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#### Research article

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# Taxonomic notes on the antlion tribe Myrmeleontini Latreille (Neuroptera, Myrmeleontidae, Myrmeleontinae) from Pakistan, with description of a new species

Muhammad Asghar HASSAN <sup>6</sup><sup>1</sup>, Yuchen ZHENG <sup>6</sup><sup>2</sup> & Xingyue LIU <sup>6</sup><sup>3</sup>,\*

1,2,3</sup>Department of Entomology, China Agricultural University, Beijing 100193, China.

\*Corresponding author: xingyue\_liu@yahoo.com

<sup>1</sup>Email: m.hassan93@cau.edu.cn

<sup>2</sup>Email: s20193192649@cau.edu.cn

<sup>1</sup>urn:lsid:zoobank.org:author:75F6DE8B-B0AB-490A-8F3F-B6E2ED7A5BBD <sup>2</sup>urn:lsid:zoobank.org:author:BF21BD87-76FF-48E0-A75C-29C7F473C79E <sup>3</sup>urn:lsid:zoobank.org:author:9BA429B5-F2CB-44BF-BF8E-4D70D016B06B

Abstract. A new species of the myrmeleontine antlion genus *Baliga* Navás, 1912 (Neuroptera: Myrmeleontidae), *Baliga kashmirensis* sp. nov., from Azad Kashmir and Khyber Pakhtunkhwa Province of Pakistan is described and illustrated, representing the first record of *Baliga* from Pakistan. Three species of *Myrmeleon* Linnaeus, 1767 are re-described: *M. hyalinus hyalinus* Olivier, 1811, *M. tenuipennis* Rambur, 1842, and *M. trivialis* Gerstaecker, 1885. *Myrmeleon bimaculatus* Yang, 1999 syn. nov. originally described from China is considered to be a junior synonym of *Myrmeleon tenuipennis* Rambur, 1842. In addition, an annotated catalogue of all species of *Myrmeleon* known from Pakistan along with their distribution map, taxonomical notes and updated identification key to known genera and species are provided.

Keywords. Antlion, lacewings, new taxa, Oriental Region, Kashmir.

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# Introduction

The tribe Myrmeleontini Latreille, 1802 is the most species-rich tribe of the subfamily Myrmeleontinae (Neuroptera: Myrmeleontidae), and currently comprises ten genera, i.e., *Australeon* Miller & Stange, 2012, *Baliga* Navás, 1912, *Dictyoleon* Esben-Petersen, 1923, *Euroleon* Esben-Petersen, 1919, *Hagenomyia* Banks, 1911, *Kirghizoleon* Krivokhatsky & Zakharenko, 1994, *Megistoleon* Navás, 1931, *Myrmeleon* Linnaeus, 1767, *Porrerus* Navás, 1913, and *Weeleus* Navás, 1912, and 228 described species (Machado *et al.* 2019; Hayashi *et al.* 2020). Among these, *Myrmeleon* is the only cosmopolitan antlion genus and comprises the highest proportion of species, with 189 extant described species (Machado *et al.* 2019). *Myrmeleon* is the only genus of Myrmeleontini known from Pakistan, including eight

species that are widely distributed throughout the Oriental and Palearctic parts of the country (Iqbal & Yousuf 1997; Akhtar *et al.* 2018; Hassan *et al.* 2019).

An interesting result in the recent phylogenetic analysis of Myrmeleontidae by Machado et al. (2019) suggested that Baliga, Hagenomyia, together with six genera (Australeon, Dictyoleon, Euroleon, Kirghizoleon, Megistoleon, and Weeleus) are potential synonyms of Myrmeleon. The results of Machado et al. (2019) could also be interpreted that Myrmeleon, as currently circumscribed, is massively paraphyletic. Despite the recent efforts on phylogeny and the larval taxonomy of antlions (Badano et al. 2017b; Machado et al. 2019), there are a few studies that inferred morphological affinities which used adult characters to infer antlion phylogeny (Stange 1994; Badano et al. 2017c, 2018; Machado & Oswald 2020). The larvae of all known genera of Myrmeleontini are pit-builders and exhibit a striking similar morphology. For example, the larvae of Myrmeleon and Euroleon are remarkably similar, but their adults can be distinguished based on wing venational characters (Mansell 1996; Stange 2004; Badano & Pantaleoni 2014; Badano et al. 2017b). In fact, many myrmeleontine genera, especially Baliga and *Hagenomyia*, are based on variable morphological characters, questioning the affinities of several taxa. Navás (1912b) originally described Baliga based on the presence of interconnected crossveins in the costal area proximal to the forewing pterostigma. Later, Esben-Petersen (1913) synonymized this genus with Hagenomyia and remarked that the description of Baliga solely based on the presence of interconnected crossveins is not justified. This view was later followed by Markl (1954), Kuwayama (1962), Oswald & Penny (1991), Ghosh (2000), Bao et al. (2007), Wang et al. (2018), and Yang et al. (2018). However, Stange & Wang (1998), Stange (2004), and Hayashi et al. (2020) considered Baliga as a valid genus based on the shorter length of anterior gonocoxites 8 than posterior gonocoxites 8 (= anterior and posterior gonapophyses in Stange 2004) in the female genitalia (these two sclerites are nearly equal in length in *Hagenomyia*).

In this paper, we first report *Baliga* from Pakistan based on a new species, namely *Baliga kashmirensis* sp. nov. Then, we summarize the present information of Myrmeleontini from Pakistan and add some new findings of this tribe from our recent field surveys in the northern parts of Pakistan. Finally, we also use molecular data, i.e., the partial DNA sequences of the mitochondrial COI and 16S rRNA genes, to verify the validity of the new species herein described. A dichotomous key as well as a distribution map are provided for *Baliga* and *Myrmeleon* species in Pakistan.

# Material and methods

#### **Taxonomy**

The adult specimens were collected at day time (around 10.00 am in the morning to evening at 6 pm) by using a sweeping net along the mountainous regions in Azad Kashmir, Khyber Pakhtunkhwa and Punjab provinces of Pakistan. Samples were preserved in 95% ethanol or pinned. The specimens examined are deposited in the Entomological Museum, China Agricultural University (CAU), Beijing, China and the following institutes in Pakistan: Pakistan Museum of Natural History (PMNH), and the National Insect Museum (NIM), Islamabad. A male of *Baliga sagax* (Walker, 1853) (collecting information: "CHINA: Fujian, Xiamen, Siming District, Huandao Road, Wanyuepo, 0 m, 25.vi.2021, leg. Yuchen Zheng") was used for morphological comparison with *Baliga kashmirensis* sp. nov.

For the species identification, the abdominal segments 7–10 were macerated for 5–6 h in a cold, saturated KOH solution. After rinsing the KOH with acetic acid and water, the apex of the abdomen was transferred to glycerin for further dissection and examination. Terminology mainly follows Badano *et al.* (2017a) for body morphology and wing venation, and Aspöck & Aspöck (2008) for genital sclerites. The species were identified based on following literature: Ohm (1965), Hölzel (1972, 1987), Aspöck *et al.* (1980), Ghosh (1983, 1984, 1990, 2000), Bao *et al.* (2009), Zhan *et al.* (2011), Akhtar *et al.* (2018),

Ábrahám & Giacomino (2020), and Hajiesmaeilian *et al.* (2020). A key is provided for all known and new records of species of *Myrmeleon* from Pakistan.

Photographs of the adult habitus were taken with a Nikon D800 or D850 digital camera with a Nikon Micro-Nikkor 105 mm lens, while the genital photographs were taken with a Canon 7D Mark II or D850 digital camera with a Nikon SMZ18 microscope. Photographs were cleaned up and laid out with Helicon focus (ver. 6.7.1), and Adobe Photoshop CS 6.0. The distribution map was prepared with ArcGIS 10.5 software (Esri, Redlands, CA) by using the original base map of Pakistan.

#### Molecular identification

Representatives of species of *Baliga* and *Myrmeleon* sampled from Pakistan and the published data of the Japanese and Chinese species of these two genera (Hayashi *et al.* 2020) were included for the molecular identification of the species from Pakistan. Moreover, the available COI data of six species of *Myrmeleon* from Pakistan (Akhtar 2018), *Myrmeleon hyalinus* from Egypt and Greece, and two subspecies of *Myrmeleon hyalinus* Olivier, 1811 from Azerbaijan retrieved from the NCBI GenBank were also included for molecular analysis. Since no 16S rRNA gene sequences were available in GenBank for those six species of *Myrmeleon* from Pakistan and two subspecies of *Myrmeleon hyalinus* from Azerbaijan, we used solely the 16S RNA gene sequences to generate the phylogenetic tree for species of *Baliga* and *Myrmeleon* sampled in the present study and those from the Chinese and Japanese species. *Norfolius howensis* (Tillyard, 1917) and *Nymphes myrmeleonoides* Leach, 1814 (Neuroptera: Nymphidae) were selected as outgroups (Winterton *et al.* 2010). The species list, along with their corresponding GenBank accession numbers and collecting data, are provided in Table 1.

The adult specimens were stored in 95% ethanol and refrigerated at -20°C. The genomic DNA was extracted from the thoracic muscle or several legs of each individual sample preserved in ethanol. The samples were incubated in the extraction buffer/proteinase-K mixture at 56°C for 12 h. We used the DNeasy Blood & Tissue kit (QIAGEN, Beijing, China) or TIANamp Micro DNA Kit (TIANGEN BIOTECH CO., LTD, Beijing, China), following manufacturers' instructions. Partial sequences of two mitochondrial genes, i.e., COI and 16S rRNA, were amplified and sequenced. PCR was carried out in a Bio-Rad/T100<sup>TM</sup> Thermal Cycler (Hercules, CA, USA) by using an AccuPower PCR Premix (Bioneer, Daejeon, Korea) with 12.5 L in a reaction volume of 25 L, which included 1.0 L DNA templet, 1.0 L forward primer, 1.0 L reverse primer and 9.5 L distilled water.

Primer sequences used to amplify the two gene fragments as: LEP-F1 (5'-ATTCAAC CAATCATAAAGATATTGG-3') and LEP-R1 (5'-TAAACTTCTGGATGTCCAAAAAATCA-3') for COI (Hebert et al. 2003), and LR-J-12887 (5'-CCGGTTTGAACTCAGATCATGT-3') and SR-N-13398b (5'-CRCYTGTTTAWCAAAAACAT-3') for 16S rRNA (Simon et al. 1994). The PCR was carried out under the following amplification conditions: an initial denaturation step at 94°C (30 s), followed by 39 cycles of denaturation at 95°C (10 s), annealing at 45–50°C (50 s) (depending on primer pair used: 45°C for 16S; 50°C for COI), and elongation at 65°C (1 min), and a final elongation step at 65°C (10 s). The PCR products were electrophoresed in 1% TAE agarose gel stained with Gold View. Sequence assembly was done by using the ContigExpress application. The fragments of COI and 16S rRNA genes were translated into amino acid under the invertebrate mitochondrial genetic code and aligned based on their amino acid sequences by ClustalW in MEGA 7 (Kumar et al. 2016). Genetic distances were calculated using the Kimura 2 Parameter (K2P, Kimura 1980) model in MEGA 7.0 (Kumar et al. 2016). The phylogenetic study included the following three analyses: Neighbour-joining (NJ, Saitou & Nei 1987) was performed in MEGA 7 under the Kumera-2-Parameter (K2P) model; Bayesian inference (BI) analysis was performed using MrBayes on XSEDE (ver. 3.2.7a) (Ronquist et al. 2012) as implemented in the CIPRES science gateway portal (Miller et al. 2010); Maximum likelihood (ML) analyses were performed using the IQ-TREE (Nguyen et al. 2015) as implemented on

Table 1 (continued on next page). List of specimens included in this study.

