* worms exposed to compacted sewage sludge was inhibited drastically. Decrease in the wet weight was observed during whole experiment, however, the most significant decrease was observed up to 42 days, then becoming a less pronounced. A similar non-linear decrease in growth rate of E.

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fetida was occurred under the action of herbicide acetochlor [19]. Likewise, the growth of earthworms *Aporrectodea caliginosa* was decreased noticeably after 7 and 14 days of exposure to insect growth regulators flufenoxuron and pyriproxyfen [20]. In general, growth inhibition is a good indicator of chemical stress, which may link effects to energy dynamics and ultimately inhibit growth of the organisms [15]. In our study, the content of heavy metals discovered in the substrate CSS before the earthworm's introduction, was strongly reduced at the end of the experiment (Table 1).

Table 1

Metal	Content of metal in CSS, mg/kg	
	before the experiment	after the experiment
Cu	76.21±2.54	5.40±0.20
Zn	218.04±8.07	17.32±0.61
Mn	794.10±30.17	54.74±1.86
Ni	29.20±1.05	0.70±0.03
Cr	38.12±1.49	2.54±0.12
Pb	39.03±1.40	3.02±0.10
Cd	5.12±0.19	-

Effect of *E. fetida* vermiculture on heavy metal content in compacted sewage sludge (CSS)

The smallest drop was found for the zinc and lead levels (respectively, 12.6 and 13 times compared to baseline). Decrease in the levels of copper, manganese and chromium was greater (14.1,14.5 and 15.2 times. respectively). Nickel content was reduced to 41.4 times; however, the most striking was the almost 100% reduction in level of cadmium, which was not identified in the substrate after the experiment. That is, the worm's biomass reduction was accompanied by a sharp decrease in heavy metal content in contaminated substrate. It is obvious that inhibition of E. fetida growth could be due to enormous amplification of processes of heavy detoxification metals. Our results are consistent with data [6] about earthworm's ability to accumulate high concentrations of mercury, lead, copper, manganese, zinc, and the extreme cadmium content (up to 100 mg per kg dry weight). It was established [21], that accumulation of a single heavy metal does not affect the worms' physiology; while in our study, the sharp decline in biomass could be

caused by presence of several heavy metals and possibly other toxicants in the substrate.

Increase in activity of glutathione-Stransferase was revealed in coelomic fluids of *E. fetida* exposed to both substrate MH (insignificant difference with control) and CSS (187% compared to control, P < 0.05), as it shown in Figure 2. F-test indicated the difference of data dispersions for two different substrates MH and CSS.

Effective metabolic degradation of different xenobiotics by the earthworms based on the functioning of glutathione-S-transferase, which can catalyze the conjugation of toxic compounds with reduced glutathione, and is involved in antioxidant defense [22]. In our study, almost 2-fold activation of GST was found in coelomic fluid of *E. fetida* after 112 days of exposition to complex contaminated substrate. This result confirms the involvement of *E. fetida* glutathione-S-transferase in detoxification of various components as well as the capacity for long-term

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functioning under the influence of substrate with an integrated pollution.

> MH Substrate

Fig. 2. GST activity (nkatal/ml) in coelomic fluid of *E. fetida* exposed to SH (sunflower husk), MH (mixed husk), and CSS (compacted sewage sludge)

SH

Outcomes are comparable with the data [13] about significant increasing GST activity of *E. fetida* in soils with copper after a period of contaminants accumulation in body tissues, as well, as about GST activation with longer duration and increasing concentrations of herbicide acetochlor [23].

In our study, the control level of reduced glutathione content in the *E*. *fetida* coelomic fluid was exceeded in the worms exposed to substrate MH (10 % above control, P < 0.05), and to substrate CSS (36% above control, P < 0.05) as well (Figure 3). F-test revealed the difference of data dispersions for substrates MH and CSS.

CSS

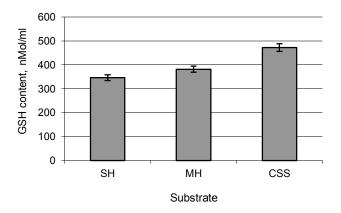


Fig. 3. Effect of substrates on GSH content (nMol/ml) in coelomic fluid of *E. fetida* exposed to SH (sunflower husk), MH (mixed husk), and CSS (compacted sewage sludge)

Thus, amount of reduced glutathione in the *E. fetida* coelomic fluid increased by 1.4 times, indicating enhancement the biosynthesis of glutathione and recovery of the oxidized glutathione during exposition to the contaminated substrate. Reduced glutathione carries out many functions besides conjugation with xenobiotics; in particular, in the worms' cells, GSH was involved in the biosynthesis of metallothioneins due to the action of copper [13] and cadmium

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[24]. In our experiment, GSH pool enhancement was less than GST activation level; this could be attributed to glutathione spending for the synthesis of metallothioneins. Anyway, the glutathione content was sufficient to ensure the growth of GST activity, which is consistent with the assertion [25] about the absence of GST activation in all earthworm species with low GSH levels.