Family	Species	GenBank accession		Locality	Reference
		number			
	D 1: 1 1 1 :	16S	COI	D-1.:-4 A 1 171	Durant starts
Myrmeleontidae	Baliga kashmirensis sp. nov.	ON263264	ON260954	Pakistan: Azad Kashmir	Present study
	Baliga asakurae	LC582310	LC582458	China: Taiwan	Hayashi et al. 2020
	Baliga asakurae	LC582312	LC582460	China: Taiwan	Hayashi et al. 2020
	Baliga kimurai	LC582297	LC582435	Japan: Iriomote Island	Hayashi et al. 2020
	Baliga kimurai	LC582275	LC582413	Japan: Ishigaki Island	Hayashi et al. 2020
	Baliga kimurai	LC582255	LC582450	Japan: Iriomote Island	Hayashi et al. 2020
	Baliga ryukyuensis	LC582272	LC582410	Japan: Amami Island	Hayashi et al. 2020
	Baliga ryukyuensis	LC582269	LC582406	Japan: Okinawa Island	Hayashi et al. 2020
	Baliga ryukyuensis	LC582263	LC582477	Japan: Okinawa Island	Hayashi et al. 2020
	Baliga micans	LC582264	LC582483	Japan: Fukuejima Island	Hayashi et al. 2020
	Baliga micans	LC582300	LC582437	Japan: Hokkaido	Hayashi et al. 2020
	Baliga micans	LC582288	LC582426	Japan: Nagano	Hayashi et al. 2020
	Myrmeleon tenuipennis	ON263263	ON260959	Pakistan: Islamabad	Present study
	Myrmeleon h. hyalinus	ON255708	ON254389	Pakistan: Islamabad	Present study
	Myrmeleon trivialis	ON255928	ON254214	Pakistan: Azad Kashmir	Present study
	Myrmeleon bore	LC582239	LC582509	Japan: Awashima Island	Hayashi et al. 2020
	Myrmeleon bore	LC582279		Japan: Fukushima	Hayashi et al. 2020
	Myrmeleon bore	LC582277	LC582415	Japan: Hokkaido	Hayashi et al. 2020
	Myrmeleon formicarius	LC582290	LC582428	Japan: Hokkaido	Hayashi et al. 2020
	Myrmeleon formicarius	LC582287	LC582425	Japan: Nagano	Hayashi et al. 2020
	Myrmeleon formicarius	LC582262	LC582454	Japan: Saitama	Hayashi et al. 2020
	Myrmeleon taiwanensis	LC582237	LC582440	Japan: Iriomote Island	Hayashi et al. 2020
	Myrmeleon taiwanensis	LC582247	LC582446	Japan: Iriomote Island	Hayashi et al. 2020
	Myrmeleon taiwanensis	LC582296	LC582434	Japan: Ishigaki Island	Hayashi et al. 2020
	Myrmeleon solers	LC582265	LC582484	Japan: Fukuejima Island	Hayashi <i>et al</i> . 2020
	Myrmeleontidae sp.		KP845997	Pakistan: Azad Kashmir	Akhtar 2018
	Neuroptera sp.		JF839535	Pakistan: Punjab	Akhtar 2018
	Neuroptera sp.		JF839541	Pakistan: Sindh	Akhtar 2018
	Myrmeleontidae sp.		KP846015	Pakistan: Azad Kashmir	Akhtar 2018
	Neuroptera sp.		HQ990310	Pakistan: Punjab	Akhtar 2018
	Neuroptera sp.		JF839546	Pakistan: Punjab	Akhtar 2018
	Myrmeleontidae sp.		KP845978	Pakistan: Punjab	Akhtar 2018
	Myrmeleontidae sp.		KP845928	Pakistan: Punjab	Akhtar 2018
	Myrmeleon h. distinguendus		MT621176	Azerbaijan	Kerimova <i>et al</i> . 2020
	Myrmeleon h. distinguendus		MT621177	Azerbaijan	Kerimova <i>et al</i> . 2020
	Myrmeleon h. hyalinus		MT621174	Azerbaijan	Kerimova <i>et al</i> . 2020

**Table 1** (continued). List of specimens included in this study.

Family	Species	GenBank accession number		Locality	Reference
		16S	COI		
	Myrmeleon hyalinus		KC182603	Greece	Directly submitted
	Myrmeleon hyalinus		KC182604	Egypt	Directly submitted
	Myrmeleon hyalinus		KC182605	Egypt	Directly submitted
hidae	Norfolius howensis	EU734882	EU839749	Australia: Queensland	Winterton 2010
Nymphidae	Nymphes myrmeleonoides	EU734884	EU839751	Australia: Queensland	Winterton 2010

the webserver (http://iqtree.cibiv.univie.ac.at/). Trees were visualized and edited with FIGTREE ver. 1.3.1 (Rambaut 2009).

#### Results

# **Taxonomy**

# Tribe Myrmeleontini Latreille, 1802

## Key to genera of Myrmeleontini from Pakistan

- Antenna obviously swollen; wings relatively narrow; forewing costal area without interconnected crossveins proximal to pterostigmal area (Fig. 7)
   Myrmeleon Linnaeus, 1767

#### Genus Baliga Navás, 1912

Baliga Navás, 1912b: 110. Type species: Myrmeleon asakurae Okamoto, 1910: 297. Original designation.

*Balaga* Navás, 1912b: 111. Type species: *Myrmeleon micans* McLachlan, 1875: 176. Original designation. *Baga* Navás, 1930a: 37. Type species: *Balaga montana* Navás, 1930a: 38. Monotypy.

#### **Diagnosis**

*Baliga* is quite similar to *Hagenomyia* and *Myrmeleon* but can be distinguished from *Hagenomyia* by the female anterior gonocoxites 8 relatively shorter than posterior gonocoxites 8 (these two female genital sclerites are nearly equal length in *Hagenomyia*) and from *Myrmeleon* by the presence of interconnected crossveins in the costal area of forewing (these interconnected crossveins are absent in *Myrmeleon*).

#### Distribution

*Baliga* currently includes 17 described species, predominantly distributed in the Oriental (12 species) and Palaearctic regions (4 spp.: China, Japan, and Korea) with a single species in Australia (Queensland). It is widely distributed in the Oriental region: Bangladesh, China, India, Myanmar, Sri Lanka, Vietnam, and the main islands of Indonesia, Malaysia, Micronesia, Philippines, Japan, and Korea (Ghosh 2000; Stange 2004; Bao *et al.* 2007; Hayashi *et al.* 2020).

# Baliga kashmirensis sp. nov.

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Figs 1-5, 20

# **Diagnosis**

Larger-sized species (forewing length: 34.5–44.8 mm), superficially resembling to *Baliga sagax* (Walker, 1853) based on similar yellow markings on vertex but can be distinguished by the yellow pronotum, with a pair of well separated median longitudinal dark brown stripes and frons mostly shining black, but medially with a narrow longitudinal yellow marking and a narrow median U-shaped yellow marking at ventral corner (Fig. 3B–D).

#### **Etymology**

The new species is named after its type locality, i.e., Azad Kashmir, Pakistan.

## Type material

# Holotype

PAKISTAN • & Azad Kashmir, District Poonch, Rawalakot Valley, Khai Gala; 33°51′4.3194″ N, 73°49′46.3434″ E; 1802 m a.s.l.; 4 Jun. 2019, Hassan M.A. leg.; CAU.

#### **Paratypes**

PAKISTAN • 7  $\circlearrowleft$   $\circlearrowleft$  , 7  $\circlearrowleft$   $\circlearrowleft$  ; Azad Kashmir, District Poonch, Rawalakot Valley, Khai Gala; 33°51′4.3194″N, 73°49′46.3434″ E; 1802 m a.s.l.; 4 Jun.–19 Aug.–3 Sep. 2019 • 2  $\circlearrowleft$   $\circlearrowleft$  , 2  $\circlearrowleft$   $\circlearrowleft$  ; Khyber Pakhtunkhwa Province, District Mansehra, Bajna; 34°27′45.036″ N, 73°15′26.028″ E; 1000 m a.s.l; 19 Jul. 2019; Hassan M.A. leg.; CAU.

# **Description**

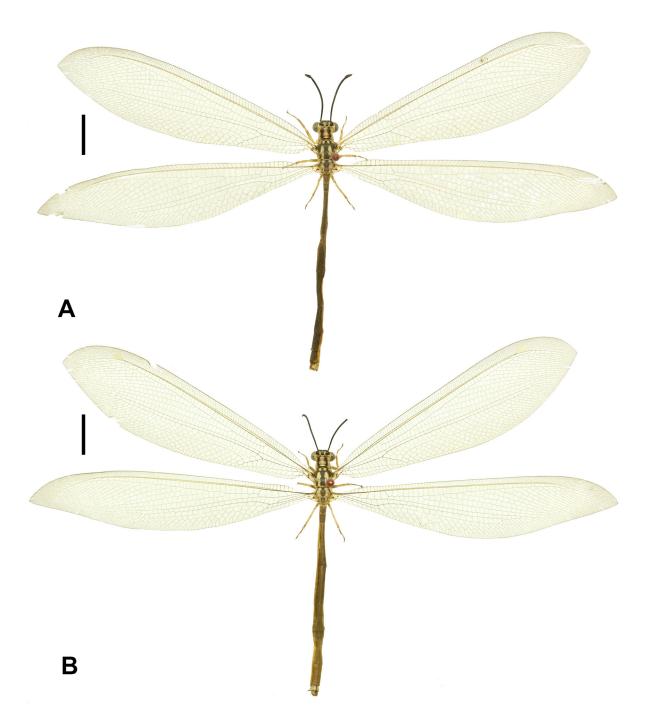
MEASUREMENTS ( $\lozenge$ n=5,  $\lozenge$ n=6). Forewing: length  $\lozenge$  34.5–38.5 mm,  $\lozenge$  37.7–44.8 mm; width  $\lozenge$  9.0–10.6 mm,  $\lozenge$  10.0–12.0 mm. Hind wing: length  $\lozenge$  36.0–39.0 mm,  $\lozenge$  39.0–46.0 mm; width  $\lozenge$  7.8–8.6 mm,  $\lozenge$  10.0 mm. Body length:  $\lozenge$  32.0–35.2 mm,  $\lozenge$  34.0–39.5 mm.

HEAD (Fig. 3A–D). Vertex moderately raised, dark brown with yellow markings; in frontal view dark brown, dorsally with a pair of C-shaped yellow markings, which are connected to ocular rim; in dorsal view dark brown but medially yellow, with a rounded dark brown marking at middle; epicranial area black, with longitudinal transverse grooves, covered with short brownish pubescence. Frons shining black, but medially with a narrow longitudinal yellow marking and a narrow median U-shaped yellow marking at ventral corner, covered with short brownish pubescence. Occiput and postorbital sclerites yellow. Clypeus pale yellow (Fig. 3B) or medially with two small rounded dark brown spots in some specimens (Fig. 3C), and with four median long brown setae. Labrum brownish yellow, with erected brownish setae at proximal margin. Genae pale yellow. Maxillary and labial palps pale yellow, with terminal labial palpomere spindle-shaped, palpimacula brownish, small and circular, with short pale yellow setae. Antennae black, covered with short black setae, dorsal ring of scape and pedicel yellowish brown, flagellum black with several distal flagellomeres widened and moderately flattened. Antennal sclerite yellow (Fig. 3B).

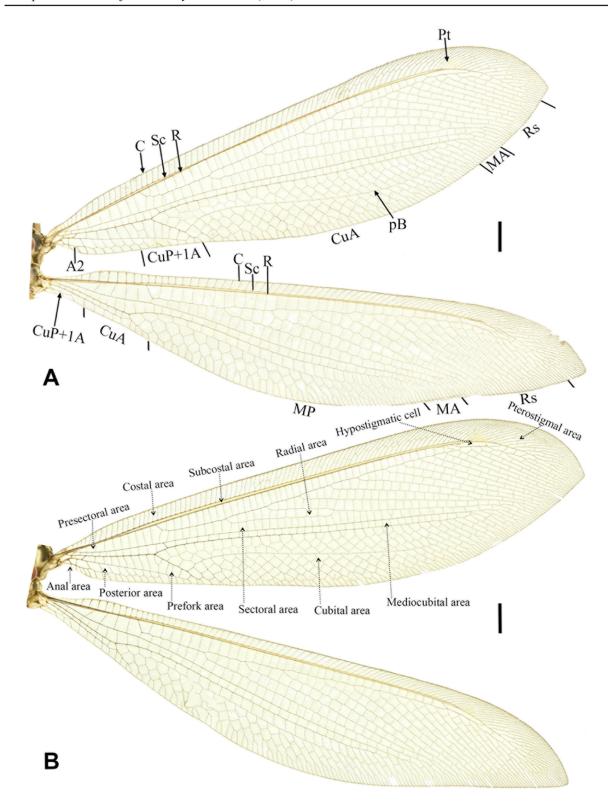
THORAX (Fig. 3A). Pronotum slightly wider than long, yellow, with two median longitudinal dark brown stripes; lateral margins narrowly dark brown after anterior transverse furrow in lateral view; covered with long sparse black setae, but anterolateral margins with short, black and pale yellow setae. Mesonotum yellow with a median and lateral dark brown stripe; prescutum laterally with a pair of rounded yellow spots, covered with long sparse black setae; mesoscutum with a median and lateral dark brown stripe, lateral stripes limited to proximal  $\frac{2}{3}$ , covered with short yellow setae; metascutellum yellow with a broad median longitudinal

dark brown stripe, covered with long sparse yellow setae. Metanotum yellow with a median and lateral dark brown stripe, lateral stripes limited to proximal half of metascutum, covered with short yellow setae. Pleuron yellow, with a median longitudinal dark brown stripe, covered with short sparse yellow setae (Fig. 3F).

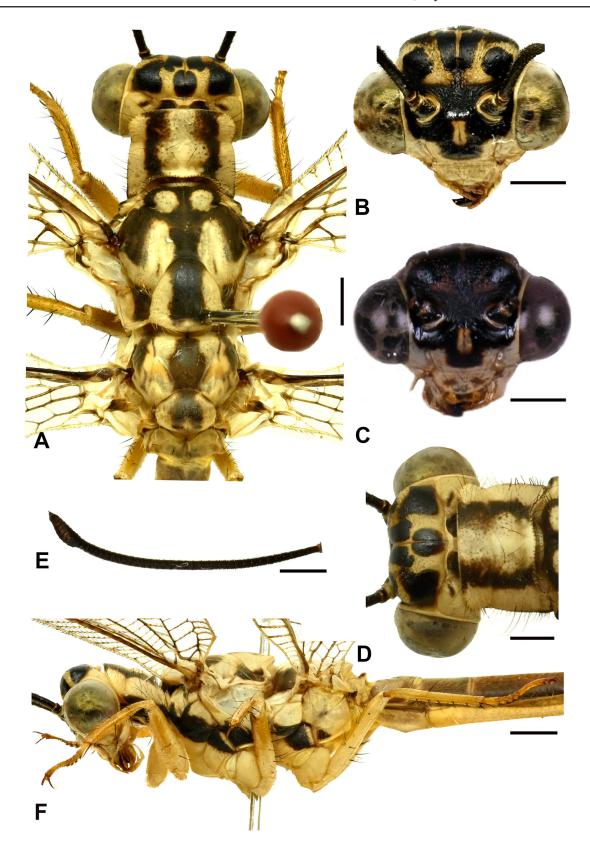
Legs (Fig. 3F). Foreleg: coxa and trochanter yellow, covered with short yellow setae. Femur yellow, covered with short, black and brownish yellow setae, proximal half with a few long black setae; femoral sense hair as long as proximal ½ of profemora. Tibia yellow, with mixed, short and thick black setae,



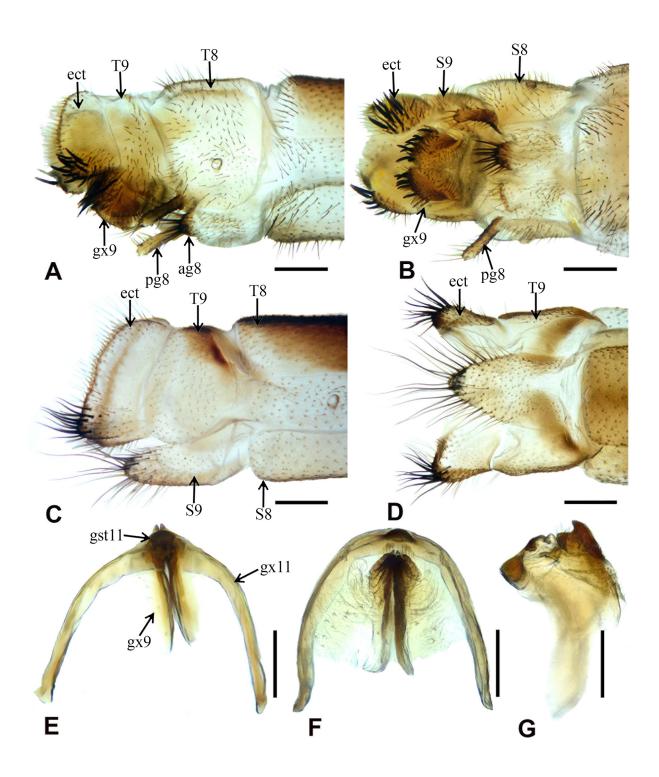
**Fig. 1.** *Baliga kashmirensis* sp. nov. Dorsal habitus. **A**. Male (CAUPK00001). **B**. Female (CAUPK00002). Scale bars = 5.0 mm.



**Fig. 2.** *Baliga kashmirensis* sp. nov. Right fore- and hind wing. **A.** Male (CAUPK00001). **B.** Female (CAUPK00002). Abbreviations: A = anal vein; C = Costa; CuA = cubitus anterior; CuP = cubitus posterior; MA = media anterior; MP = media posterior; pB = posterior Banksian line; Pt = pterostigma; Sc = subcosta; R = radius.Scale bars = 2.0 mm.



**Fig. 3.** *Baliga kashmirensis* sp. nov. **A.** Dorsal habitus of head and thorax. **B–C.** Head, frontal view. **D.** Head and pronotum, dorsal view. **E.** Antenna. **F.** Lateral view. A–B, D–F:  $\lozenge$  (CAUPK00001); C:  $\lozenge$  (CAUPK00003). Scale bars: A = 2.0 mm; B–E = 1.0 mm; F = 3.0 mm.



**Fig. 4.** *Baliga kashmirensis* sp. nov. **A.** Female genitalia, lateral view. **B.** Same, ventral view. **C.** Male genitalia, lateral view. **D.** Same, ventral view. **E.** Complex of gonocoxites 9 + gonocoxites 11, dorsal view. **F.** Same, ventral view. **G.** Same, lateral view. Abbreviations: ag8 = anterior gonocoxites 8; ect = ectoproct; gst11 = gonostylus 11; gx9 = gonocoxites 9; gx11 = gonocoxites 11; pg8 = posterior gonocoxites 8; pp = pregenital plate; S = sternites; T = tergites. A–B:  $\delta$  (CAUPK00002); C–G:  $\varphi$  (CAUPK00001). Scale bars = 0.5 mm.

antennal cleaning setae yellow; tibial spurs brownish, straight, as long as Ta1. Tarsomeres yellow, covered with short black setae; Ta1 equal to combined length of Ta2—Ta3; Ta2, Ta3 and Ta4 nearly equal in size; Ta5 equal to combined length of Ta1—Ta4. Pretarsal claws brownish, curved. Mid leg similar to foreleg. Hind leg similar to mid leg, but femoral sense hair absent.

Wings (Fig. 2). Forewing: relatively broad, subacute at apex; membrane hyaline; longitudinal veins dark brown, except Sc with alternate dark brown and yellow patches, covered with sparse short black setae; costal area with interconnected crossveins proximal to pterostigma for at least ½ of forewing length; seven to nine presectoral crossveins; initial branching point of CuA at same level to Rs origin; Rs with 12–16 branches; pterostigma milky white; anterior Banksian line absent; posterior Banksian line present. Hind wing: slightly longer than forewing, acutely pointed at apex; membrane hyaline; longitudinal veins dark brown, except Sc with alternate dark brown and yellow patches, covered with sparse short black setae; costal veinlets simple, except a few marginally forked crossveins around poststigmal area; five presectoral crossveins; median fork proximal to Rs origin; Rs with 14–18 branches; pterostigma milky white, relatively smaller than that on forewing; anterior Banksian line absent; posterior Banksian line present; pilula axillaris with rounded knob, covered with dense brown setae.

ABDOMEN (Fig. 1). Tergites brownish, covered with short brownish yellow setae. Sterites brownish yellow, covered with short brownish yellow setae.



**Fig. 5.** Habitat of *Baliga kashmirensis* sp. nov. **A–B**. Khyber Pakhtunkhwa: Bajna. **C–D**. Azad Kashmir: Khai Gala.

MALE GENITALIA (Fig. 4C–G). Tergum 9 trapezoidal, with anterior margin slightly prominent in lateral view. Sternum 9 slenderly triangular in ventral view, covered with long black setae at distal half. Ectoproct nearly rectangular in lateral view, posterodorsal margin rounded, covered with yellow setae, posteroventral corner slightly prominent, covered with long thick black setae. Gonocoxites 11 highly sclerotized, lateral arms straight, apex curved ventrad in dorsal view. Gonostylus 11 slightly prominent in ventral view. Gonocoxites 9 broad and elongate, curved with pointed apex in lateral view.

Female Genitalia (Fig. 4A–B). Tergum 9 quadrate in lateral view, with short black setae. Ectoproct subquadrate, posterodorsal margin rounded, with short yellow setae, but proximal ½ with robust digging setae. Anterior gonocoxites 8 short, as long as wide, with thick long black setae. Posterior gonocoxites 8 long, digitiform, with long black setae. Gonocoxites 9 broad and rounded, with robust digging setae, anterolaterally with a bunch of erected short black setae at proximal ½.

#### Distribution

Pakistan (Azad Kashmir and Khyber Pakhtunkhwa).

# Genus Myrmeleon Linnaeus, 1767

Myrmeleon Linnaeus, 1767: 913: Type species: Myrmeleon formicarium Linnaeus, 1767: 914. Subsequent designation by Latreille, 1810: 435.

Macroleon Banks, 1909: 4. Type species: Myrmeleon validus McLachlan, 1894: 515. Original designation.

Enza Navás, 1912b: 113. Type species: Enza otiosus Navás, 1912b: 114. Monotypy.

Myrmeleodes Navás, 1912c: 242. Type species: Myrmeleodes medius Navás, 1912c: 243. Monotypy.

Moreyus Navás, 1914b: 55. Type species: Moreyus brasiliensis Navás, 1914b: 55. Monotypy.

Morter Navás, 1915a: 466. Type species: Myrmeleon hyalinus Olivier, 1811: 126. Original designation. Neleon Navás, 1915b: 53. Type species: Myrmeleon immaculatum De Geer, 1773: 564. Original designation.

*Neseurus* Navás, 1916: 53. Type species: *Myrmeleon alternans* Brullé in Webb & Berthelot, 1839: 83. Original designation.

Cocius Navás, 1919: 296. Type species: Cocius angustatus Navás, 1919: 297. Monotypy.

Leptoleon Esben-Petersen, 1918: 18. Type species: Leptoleon regularis Esben-Petersen, 1918: 18. Monotypy.

*Myrmeleonellus* Esben-Petersen, 1918: 17. Type species: *Myrmeleonellus pallidus* Esben-Petersen, 1918: 18. Monotypy.

Dicholeon Navás, 1920: 193. Type species; Dicholeon nigritarsis Navás, 1920: 193. Monotypy.

Tafanerus Navás, [1921] 1919: 62. Type species: Tafanerus indicus Navás, [1921] 1919: 63. Original designation.

Talosus Navás, 1923a: 35. Type species: Talosus oberthuri Navás, 1923a: 35. Monotypy.

Banya Navás, 1923b: 145. Type species: Banya trifasciata Navás, 1923b: 145. Monotypy.

Grocus Navás, 1925: 185. Type species: Grocus gerstaeckeri Navás, 1925: 185. Original designation.

Colinus Navás, 1925: 187. Type species: Colinus philippinus Navás, 1925: 187. Monotypy.

Afroleon Navás, 1927: 13. Type species: Afroleon basutus Navás, 1927: 13. Monotypy.

Neurocolinus Navás, 1930b: 42. Type species: Colinus philippinus Navás, 1925: 187. Monotypy.

Nemeyus Navás, 1934a: 502. Type species: Nemeyus sanaanus Navás, 1934a: 503. Monotypy.

Nezuela Navás, 1934b: 155. Type species: Nezuela geayana Navás, 1934b: 156. Monotypy.

Bordus Navás, 1936a: 165. Type species: Bordus temeratus Navás, 1936a: 166. Monotypy.

Congoleon Navás, 1936b: 337. Type species: Congoleon sociatus Navás, 1936b: 337. Monotypy.

Hypsoleon Navás, 1936c: 103. Type species: Hypsoleon chappuisinus Navás, 1936c: 103. Monotypy.

Nelneja Navás, 1936c: 104. Type species: Nelneja guttata Navás, 1936c: 105. Monotypy.

#### **Diagnosis**

*Myrmeleon* is similar to *Baliga* by the presence of anterior gonocoxites 8 relatively shorter than posterior gonocoxites 8 in the female genitalia (Fig. 9A–B) but can be distinguished by the absence of interconnected crossveins in the costal area of forewing (Fig. 7).

# Key to species of the genus Myrmeleon from Pakistan\* 2. Clypeus yellow with two median rounded brownish markings; pronotum dark brown, medially with a narrow longitudinal complete yellow stripe, rounded in center, laterally with a narrow yellow stripe at proximal half (Hölzel 1972: fig. 97); male gonocoxite 9 arcuate at distal margin in ventral Clypeus yellow without distinct brownish markings (Fig. 18B); pronotum yellow, medially with two longitudinal dark brown stripes (Fig. 18C); male gonocoxite 9 pointed at distal margin in ventral 3. Pronotum predominantly dark brown 4 Pronotum yellow, medially with two longitudinal dark brown stripes, separated by a narrow yellow 4. Pronotum dark brown, medially with two yellow stripes on anterior half at proximal to anterior transverse furrow, laterally with a narrow yellow stripe at proximal half (Iqbal & Yousuf 1992: - Pronotum dark brown, medially with a narrow longitudinal yellow stripe at proximal half and two rounded yellow markings at distal half, laterally with a narrow yellow stripe at proximal 3/3 (Aspöck 5. Pronotum yellow, medially with two longitudinal dark brown stripes, narrowly separated by central Pronotum yellow, medially with a longitudinal dark brown stripe, anterior transverse furrow with medially dark brown stripe, distal margin with two transverse dark brown stripes (Fig. 14C) ......... 6. Vertex black, posteromedially with a pair of longitudinal yellow markings (Fig. 8C)..... Vertex black, posteriorly with four yellow markings, two at medially and two at lateral margins

# *Myrmeleon tenuipennis* Rambur, 1842 Figs 6–11, 20

Myrmeleon tenuipennis Rambur, 1842: 405. Type locality: India (Maharashtra: Mumbai). Myrmeleon fryeri Navás, 1914c: 135. Type locality: Sri Lanka. Myrmeleon bimaculatus Yang, 1999: 149. Type locality: China (Fujian: Nanping). Syn. nov.

<sup>\*</sup> excluding Myrmeleon bore and M. noacki.

# **Diagnosis**

*Myrmeleon tenuipennis* can be distinguished based on the presence of two narrow median longitudinal yellow markings at posteromedially on vertex (Fig. 8C) and with a pair of median dark brown stripes on pronotum, which is narrowly separated by a central yellow line.

#### **Material examined**

PAKISTAN – **Islamabad Capital Territory** • 11  $\Diamond \Diamond$ , 3  $\Diamond \Diamond$ ; Margalla Hills; 33°43′53.76″ N, 73°2′9.96″ E; 562 m a.s.l.; 16 Aug. 2019, Hassan M.A. leg.; CAU • 1  $\Diamond$ ; Faiz Abad; 33°39′50.6154″ N, 73°9′9.36″ E; 480 m a.s.l.; 11 Aug. 2019, Hassan M.A. leg.; CAU. – **Khyber Pakhtunkhwa Province** • 1  $\Diamond$ , 3  $\Diamond \Diamond$ ; District Swat; 35°3′14.7306″ N, 72°33′53.4492″ E; 760 m a.s.l.; 11 Sep. 2019; Fazullah leg.; NIM • 1  $\Diamond$ ; District Haripur, Sarai Saleh; 33°59′07.64″ N, 72°59′20.97″ E; 610 m a.s.l.; 12 Aug. 2019; Hussain R. leg.; PMNH. – **Punjab Province** • 1 $\Diamond$ ; District Faisalabad, UAF; 31°25′46.8048″ N, 73°4′14.3112″ E; 192 m a.s.l.; 28 Aug. 2019; Hassan M.A. leg.; NIM.