In our study, SDS-PAGE analysis of *E. fetida* coelomic fluids revealed increase in intensity of certain protein bands in the spectra of the worms exposed to both substrates MH and CSS, compared with the control spectrum (Figure 4).

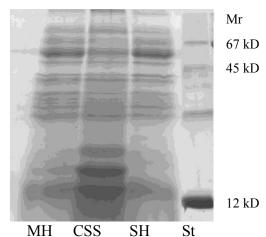


Fig. 4. Polypeptide content of coelomic fluid of the *E. fetida* worms exposed to different substrates: MH – mixed husk; CSS – compacted sewage sludge; SH – sunflower husk (control). St – standard proteins (cytochrome C, $M_r = 12$ kD; egg albumin, $M_r = 45$ kD; albumin bovine serum, $M_r = 67$ kD).

E. fetida coelomic fluid proteins have been shown to have molecular masses ranging from 14.3 kDa to 97.8 kDa. Noteworthy, the components with M_r=18.2; M_r=78.5, and M_r=81.3 kDa were absent in the spectrum of the worms exposed to contaminated silt. At the same time, strengthening the bands corresponding to low-molecular polypeptides with Mr=14.3; Mr=16.6, M_r=21.4 kDa, and to high-molecular zone with M_r=97.8 kDa was observed in the spectrum of the worms exposed to silt compared to control spectrum. Beside them, proteins with $M_r=16.6$; $M_r=41.3$; $M_r=84.1$, and $M_r=87.1$ kDa were presented only in the samples exposed to SCC. Therefore, the appearance of these proteins in the worm's coelomic fluid reflects the changes in the protein metabolism including induction and / or

enhancement of biosynthesis of enzymes responsible for detoxification of contaminated substrate. So, it can be concluded the metabolic systems of *E*. *fetida* generally actively reacted to substrate content, providing a high adaptive capacity of worm organism to polluted environment by means of the xenobiotics detoxification.

4. Conclusion

Summing up, the studies confirmed the ability of *E. fetida* to simultaneously reduce the content of copper, zinc, manganese, nickel, chromium, lead and cadmium in contaminated substrate. Processes of the substrate cleaning against nickel and cadmium were the most effective. Induction of four new proteins biosynthesis along with activation of glutathione-S-transferase

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was found in the coelomic fluid of earthworms exposed to polluted silt. Moreover, increasing pool of reduced glutathione was also observed in the coelomic fluid at the end of earthworms' exposition to contaminated substrate. Hence, vermiculture of *E. fetida* can be an effective way of sewage sludge purification in the case of integrated pollution by enhancing the

6. References

[1] MARKIN V., Utilization of domestic sewage sludge, *Donbas National Academy of Civil Engineering and Architecture*, 3(101): 138-140, (In Russian), (2013)

[2] SIGUA G.C, ADJEI M.B., RECHCIGL J.E., Cumulative and residual effects of repeated sewage sludge applications: forage productivity and soil quality implications in South Florida, USA, *Environ Sci. Pollut. Res. Int.*, 12 (2), 80-88, (2005)

[3] SMITH S.R., A critical review of the bioavailability and impacts of heavy metals in municipal solid waste composts compared to sewage sludge, *Environment International*, 35(1), 142–156, (2009)

[4] ZHANG H., ZHANG Q., YANG B., WANG J., Compacted sewage sludge as a barrier for tailings: the heavy metal speciation and total organic carbon content in the compacted sludge specimen, *PLoS One.*, 30: 9(6), (2014)

[5] BABEL S, DEL MUNDO DACERA D., Heavy metal removal from contaminated sludge for land application: a review, *Waste Manag.*, 26 (9), 988-1004, (2006)

[6] SINHA R.K., CHANDRAN V., SONI B.K., PATEL U., GHOSH A., Earthworms: nature's chemical managers and detoxifying agents in the environment: an innovative study on treatment of toxic wastewaters from the petroleum industry by vermifiltration technology, *Environmentalist*, 32, 445-452, (2012)

[7] ALY M.A, SCHRODER P., Effect of herbicides on glutathione S-transferases in the earthworm, Eisenia fetida, *Environonmental Science and Pollution Research International*, 15(2), 143-149, (2008)

[8] TORKHANI A.L., KOZUH ERZEN N., KOLAR L., VIRANT C.T., LESTAN D.,

activity of glutathione-dependent system.