CHINA – **Fujian Province** • 1 ♀ (paratype of *Myrmeleon bimaculatus*); Sha County; 27 Aug. 1979; Bangkan Huang leg.; CAU • 3 ♂♂, 3 ♀♀; 6 larvae reared to adults; Longyan, Xinluo District, Mt. Tiangongshan; 15 Mar. 2020; Yuchen Zheng leg.; CAU • 1 ♂; Xiamen, Siming District, Hudietan; 11 Jun. 2021; Yuchen Zheng leg.; CAU. – **Hainan Province** • 1 ♂; Ledong, Jianfengling; 14 Jun. 1983; Maobin Gu leg.; CAU. – **Guangdong Province** • 1 ♂, 1 ♀; Guangzhou, Fanyu District, Sun Yat-sen University; Jun. 1987; Xuanda Zhang leg.; CAU. – **Guangxi Province** • 1 ♂, 3 ♀♀; Nanning; 23 May 1982; Fasheng Li leg.; CAU • 1 ♂; Congzuo, Pingxiang; 10 May 1963; Chikun Yang leg.; CAU.

# **Re-description**

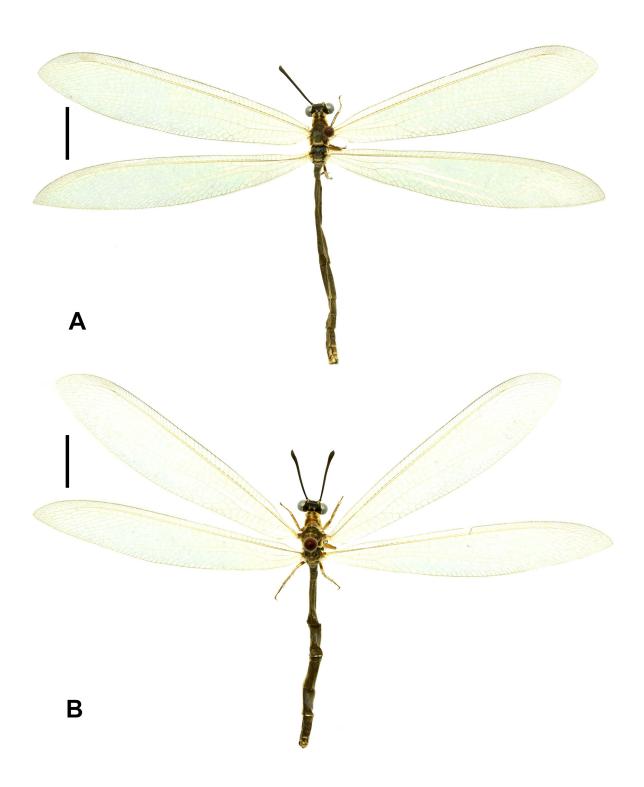
MEASUREMENTS ( $\lozenge$ n=5,  $\lozenge$ n=6). Forewing: length  $\lozenge$  24.5–31.0 mm,  $\lozenge$  25.0–32.5 mm; width  $\lozenge$  5.5–6.8 mm,  $\lozenge$  5.6–6.6 mm; hind wing: length  $\lozenge$  24.5–30.0 mm,  $\lozenge$  27.5–32.0 mm; width  $\lozenge$  4.2–5.4 mm,  $\lozenge$  4.7–5.5 mm; body length:  $\lozenge$  24.5–25.5 mm,  $\lozenge$  22.5–28.0 mm.

HEAD (Fig. 8B–C). Vertex moderately raised; in frontal view black, without yellow markings; dorsally black, posteromedially with two longitudinal yellow markings; epicranial area shining black. Frons shining black, but ventral corner yellow, covered with short brownish pubescence. Occiput shining black. Postorbital sclerite yellow. Clypeus pale yellow, distally with four long black setae. Labrum yellow, covered with erected brownish setae at proximal margin. Genae pale yellow. Maxillary and labial palps pale yellow, terminal labial palpomere spindle-shaped, palpimacula brownish, small and circular, with short black setae. Antennae black, scape mostly yellow, pedicel yellow at proximal ½, flagellum dark brown, covered with short black setae. Antennal sclerite yellow (Fig. 8C–D).

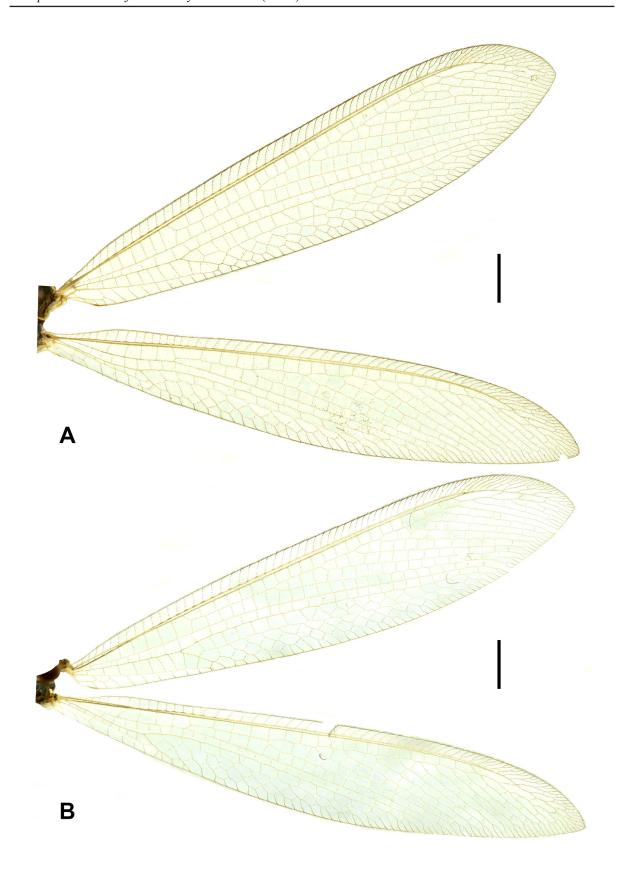
THORAX (Fig. 8A, C). Pronotum slightly wider than long, yellow, with two median longitudinal dark brown stripes, separated by a narrow yellow central stripe, lateral margins yellow, covered with long yellow setae. Mesonotum dark brown, medially with faintly brownish yellow marking, distal margin yellow, covered with sparse yellow setae, but prescutum with long dark brown setae. Metanotum dark brown, medially with faintly brownish yellow markings, distal margin of metascutellum yellow, covered with sparse yellow setae. Pleuron dark brown, covered with sparse yellow setae (Fig. 8E).

Legs (Fig. 8E). Foreleg: coxa and trochanter yellow, covered with short yellow setae. Femur yellow, posterolaterally brownish at apex, covered with short black setae, but posterolaterally with a few long black setae at proximal ½ and ventrally with short yellow setae; femoral sense hair shorter than proximal ½ of profemora. Tibia yellow, anterolaterally brownish, covered with short black setae, posterolaterally with a few long black setae, antennal cleaning setae yellow; tibial spurs brownish, straight, as long as Ta1. Tarsomeres yellow, covered with short black setae; Ta1 equal to combined length of Ta2–Ta4; Ta2, Ta3 and Ta4 nearly equal in size; Ta5 equal to combined length of Ta1–Ta3. Pretarsal claws brownish, moderately

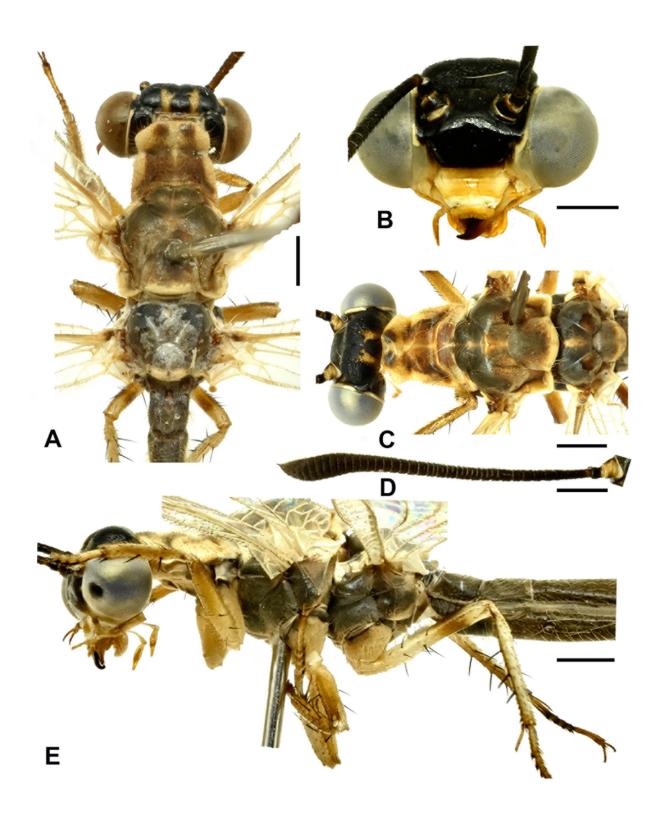
curved. Mid leg: coxa and trochanter similar to foreleg. Femur yellow, anterolaterally brownish, covered with short black setae, ventrally with soft yellow and laterally with a few long black setae at proximal half; femoral sense hair shorter longer than proximal half of mid femora. Tibia similar to foreleg, but laterally



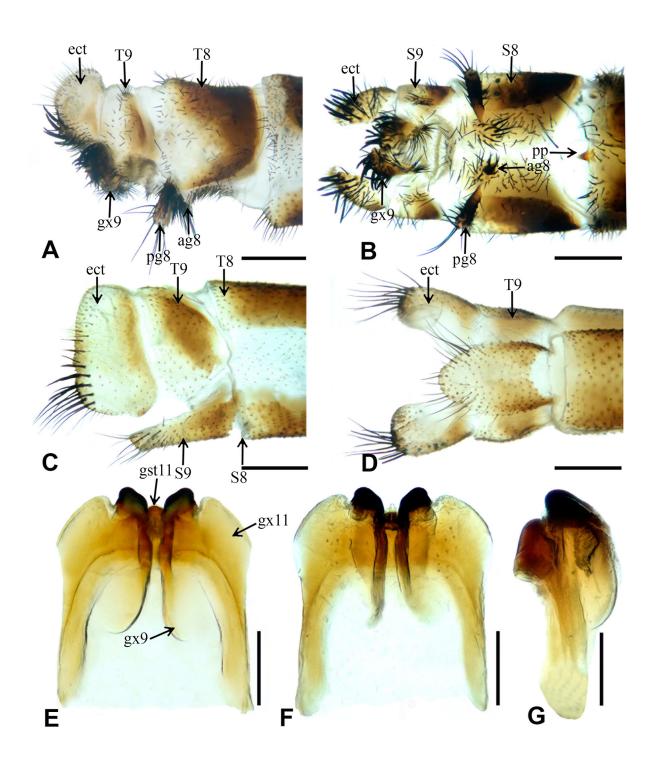
**Fig. 6.** *Myrmeleon tenuipennis* Rambur, 1842. Dorsal habitus. **A**. Male (CAUPK000010). **B**. Female (CAUPK000011). Scale bars = 5.0 mm.



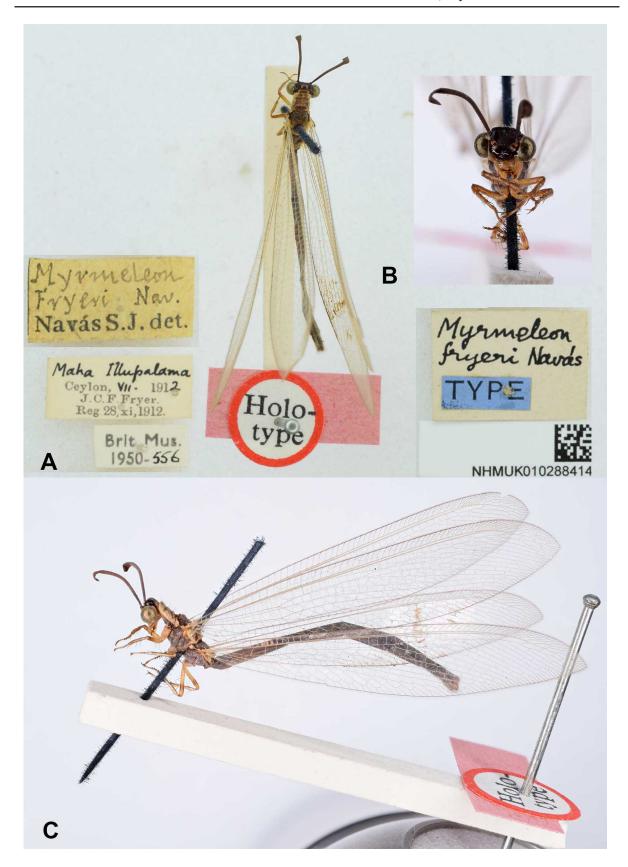
**Fig. 7.** *Myrmeleon tenuipennis* Rambur , 1842. Right fore- and hind wing. **A.** Male (CAUPK000010). **B.** Female (CAUPK000011). Scale bars = 2.0 mm.



**Fig. 8.** *Myrmeleon tenuipennis* Rambur, 1842. **A.** Dorsal habitus of head and thorax. **B.** Head, frontal view. **C.** Head and pronotum, dorsal view. **D.** Antenna. **E.** Lateral view. A–B, D–E:  $\Diamond$  (CAUPK000010); C:  $\Diamond$  (CAUPK000011). Scale bars: A = 2.0 mm; B–D = 1.0 mm; E = 3.0 mm.



**Fig. 9.** *Myrmeleon tenuipennis* Rambur, 1842. **A.** Female genitalia, lateral view. **B.** Same, ventral view. **C.** Male genitalia, lateral view. **D.** Same, ventral view. **E.** Complex of gonocoxites 9 + gonocoxites 11, dorsal view. **F.** Same, ventral view. **G.** Same, lateral view. A–B: ♀ (CAUPK000011); C–G: ♂ (CAUPK000010). Abbreviations: ag8 = anterior gonocoxites 8; ect = ectoproct; gst11 = gonostylus 11; gx9 = gonocoxites 9; gx11 = gonocoxites 11; pg8 = posterior gonocoxites 8; pp = pregenital plate; S = sternites; T = tergites. Scale bars = 0.5 mm.



**Fig. 10.** Holotype of *Myrmeleon fryeri* Navás, 1914 (NHMUK). **A**. Male, dorsal habitus. **B**. Head, frontal view. **C**. Lateral view.

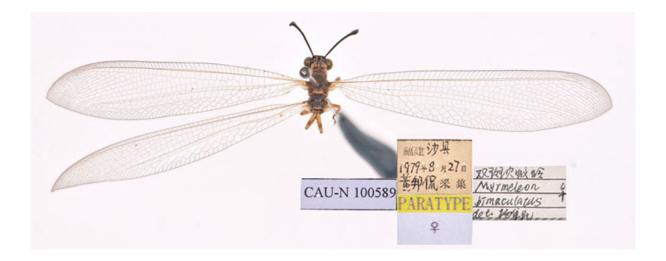
with a few long black setae; tibial spurs similar to foreleg. Tarsomeres and pretarsal claws similar to foreleg. Hind leg: coxa and trochanter similar to mid leg. Femur yellow, with distinct subapical brownish ring, covered with short black setae, but at proximal half with a few long black setae; femoral sense hair absent. Tibia yellow, ventrally brownish, covered with short black setae, ventrally with a row of long black setae; tibial spurs similar to middle leg. Tarsomeres and pretarsal claws similar to mid leg.

Wings (Fig. 7). Forewing: slightly longer and wider than hind wing, subacute at apex; membrane hyaline; costal area slightly narrow at proximal region; venation yellow, except Sc with alternate brownish and yellow patches at proximal half, covered with sparse short black setae; poststigmal area with a few interconnected crossveins; seven to nine presectoral crossveins; initial branching point of CuA at same level or proximal to Rs origin; Rs with 10–13 branches; CuP origin at same level to basal crossveins, fused with 1A after a short free base; pterostigma small, milky white; anterior Banksian line absent; posterior Banksian line present. Hind wing: relatively narrower than forewing, acute at apex; membrane hyaline; venation similar to forewing; four presectoral crossveins; median fork proximal to Rs origin; Rs with 12–14 branches; pterostigma indistinct; anterior Banksian line absent; posterior Banksian line present; pilula axillaris small, with rounded knob, covered with dense brown setae.

ABDOMEN (Fig. 6). Tergites dark brown, but in some specimen with a narrow transverse yellow stripe at distal margin of terga 4–8, covered with short brownish yellow setae. Sternites dark brown, covered with short brownish yellow setae.

MALE GENITALIA (Fig. 9C–G). Tergum 9 trapezoidal, with anterior margin slightly prominent in lateral view. Sternum 9 ovoid in ventral view, covered with elongated black setae at distal half. Ectoproct nearly rectangular in lateral view, covered with yellow setae at distal half, posteroventral corner at proximal half with long thick black setae in lateral view. Gonocoxites 11 highly sclerotized, lateral arms straight, posterolaterally wide and rounded in dorsal view. Gonocoxites 11 slightly prominent in dorsal view. Gonocoxites 9 broad and elongated, proximally diverged in dorsal view, distally rounded in lateral view.

Female Genitalia (Fig. 9A–B). Tergum 9 quadrate in lateral view, covered with short black setae. Ectoproct rounded in lateral view, posterodorsal margin rounded, with short yellow setae, but proximal ½ with robust digging setae. Anterior gonocoxites 8 short, as long as wide, covered with thick long black setae. Posterior gonocoxites 8 long, digitiform, covered with long black setae. Gonocoxites 9 broad and



**Fig. 11.** Paratype of *Myrmeleon bimaculatus* Yang, 1999, ♀ (CAU-N 100589), dorsal habitus.

rounded, covered with robust digging setae, anterolaterally with a bunch of erected short black setae at proximal 1/3. Pregenital plate small, pointed at apex in ventral view.

#### Note

Myrmeleon tenuipennis is rarely mentioned in literature since its original description (Rambur 1842; Ghosh 1983; Stange 2004). Previously, it was only known from India, Sri Lanka, and Vietnam (Stange 2004; Oswald 2020). But we thought this is the most common antlion species in Pakistan, previously misidentified as M. assamensis. The marking patterns on frons and pronotum of M. assamensis reported from Pakistan (Akhtar et al. 2018: fig. 1a) and the paratype of Myrmeleon bimaculatus Yang, 1999 (Fig. 11) from China are almost identical to the type photographs of M. fryeri Navás, 1914 (Fig. 10). However, it can be distinguished from these closely related species by the presence of two yellow markings on vertex in dorsal view (with four yellow markings in M. assamensis: two at median and two at lateral margins in dorsal view). After careful examination of the holotype photographs of M. fryeri (Fig. 10), which is a junior synonym of M. tenuipennis proposed by Esben-Petersen (1931), it is concluded that the specimens presently collected from Pakistan are M. tenuipennis based on the presence of two yellow markings on vertex, instead of four in M. assamensis. We also examined the paratype of M. bimaculatus Yang, 1999 (holotype lost), and confirm that this species is a synonym of M. tenuipennis. Myrmeleon tenuipennis is widely distributed in coastal areas of southern China.

## Distribution

Pakistan: Punjab Province (Islamabad Capital Territory: District Jhelum); China (Fujian, Guangdong, Guangxi, Hainan, Taiwan), India (Maharashtra), Sri Lanka, Vietnam (Ghosh 1983; Yang 1999; Stange 2004; Bao & Wang 2006; Bao *et al.* 2009; Oswald 2020).

*Myrmeleon hyalinus hyalinus* Olivier, 1811 Figs 12–15, 20

Myrmeleon hyalinus hyalinus Olivier, 1811: 126. Type locality: Saudi Arabia.

#### **Diagnosis**

It can be distinguished by the distinctive head and thoracic markings: frons black, except for ventral corner yellow (Fig. 14B); vertex black, median and posterior portions with yellow markings in dorsal view; pronotum yellow, medially with a longitudinal brownish stripe, anterior transverse furrow dark brown, distally with a pair of well-separated transverse brownish stripes (Fig. 14C); wings narrowly elongated, acutely pointed at apex, initial branching point of CuA distad Rs origin (Fig. 13). Moreover, male genitalia is distinctive among species of *Myrmeleon* in Pakistan: gonocoxites 11 highly sclerotized, lateral arms elongated; gonostylus 11 rounded in lateral view; gonocoxites 9 narrow and elongated, wider in lateral view with pointed apex.

#### **Material examined**

PAKISTAN – **Islamabad Capital Territory** • 1 ♂; Quaid-e-Azam University, Shahdarah; 33°45′5.1474″ N, 73°9′40.1754″ E; 555 m a.sl.; 24 Aug. 2019; Hassan M.A. leg.; CAU.

# **Re-description**

MEASUREMENT ( $\Im n=1$ ). Forewing: length 26.2 mm, width 5.5 mm; hind wing: length 26.2 mm, width 4.6 mm; body length: 24.0 mm.

HEAD (Fig. 14B–C). Vertex moderately raised; in frontal view black, without yellow markings; in dorsal view black, medially with a pair of transverse and posteriorly with a pair of longitudinal yellow markings;

epicranial area black, with longitudinal grooves, covered with short brownish pubescence. Frons black, but ventral corner yellow, covered with short yellowish pubescence. Occiput and postorbital sclerites yellow. Clypeus yellow, medially with two indistinct dark brown markings. Labrum yellow, covered with erected brownish setae at proximal margin. Genae pale yellow. Maxillary and labial palps pale yellow, terminal labial palpomere spindle-shaped, palpimacula brownish, small and circular, with short black setae. Antennae brownish, dorsal ring of scape and pedicel yellow, covered with short black setae, flagellum brownish with proximal and distal margin dark brown. Antennal sclerites yellow (Fig. 14B).

Thorax (Fig. 14A). Pronotum slightly wider than long, yellow, medially with a longitudinal brownish stripe, slightly interrupted at anterior transverse furrow; dark brown stripe along anterior transverse furrow, not reaching at lateral margins; distally with a pair of well-separated transverse brownish stripes; covered with sparse yellow setae. Mesonotum dark brown; prescutum laterally with a narrow yellow stripe; mesoscutum with yellow markings on median and posterolateral margins; mesoscutellum at distal ½ yellow; covered with sparse yellow setae, but prescutum with long dark brown setae. Metanotum dark brown; prescutum medially with faintly brownish longitudinal yellow marking; metascutum distally with a pair of large yellow markings; metascutellum laterally and distally brownish yellow; covered with sparse yellow setae. Pleuron dark brown, with yellow markings, covered with sparse yellow setae (Fig. 14E).

Legs (Fig. 14E). Foreleg: coxa and trochanter yellow, with short yellow setae. Femur yellow, light brownish at distal ½, with short black setae, but posterolaterally with a few long black setae at proximal



**Fig. 12.** *Myrmeleon hyalinus hyalinus* Olivier, 1811 (CAUPK000024). Male, dorsal habitus. Scale bar = 0.5 mm.

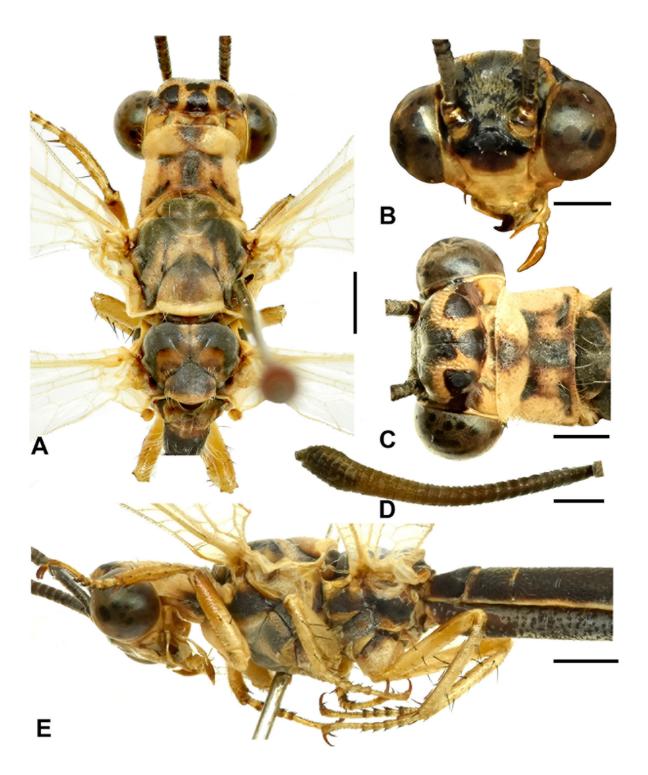
half; femoral sense hair shorter than proximal ½ of profemora. Tibia yellow, with mixed, short and long black setae at proximal half, antennal cleaning setae yellow; tibial spurs brownish, straight, as long as Ta1. Tarsomeres yellow, with short black setae; Ta1 equal to combined length of Ta2–Ta3; Ta2, Ta3 and Ta4 nearly equal in size; Ta5 equal to combined length of Ta1–Ta3. Pretarsal claws brownish, curved. Mid leg: coxa yellow, slightly brownish at proximal ½, with yellow setae. Trochanter yellow, with short black setae. Femur yellow, light brownish at distal ½, covered with short black setae, laterally with a few long black setae at proximal half; femoral sense hair shorter than proximal half of mid femora. Tibia yellow, covered with mixed, short and long black setae; tibial spurs similar to foreleg. Tarsomeres and pretarsal claws similar to foreleg. Hind leg: coxa and trochanter similar to mid leg. Femur yellow, light brownish at distal ⅓, covered with short black setae, proximal half with a few long black setae; femoral sense hair absent. Tibia, tarsomeres, and pretarsal claws are similar to mid leg.

Wings (Fig. 13). Forewing as long as hind wing, acute at apex; membrane hyaline; costal area slightly narrow at proximal region; venation brownish yellow, covered with sparse short black setae; poststigmal



**Fig. 13.** *Myrmeleon hyalinus hyalinus* Olivier, 1811 (CAUPK000024). Right fore- and hind wing. Scale bar = 5.0 mm

area with a few interconnected crossveins; nine presectoral crossveins; initial branching point of CuA proximal to Rs origin; Rs with 11 branches; CuP origin at the same level to basal crossveins, fused with 1A after a short free base; pterostigma indistinct; anterior Banksian line absent; posterior Banksian

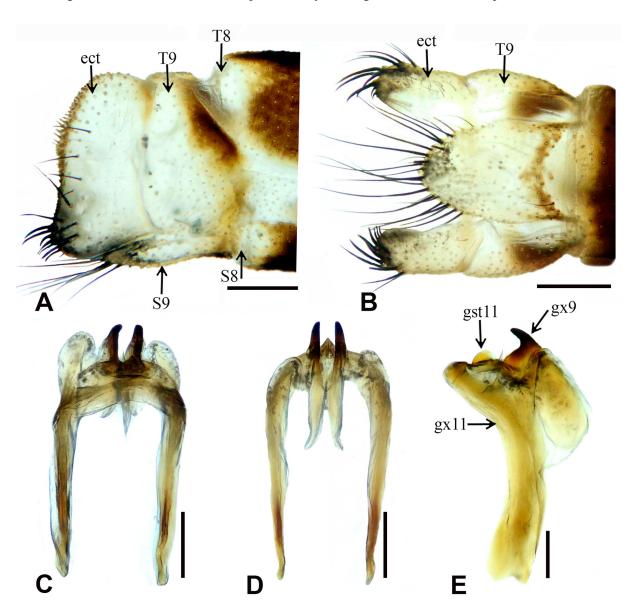


**Fig. 14.** *Myrmeleon hyalinus hyalinus* Olivier, 1811 (CAUPK000024). **A.** Dorsal habitus of head and thorax. **B.** Head, frontal view. **C.** Head and pronotum, dorsal view. **D.** Antenna. **E.** Lateral view. Scale bars: A = 2.0 mm; B - D = 1.0 mm; E = 3.0 mm.

present. Hind wing: relatively narrow, acute at apex; membrane hyaline; venation similar to forewing; five presectoral crossveins; median fork at proximal to Rs origin; Rs with 12 branches; pterostigma indistinct; anterior Banksian line absent; posterior Banksian indistinct; pilula axillaris small, with rounded knob covered with dense brown setae.