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Does ivermectin attract earthworms, Journal of

Soils and Sediments, 11(1), 124-128, (2011) [9] SAINT-DENIS M., LABROT F., NARBONNE J.F., RIBERA D., Glutathione, glutathione-related enzymes, and catalase activities in the earthworm Eisenia-fetida-Andrei, *Arch. Environ. Contam. and Toxicol.*, 4 (35), 602-614, (1998)

[10] WANG J.H., ZHU L.S., LIU W., WANG J., XIE H., Biochemical responses of earthworm *(Eisenia foetida)* to the pesticides chlorpyrifos and fenvalerate, *Toxicol. Mech. Methods*, 22(3), 236-241, (2012)

[11] XIAO N.-W., SONG Y., GE F., LIU X.-H., OU-YANG Z.-Y., Biomarkers responses of the earthworm *Eisenia fetida* to acetochlor exposure in OECD soil, *Chemosphere*, 65 (6), 907–912, (2006)

[12] LUKKARI T., Earthworm Responses to Metal Contamination. Tools for Soil Quality Assessment, *Jyvaskyla University Printing House*, Jyvaskyla. 64 p. (2004)

[13] CATALDO J.R., HIDALGO M.E., NEAMAN A., GAETE H.O., Use of molecular biomarkers in *Eisenia foetida* to assess copper toxicity in agricultural soils affected by mining activities, *Journal of Soil Science and Plant Nutrition*, 11(3), 57-70, (2011)

[14] RIBERA D., NARBONNE J.F, ARNAUD C, SAINT-DENIS M., Biochemical responses of the earthworm *Eisenia fetida andrei* exposed to contaminated artificial soil, effects of carbaryl, *Soil Biology and Biochemistry*, 33 (7–8), 1123–1130, (2001)

[15] BADAWY M.E., KENAWY A., EL-ASWAD A.F., Toxicity Assessment of Buprofezin, Lufenuron, and Triflumuron to the Earthworm *Aporrectodea caliginosa*, *International Journal of Zoology*, Article ID 174523, 9 p., (2013)

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[16] HABIG W.H., PABST M.J., JAKOBY W.B., Glutathione S-transferase. The first step in mercapturic acid formation, *J. Biol. Chem.*, 249(22), 7130 – 7139, (1974)

[17] ANDERSON M.E., Determination of glutathione and glutathione disulfide in biological samples, *Methods Enzymol.*, 548 – 555, (1985)

[18] LAEMMLI U.K., Cleavage of structural of bacteriophage T-4, *Nature*, 227, 680-685, (1970)

[19] XIAO N., JING B., GE F, LIU X., The fate of herbicide acetochlor and its toxicity to *Eisenia fetida* under laboratory conditions, *Chemosphere*, 62(8), 1366 – 1373, (2006)

[20] NASR H.M., BADAWY M.E., Biomarker Response and Biomass Toxicity of Earthworms *Aporrectodea caliginosa* Exposed to IGRs Pesticides, *J Environ. Anal. Toxicol.*, 5, 332, (2015)

[21] HARTENSTEIN R., NEUHAUSER E.F., COLLIER J., Accumulation of heavy metals in the earthworm *Eisenia foetida*, *J. Environ. Qual.*, 9, 23-26, (1980) [22] MARTIN-DIAZ M., BLASCO J., SALES D., DELVALLS T., Field validation of a battery of biomarkers to assess sediment quality in Spanish ports, *Environ. Pollut.*, 151, 631-640, (2008)

[23] XIAO N_.W., LIU X_.H., LI W., GE F., Effect of herbicide acetochlor on cytochrome P450 monooxygenases and GST of earthworms *Eisenia fetida*, *J Environ Sci.* (China)., 18(1), 135-140, (2006)

[24] GILLIS P., REYNOLDSON T., DIXON G., Natural variation in a metallothioneinlike protein in *Tubifex tubifex* in the absence of metal exposure, *Ecotox. Environ. Safe*, 58, 22-28, (2004)

[25] HERMANM J., STENERSEN V, QIEN N.I., Glutathione S-transferases in earthworms (Lumbricidae). Substrate specificity, tissue and species distribution and molecular weight, *Comparative Biochemistry and Physiology. Part C Comparative Pharmacology*, 69(2), 243-52, (1981)

Nataliya MITINA, Nina KHROMYKH, Larisa SHUPRANOVA, Inna ZUBAREVA, *Activating the glutathione system of Eisenia Fetida during exposure to contaminated silt sludge*, Food and Environment Safety, Volume XV, Issue 4 – 2016, pag. 276 – 283