ABDOMEN (Fig. 12). Tergites dark brown, distally with a narrow yellow stripe on terga 1–8, lateral margins yellow, covered with short brownish yellow setae. Sternites dark brown, distally with a narrow yellowish stripe, covered with short brownish yellow setae.

MALE GENITALIA (Fig. 15). Tergum 9 subtrapezoidal in lateral view. Sternum 9 ovate-shaped, covered with long black setae at distal ½. Ectoproct nearly rectangular in lateral view, posteroventral corner



**Fig. 15.** *Myrmeleon hyalinus hyalinus* Olivier, 1811 (CAUPK000024). **A.** Male genitalia, lateral view. **B.** Same, ventral view. **C.** Complex of gonocoxites 9 + gonocoxites 11, dorsal view. **D.** Same, ventral view. **E.** Same, lateral view. Abbreviations: ect = ectoproct; gst11 = gonostylus 11; gx9 = gonocoxites 9; gx11 = gonocoxites 11; S = sternites; T = tergites. Scale bars = 0.5 mm.

slightly prominent, covered with long thick black setae. Gonocoxites 11 highly sclerotized, lateral arms elongated, apex broad and curved ventrad in dorsal view. Gonostylus 11 rounded in lateral view. Gonocoxites 9 narrow and elongated, wider in lateral view with pointed apex.

#### Note

Myrmeleon hyalinus currently includes five subspecies: M. h. hyalinus Olivier, 1811 (widespread in Northern Africa and the Middle East, Atlantic islands, India, and Pakistan), M. h. afghanus Hölzel, 1987 (Afghanistan), M. h. caboverdicus Hölzel, 1987 (Cape Verde Islands), M. h. cabrerai Navás, 1912 (Canary Islands), and M. h. distinguendus Rambur, 1842 (widespread in Southern Europe to the Middle East), which can be distinguished in adult morphology and geographical distribution (Hölzel 1987). The morphological characters to delimit the subspecies of M. hyalinus are largely unreliable, particularly the frontal and prothoracic marking patterns (Fig. 14A–C; Hölzel 1987: fig. 2; Akhtar et al. 2018: fig. 3a; Hajiesmaeilian et al. 2020: fig. 11). For example, the marking pattern of frons in our examined specimen is similar to M. h. cabrerai with lower margin of frons narrowly yellow (Fig. 13B; Hölzel 1987: fig. 10), but different from M. h. hyalinus that is distinguished by a median yellow marking on frons at lower margin (Fig. 14B; Hölzel 1987: fig. 8; Akhtar et al. 2018: fig. 7a; Hajiesmaeilian et al. 2020: 20). However, the intraspecific divergence herein observed for M. h. hyalinus and M. h. distinguendus was 0.021-0.049 based on COI gene data. Furthermore, the monophyly of M. h. hyalinus was not recovered based on our analysis (Fig. 21). In conclusion, a combined morphological and molecular data should be applied for all subspecies to resolve the status of these geographically isolated subspecies.

#### **Distribution**

Pakistan: (Punjab Province: District Bahawalpur, Lal Sohanra National Park); widespread: Southern Europe, Northern Africa, Middle East, East to Western India and Macaronesia (Aspöck *et al.* 2001; Stange 2004; Ábrahám 2010, 2011, 2017; Akhtar *et al.* 2018; Hassan *et al.* 2019; Oswald 2020).

# *Myrmeleon trivialis* Gerstaecker, 1885 Figs 16–20

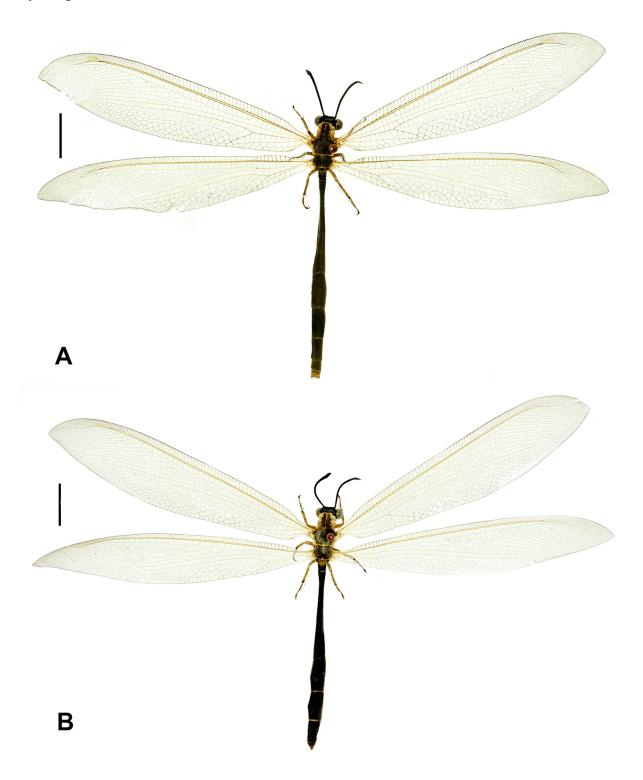
*Myrmeleon trivialis* Gerstaecker, [1885] 1884: 23. Type locality: India (Himalayas). *Myrmeleon montanus* Navás, 1914a. 234. Type locality: India (Darjeeling).

# **Diagnosis**

Body coloration generally dark brown; frons and vertex wholly black, without yellow markings (Fig. 18B–C); pronotum slightly wider than long, mostly yellow, with two broad median dark brown stripes, narrowly separated by a central yellow line; meso- and metanotum dark brown, posteriorly yellow, covered with scattered fine yellowish setae (Fig. 18A).

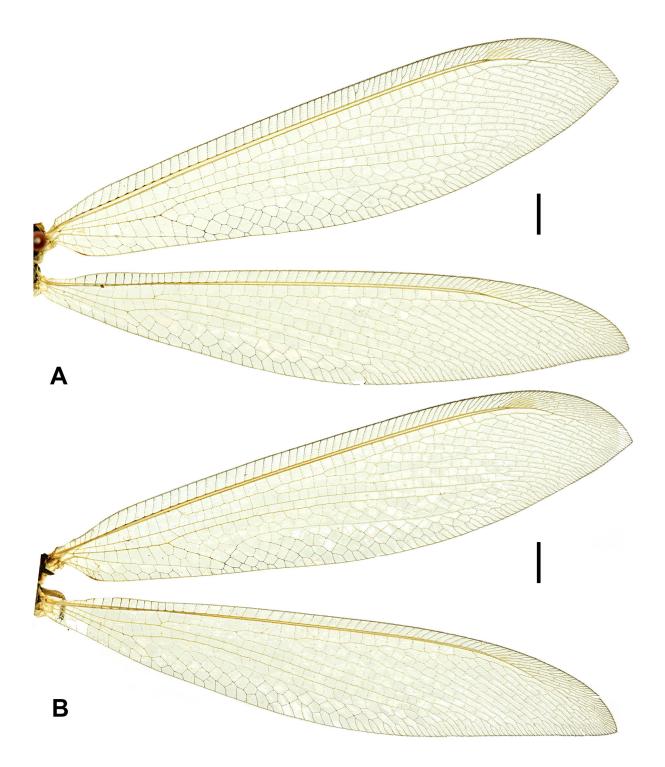
# **Material examined**

• 1  $\,$  Garhi Habibullah; 34°24′23.0394″ N, 73°22′28.1994″ E; 770 m a.s.l.; 22 Aug. 2019; Hassan M.A. leg.; NIM • 1  $\,$  Swat, 35°3′14.7306″ N, 72°33′53.4492″ E; 760 m a.s.l.; 11 Sep. 2019; Fazullah leg.; NIM • 1  $\,$  District Orakzai, Tirah Valley; 33°43′48″ N, 71°0′36″ E; 2300 m a.s.l.; 23 Jun. 2016; Syed leg.; NIM.

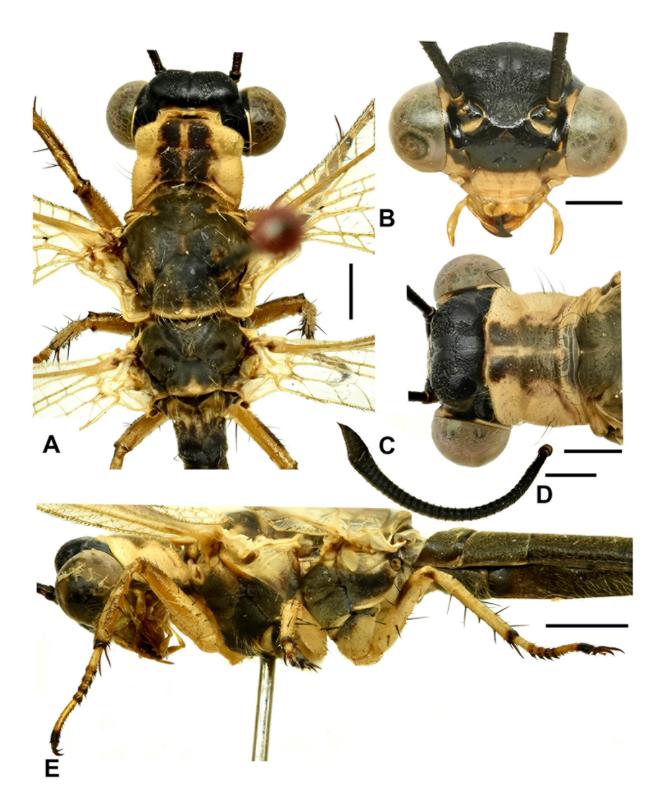


**Fig. 16.** *Myrmeleon trivialis* Gerstaecker, 1885. Dorsal habitus. **A**. Male (CAUPK000025). **B**. Female (CAUPK000026). Scale bars = 5.0 mm.

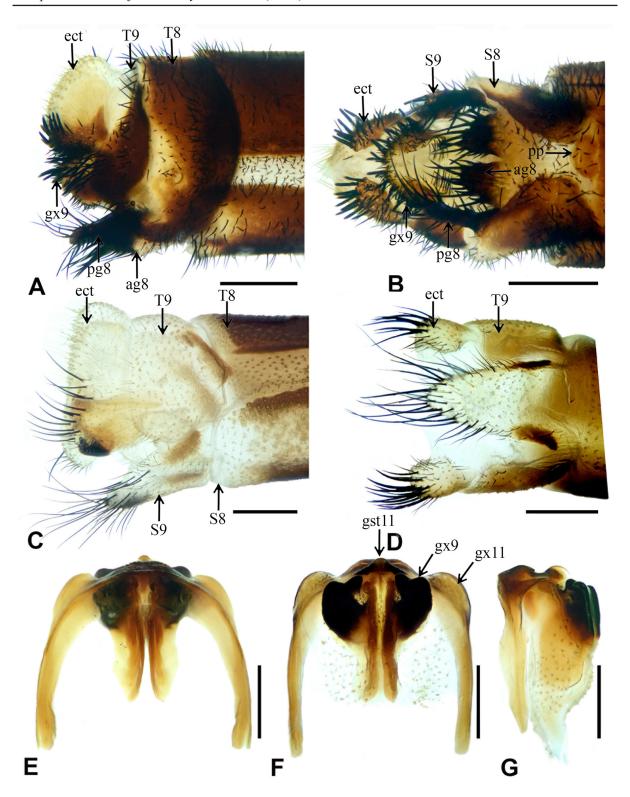
CHINA – **Tibet •** 10 ♂♂, 13 ♀♀; Shigatse, Gyirong County, Gyirong Town; 2650 m a.s.l.; 25 Jun. 2020; Yuchen Zheng leg.; CAU. – **Yunnan Province •** 2 ♂♂; Baoshan, Longyang District, Mangkuan Township, Baihualing Village, Hanlong Camp; 1400 m a.s.l.; 7 Jun. 2020; Yuchen Zheng and Jiazhi Zhang leg.; CAU.



**Fig. 17.** *Myrmeleon trivialis* Gerstaecker, 1885. Right fore- and hind wing. **A.** Male (CAUPK000025). **B.** Female (CAUPK000026). Scale bars = 2.0 mm.



**Fig. 18.** *Myrmeleon trivialis* Gerstaecker, 1885 (CAUPK000025). **A.** Dorsal habitus of head and thorax. **B.** Head, frontal view. **C.** Head and pronotum, dorsal view. **D.** Antenna. **E.** Habitus, lateral view. Scale bars: A = 2.0 mm; B - D = 1.0 mm; E = 3.0 mm.



**Fig. 19.** *Myrmeleon trivialis* Gerstaecker, 1885. **A.** Female genitalia, lateral view. **B.** Same, ventral view. **C.** Male genitalia, lateral view. **D.** Same, ventral view. **E.** Complex of gonocoxites 9 + gonocoxites 11, dorsal view. **F.** Same, ventral view. **G.** Same, lateral view. A-B: 9 + gonocoxites (CAUPK000025). Abbreviations: ag8 = anterior gonocoxites 9 + gonocoxites 21; gx9 = gonocoxites 9 + gonocoxites 32; gx11 = gonocoxites 9 + gonocoxites 33; gx9 = posterior gonocoxites 9 + gonocoxites 34; gx9 = posterior gonocoxites 9 + gonocoxites 35; gx11 = gonocoxites 9 + gonocoxites 36; gx11 = gonocoxites 9 + gonocoxites 37; gx9 = posterior gonocoxites 9 + gonocoxites 38; gx11 = gonocoxites 9 + gonocoxites 39; gx11 = gonocoxites 9 + gonocoxites 31; gx9 = posterior gonocoxites 9 + gonocoxites 31; gx9 = posterior gonocoxites 9 + gonocoxites 31; gx9 = gonocoxites 9 + gonocoxites 31; gx9 = posterior gonocoxites 9 + gonocoxites 31; gx9 = gonocoxites 9 + gonocoxites 31; gx9 = gonocoxites 9 + gonocoxites 31; gx9 = gonocoxites 9 + gonocoxites 32; gx11 = gonocoxites 9 + gonocoxites 31; gx9 = gonocoxites 9 + gonocoxites 32; gx11 = gonocoxites 9 + gonocoxites 32; gx11 = gonocoxites 9 + gonocoxites 33; gx11 = gonocoxites 9 + gonocoxites 34; gx12 = gonocoxites 9 + gonocoxites 35; gx13 = gonocoxites 9 + gonocoxites 36; gx13 = gonocoxites 9 + gonocoxites 36; gx13 = gonocoxites 9 + gonocoxites 37; gx14 = gonocoxites 9 + gonocoxites 38; gx15 = gonocoxites 9 + gonocoxites 39; gx15 = gonocoxites 9 + gonocoxites 30; gx15 = gonocoxites 9 + gonocoxites 30; gx15 = gonocoxites 9 + gonocoxites 30; gx15 = gonocoxites 9 + gonocoxites 31; gx15 = gonocoxites 9 + gonocoxites 31; gx15 = gonocoxi

# **Re-description**

MEASUREMENT ( $\lozenge$ n=4,  $\lozenge$ n=5). Forewing: length  $\lozenge$  36.0–39.0 mm,  $\lozenge$  36.0–43.5 mm; width  $\lozenge$  7.5–9.0 mm,  $\lozenge$  8.2–9.5 mm; hind wing: length  $\lozenge$  35.5–38.5 mm,  $\lozenge$  35.4–42.5 mm; width  $\lozenge$  7.0–7.3 mm,  $\lozenge$  6.3–7.8 mm; body length:  $\lozenge$  32.0–37.0 mm,  $\lozenge$  31.4–38.0 mm.

HEAD (Fig. 18B–C). Vertex moderately raised; in frontal and dorsal view black, without yellow markings; epicranial area with longitudinal grooves, covered with short yellowish pubescence. Frons shining black, with short yellowish pubescence. Occiput shining black. Postorbital sclerite yellow. Clypeus yellow, with four long black setae on proximal margin. Labrum yellow, covered with erected brownish setae at proximal margin. Genae pale yellow. Maxillary and labial palps pale yellow, terminal labial palpomere spindle-shaped, palpimacula brownish, small and circular, with short black setae. Antennae black, with distal ring of scape and pedicel yellow, flagellum dark brown with several distal flagellomeres pointed at apex, covered with short black setae. Antennal sclerites yellow (Fig. 18D).

THORAX (Fig. 18A). Pronotum slightly wider than long, yellow, medially with pair of longitudinal dark brown stripes, separated by a narrow central yellow line, covered with short sparse yellow setae, anterolaterally with short black setae, posterolaterally and distally with a few long black setae. Mesonotum dark brown, covered with sparse yellow setae; posterolateral margins on pre- and mesoscutum with yellow markings, prescutum covered with long dark brown setae; metascutellum distally with a narrow yellow stripe, covered with sparse yellow setae. Metanotum dark brown, metascutellum distally with a narrow yellow stripe, covered with sparse yellow setae. Pleuron dark brown, covered with sparse yellow setae (Fig. 18E).

Legs (Fig. 18E). Foreleg: coxa yellow, slightly brownish at proximal 1/3, with short yellow setae. Trochanter yellow, covered with mixed, short, black and yellow setae. Femur yellow, posterolaterally brownish at apex, covered with short black setae, proximal half with a few long black setae, proximal 1/3 with short yellow setae; femoral sense hair shorter than proximal half of profemora. Tibia yellow, laterally brownish in some specimens, distally shiny black, with short black setae, posterolaterally with a few long black setae, antennal cleaning setae yellow; tibial spurs brownish, straight, as long as Ta1. Tarsomeres yellow, distally each tarsomere dark brown, with short black setae; Ta1 longer than Ta2; Ta2, Ta3 and Ta4 nearly equal in size; Ta5 nearly equal to combined length of Ta1-Ta4. Pretarsal claws brownish, moderately curved. Mid leg: coxa and trochanter similar to foreleg. Femur yellow, anterolaterally dark brown, covered with short black setae, proximal 1/3 with short yellow setae but proximal half with a few long black setae; femoral sense hair shorter than proximal half of mid femora. Tibia yellow, anterolaterally brownish, distally shiny black, covered with short and long black setae; tibial spurs, tarsomeres and pretarsal claws similar to foreleg. Hind leg: coxa and trochanter similar to mid leg. Femur yellow, anterolaterally at distal 1/3 brownish, covered with long black setae, but proximal half with short yellow setae; femoral sense hair absent. Tibia, tibial spurs, tarsomeres, and pretarsal claws similar to mid leg.

Wings (Fig. 17). Forewing: relatively broad, slightly longer than hind wing, subacute at apex; membrane hyaline; costal area slightly narrow at proximal region; longitudinal veins yellow, except Sc and Cu at proximal half with alternate dark brown and yellow patches; crossveins yellow, except cubital area after posterior Banksian line with crossveins black; six to nine presectoral crossveins; initial branching point of CuA proximal to Rs origin; Rs with 11–14 branches; pterostigma small, milky white; anterior Banksian line indistinct as compared to more prominent posterior Banksian line which is proximally brownish black. Hind wing: relatively narrower than forewing, acute at apex; membrane hyaline; longitudinal veins yellow except Sc at proximal half with alternate dark brown and yellow patches; crossveins yellow, but median area after posterior Banksian line with crossveins black; five to six presectoral crossveins; median fork proximal to Rs origin; Rs with 12–14 branches; pterostigma milky

white; anterior Banksian line absent; posterior Banksian line indistinct; pilula axillaris with rounded knob, covered with dense brown setae.

ABDOMEN (Fig. 16). Tergites dark brown, distally with a narrow yellow stripe on terga 4–8; terga 4–7 relatively broader in both sexes; covered with short yellowish setae, but posterior and posterolateral margins of terga 6–7 with mixed, short, black and brownish setae, tergum 8 with short black setae. Sternites dark brown, sterna 7–8 each with distally a narrow yellow stripe; covered with short yellowish setae.

MALE GENITALIA (Fig. 19C–G). Tergum 9 trapezoidal, with anterior margin slightly prominent in middle in lateral view. Sternum 9 triangular in ventral view, covered with long and elongated black setae. Ectoproct nearly rectangular in lateral view, posterodorsal margin rounded, covered with yellow setae, posteroventral corner slightly prominent, covered with long thick black setae. Gonocoxites 11 highly sclerotized, lateral arms straight, rounded at apex in ventral view, apex curved ventrad in dorsal view. Gonostylus 11 cone-shaped, prominent in ventral view. Gonocoxites 9 separated, narrow and straight proximally, wide and curved distally with pointed apex in lateral view.

Female Genitalia (Fig. 19A–B). Tergum 9 subquadrate, covered with short black setae. Ectoproct rounded in lateral view, distal ½ yellow with short yellow setae, proximal ½ dark brown with robust digging setae. Anterior gonocoxites 8 short, as long as wide, covered with thick long black setae. Posterior gonocoxites 8 long, digitiform, covered with elongated black setae. Gonocoxites 9 broad and

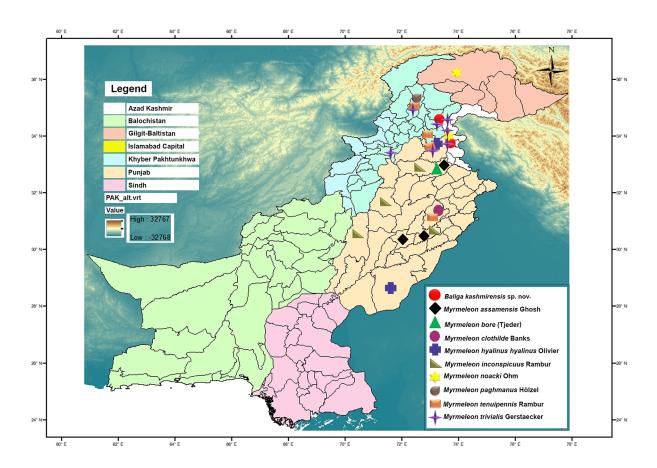
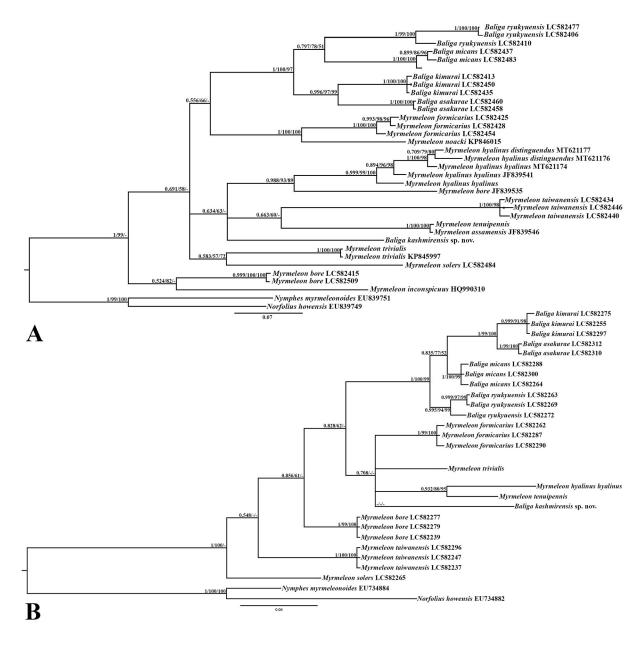


Fig. 20. Distribution map of species of *Baliga* Navás, 1912 and *Myrmeleon* Linnaeus, 1767 from Pakistan.

rounded, covered with robust digging setae, anterolaterally with a bunch of erected short black setae at proximal ½. Pregenital plate small, heart-shaped.

#### Distribution

Pakistan: Khyber Pakhtunkhwa Province (District Mansehra, Garhi Habib Ullah), Azad Kashmir (District Muzaffarabad, Peer Chanasi), Punjab Province (District Rawalpindi, Murree); China (Guangxi, Guizhou, Henan, Shaanxi, Tibet, Yunnan), India (Arunachal Pradesh, Darjeeling, Himachal Pradesh, Sikkim, West Bengal), Nepal, Thailand, Vietnam (Ghosh 1984, 2000; Zhan *et al.* 2011; Akhtar *et al.* 2018; Yang *et al.* 2018; Hassan *et al.* 2019; Ábrahám & Giacomino 2020; Oswald 2020).



**Fig. 21.** Phylogenetic relationships among the species of *Baliga* Navás, 1912 and *Myrmeleon* Linnaeus, 1767 from Pakistan, China, Egypt, Greece, and Japan, based on COI (A) and 16S rRNA (B) genes. The numbers at each node show the posterior probabilities and bootstrap values more than 50% in BI/ML/NJ analysis.

## Myrmeleon assamensis Ghosh, 1984

Myrmeleon assamensis Ghosh, 1984: 23. Type locality: India (Assam).

#### Diagnosis

Myrmeleon assamensis can be distinguished by the marking patterns on vertex and pronotum: vertex black, posteriorly with four yellow markings, two at middle and two at lateral margins in dorsal view; pronotum yellow, medially with a pair of dark brown stripes, which are separated by a narrow median yellow line.

## Note

In the original description (solely based on male), Ghosh (1984) characterized this species based on the presence of two black stripes on the pronotum and the number of presectoral crossveins and the radial branches in both wings. Subsequently, Ghosh (1990) described the female of this species and found that the number of presectoral crossveins and the radial branches are different from that in the male previously described by him. Later, Iqbal & Yousuf (1992: fig. 2) re-described this species based on specimens from Pakistan and provided the line drawings of the head and pronotum in dorsal view as well as the male genitalia. Recently, Akhtar *et al.* (2018: fig. 1a) also recorded this species in Pakistan, which, however, is a case of misidentification of *M. tenuipennis* (see Note to this species). In the original description of *Myrmeleon assamensis*, the marking patterns on vertex are as follows: vertex at distally with two longitudinal and two transverse dark spots in dorsal view (vertex with only two longitudinal yellow markings in *M. tenuipennis*). No new specimens of this species were found in this study.

#### Distribution

Pakistan: Punjab Province (District Jhelum, District Khanewal, District Sahiwal); India (Assam State) (Ghosh 1984, 1992, 2000; Iqbal & Yousuf 1992, 1997; Stange 2004; Akhtar *et al.* 2018; Hassan *et al.* 2019; Oswald 2020).

# Myrmeleon bore (Tjeder, 1941)

Grocus bore Tjeder, 1941: 74. Type localities: Sweden and Norway.

Myrmeleon exigus Yang, 1999: 148. Type locality: China (Fujian: Dongshan).

Myrmeleon tschernovi Krivokhatsky & Shapoval, 2014: 173. Type locality: Russia.

#### **Diagnosis**

Myrmeleon bore can be characterized by wholly dark brown vertex without yellow markings and the pronotum with lateral margins narrowly yellow on proximal half (Aspöck *et al.* 1980: fig. 822; Ábrahám & Papp 1991: fig. 2; Monserrat & Acevedo 2013: fig. 39; Tillier *et al.* 2013: fig. 13; Ábrahám & Giacomino 2020: fig. 3), while the species recorded from Pakistan have yellow markings on a vertex in dorsal view and the pronotum medially brownish yellow (Akhtar *et al.* 2018: fig. 2a). This character suggests that the specimens from Pakistan identified as Myrmelon bore belong to a different species.

#### Remarks

Myrmeleon bore seems widely distributed in the Palaearctic Region and was recently recorded from Pakistan. Notably, Enza otiosus Navás, 1912 has long been considered a secondary synonym of M. bore (Stange 2004; Sekimoto 2014; Wang et al. 2018). However, based on the priority of the nomenclature of the ICZN (International Code of Zoological Nomenclature), the validity of E. otiosus should be restored and transferred to Myrmeleon, then M. bore should be treated as a synonym of the former. Meanwhile, Kuwayama (1962) did not treat E. otiosus as a synonym for M. bore formally. Considering the type

localities between *E. otiosus* (holotype in Japan) and *M. bore* (syntypes in Sweden and Norway) have great distance, the relationship between both species needs to be further investigated. Hence, we do not include *E. otiosus* in the citation of *M. bore*.

#### Distribution

Pakistan? Widespread in Palaearctic Region (Röhricht 1998; Aspöck et al. 2001; Bao & Wang 2006; Akhtar et al. 2018; Yang et al. 2018; Hassan et al. 2019; Ábrahám & Giacomino 2020; Oswald 2020).

# Myrmeleon clothilde Banks, 1913

Myrmeleon clothilde Banks, 1913: 223. Type locality: India (Bihar: Samastipur, Pusa).

#### **Diagnosis**

Myrmeleon clothilde can be characterized by wholly dark brown vertex with yellow markings at distal half in dorsal view; pronotum dark brown, laterally slightly narrow yellow at proximal half; medially with two narrow longitudinal yellow stripes at proximal to anterior transverse furrow (Iqbal & Yousuf 1992: fig. 1a).

#### Note

Since its original description, this species was rarely mentioned in literature (Iqbal & Yousuf 1992, 1997; Ghosh 2000; Stange 2004). After reviewing the aforementioned literature from Pakistan and India, we found that the male genitalia of this species have not been described so far. However, further additional data on the male genitalia and distribution of this rarely known species in Pakistan need to be updated in further studies. No new specimens of this species were found in this study.

#### Distribution

Pakistan: Punjab Province (District Faisalabad); India, Sri Lanka (Iqbal & Yousuf 1992, 1997; Ghosh 2000; Stange 2004; Hassan *et al.* 2019; Oswald 2020).

# Myrmeleon inconspicuus Rambur, 1842

Myrmeleon inconspicuus Rambur, 1842: 406. Type locality: unknown.

Myrmeleon incertus Rambur, 1842: 406. Type locality: probably from Southern France.

Myrmeleon erberi Brauer, 1868: 190. Type locality: unknown.

Myrmeleon ariasi Navás, 1913b: 114. Type locality: Morocco.

Myrmeleon inconspicuus leoninus Navás, 1912d: 30. Type locality: unknown.

#### **Diagnosis**

*Myrmeleon inconspicuus* can be characterized by a wholly dark black vertex with yellow markings at distal half in dorsal view; pronotum dark brown with a narrow median longitudinal yellow marking at proximal to anterior transverse furrow and two rounded yellow markings at distal half (Akhtar *et al.* 2018: fig. 4a; Hajiesmaeilian *et al.* 2020: figs 11–12). No new specimens of this species were found in this study.

## **Distribution**

Pakistan: Punjab Province (District Chakwal, Lal Sunahara National Park, District Bhakkar, Darya Khan, Mithi, District Dera Ghazi Khan, Chak Talpur, District Sahiwal, Harappa); Southern Europe (widespread), Northern Africa (widespread), Middle East to Iran (Aspöck *et al.* 1980, 2001; Akhtar *et al.* 2018; Hassan *et al.* 2019; Hajiesmaeilian *et al.* 2020; Oswald 2020).

# Myrmeleon noacki Ohm, 1965

Myrmeleon noacki Ohm, 1965: 108. Type locality: Greece (West Greece: Zachlorou).

#### **Diagnosis**

*Myrmeleon noacki* can be characterized by a wholly dark brown vertex, without yellow markings; pronotum dark brown, laterally yellow, medially with a narrow longitudinal yellow line at proximal half wings lack pipula axillaris in males; apex of male gonocoxites 9 narrowly arcuated at anterolateral margins in ventral view.

#### Note

Myrmeleon noacki is known from the southeastern parts of Europe to Turkey, and was recently reported from Iran and Pakistan (Akhtar et al. 2018; Hajiesmaeilian et al. 2020). The reports of Myrmelon noacki from Pakistan need to be re-evaluated and compared with European specimens. Typically, this species is characterized by a narrow median longitudinal yellow marking at proximal half of pronotum and the male genitalia with gonocoxites 9 at anterolateral margins arcuated at anterolateral margins at apex in ventral view (see Ohm 1965: figs 2, 6; Hajiesmaeilian et al. 2020: figs 10, 14). The prothoracic markings and the shape of male genitalia of this species recorded from Pakistan match the typical diagnosis for M. paghmanus: pronotum dark brown, medially with a narrow longitudinal yellow marking, rounded at middle; male gonocoxites 9 arcuate at distal margin in ventral view (see Akhtar et al. 2018: fig. 5a–b; Hölzel 1972: figs 97, 101–102). No new specimens of this species were found in this study.

#### Distribution

Pakistan: Azad Kashmir (District Bagh); Gilgit-Baltistan (District Gilgit); Bulgaria, Greece, Macedonia, Iran, Turkey (Aspöck *et al.* 2001; Pantaleoni & Badano 2012; Akhtar *et al.* 2018; Hassan *et al.* 2019; Hajiesmaeilian *et al.* 2020; Oswald 2020).

# Myrmeleon paghmanus Hölzel, 1972

*Myrmeleon paghmanus* Hölzel, 1972: 37. Type locality: Pakistan (Khyber Pakhtunkhwa: Swat; Gabral-Tal).

#### **Diagnosis**

*Myrmeleon paghmanus* can be characterized by the combination of the following characters: clypeus yellow, medially with two rounded brownish markings; pronotum dark brown, medially with a narrow longitudinal complete yellow stripe, laterally with a narrow yellow stripe at proximal half wings lack pipula axillaris in males; male gonocoxites 9 arcuate at distal margin in ventral view.

#### Note

Up till now, this species is only known from Afghanistan and Pakistan. No new specimens of this species were found in this study.

# **Distribution**

Pakistan: Khyber Pakhtunkhwa Province (District Swat, Gabral-Tal); Afghanistan (Hölzel 1972; Stange 2004; Hassan *et al.* 2019; Oswald 2020).

## Molecular identification

The present phylogenetic analysis based on COI and 16S rRNA genes shows that there is strong support for the monophyly of *Baliga* clade for Japanese species by BI, ML, and NJ methods, which, however, did not

comprise *Baliga kashmirensis* sp. nov. from Pakistan. Based on COI genes, *Baliga kashmirensis* sp. nov. was assigned to be within a monophylum with *M. tenuipennis* and *M. taiwanensis* Miller & Stange, 1999. However, the monophyly of *Baliga kashmirensis* sp. nov. with *M. tenuipennis*, *M. hyalinus*, *M. trivialis*, and the *M. formicarius* clade is recovered with relatively low nodal support values. For now, the present phylogenetic analysis is largely focused on species identification due to incomplete taxon sampling. The genetic divergence between *B. kashmirensis* sp. nov. and the species of *Myrmeleon* was 0.139–0.188 and that between this new species and the other species of *Baliga* was 0.153–0.186. The greatest intraspecific divergence (0.049) was found respectively in *B. ryukyuensis* Hayashi & Matsumoto, 2020 and *M. hyalinus*. The minimum and maximum interspecific genetic divergence between species of *Baliga* and *Myrmeleon* ranged from 0.074–0.186, and 0.123–0.188, respectively.

## **Discussion**

In most recent taxonomic and molecular studies on Myrmeleontini from Japan, Hayashi et al. (2020) considered Baliga as a valid genus based on the generic classification system proposed by Stange (2004). According to Stange (2004), Baliga can be characterized by the presence of interconnected crossveins proximal to pterostigma in the forewing and the anterior gonocoxites 8 shorter than the posterior gonocoxites 8 in the female genitalia. However, the BI, ML and NJ trees herein reconstructed respectively based on COI and 16S rRNA genes for six species of Baliga and 11 Myrmeleon (Table 1; Fig. 21) albeit receiving low supports at deep-level nodes recovered Myrmeleon as paraphyletic. Similar results were also recovered in the recent molecular phylogenetic studies by Michel et al. (2017) and Machado et al. (2019). In Hayashi et al. (2020), the species of Baliga and those of Myrmeleon, respectively, constituted a monophylum, which, however, might be due to incomplete taxon sampling. Nevertheless, here we do not propose any new generic synonym but still follow the generic classification system of Stange (2004). The validity of Baliga needs a major phylogenetic revision with larger datasets in future studies. Furthermore, the additional proposed synonymized genera by Machado et al. (2019) need to be deciphered in future studies. In addition, concerning the great intraspecific divergence, it is necessary to clarify the taxonomic status of all subspecies of M. hyalinus and the two insular populations of B. ryukyuensis (Amami/Tokunoshima and Okinawa) based on both morphological and molecular data.

With respect to the distribution of antlions in Pakistan, our results corroborate the mixed fauna from the Oriental and Palaearctic regions in the northern parts of Pakistan (Fig. 20) due to the unique geographical position – the extreme edge of western Himalayas and the junction point of the world's two largest zoogeographical regions: the Oriental and the Palaearctic. The present diversity and distribution of antlions in Pakistan is consistent with our recent studies on the subfamily Ascalaphinae (Neuroptera: Myrmeleontidae) and the following genera of Myrmeleontidae and Megaloptera from northern Pakistan: *Distoleon* Banks, 1910 (Neuroptera: Myrmeleontidae), *Nevromus* Rambur, 1842 and *Protohermes* van der Weele, 1907 (Megaloptera: Corydalidae) (Hassan *et al.* 2019, 2020a, 2020b; Hassan & Liu 2021). Nevertheless, this pattern may be also due to the present limitation of sampling (extensive collecting primarily confined to the northern parts of Pakistan). Broader sampling across the country, particularly for the southern parts, may reveal the true diversity and distribution of antlions in the future.

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## References

Ábrahám L. 2010. Short report on the fauna of ant-lion and owl-fly (Neuroptera) from Socotra Archipelago. *Natura Somogyiensis* 17: 177–191.

Ábrahám L. 2011. Further data to the ant-lion fauna (Neuroptera) of Socotra Island (Yemen). *Natura Somogyiensis* 19: 101–108.

Ábrahám L. 2017. New data to the Moroccan Myrmeleontiformia (Nemopteridae, Myrmeleontidae, Ascalaphidae) fauna. *Natura Somogyiensis* 30: 75–138.

Ábrahám L. & Giacomino M. 2020. A little known and synonym ant-lions 2 (Neuroptera: Myrmeleontidae). *Natura Somogyiensis* 34: 21–72. https://doi.org/10.24394/NatSom.2020.34.21

Ábrahám L. & Papp Z. 1991. *Myrmeleon bore* (Tjeder, 1941) in Hungary (Planipennia, Myrmeleontidae). *Neuroptera International* 6: 137–139.

Akhtar S. 2018. *DNA Based Identification and Genetic Diversity Studies of Antlion Species of Pakistan*. PhD dissertation, University of Agriculture, Peshawar, Pakistan.

Akhtar S., Ashfaq M., Zia A., Ali S., Ali G.M., Farhatullah & Zafar Y. 2018. First report and redescription of five species of genus *Myrmeleon* (Neuroptera: Myrmeleontidae) from Pakistan. *Journal of Biodiversity and Environmental Sciences* 13: 180–190.

Aspöck, U. & Aspöck, H. 2008. Phylogenetic relevance of the genital sclerites of *Neuropterida* (Insecta: Holometabola). *Systematic Entomology* 33: 97–129. https://doi.org/10.1111/j.1365-3113.2007.00396.x

Aspöck H., Aspöck U. & Hölzel H. 1980. Die Neuropteren Europas. Eine Zusammenfassende Darstellung der Systematik, Ökologie und Chorologie der Neuropteroidea (Megaloptera, Raphidioptera, Planipennia) Europas. 2 Volumes. Goecke & Evers, Krefeld, Germany.

Aspöck H., Hölzel H. & Aspöck U. 2001. Kommentierter Katalog der Neuropterida (Insecta: Raphidioptera, Megaloptera, Neuroptera) der Westpaläarktis. *Denisia* 2: 1–606.

Badano D. & Pantaleoni R.A. 2014. The larvae of European Myrmeleontidae (Neuroptera). *Zootaxa* 3762: 1–71. https://doi.org/10.11646/zootaxa.3762.1.1

Badano D., Aspöck H. & Aspöck U. 2017a. Taxonomy and phylogeny of the genera *Gymnocnemia* Schneider, 1845, and *Megistopus* Rambur, 1842, with remarks on the systematization of the tribe Nemoleontini (Neuroptera, Myrmeleontidae). *Deutsche Entomologische Zeitschrift, Berlin* 64: 43–60. https://doi.org/10.3897/dez.64.11704

Badano D., Aspöck U., Aspöck H. & Cerretti P. 2017b. Phylogeny of Myrmeleontiformia based on larval morphology (Neuropterida: Neuroptera). *Systematic Entomology* 42: 94–117. https://doi.org/10.1111/syen.12200

Badano D., Aspöck H., Aspöck U. & Haring E. 2017c. Eyes in the dark ... shedding light on the antlion phylogeny and the enigmatic genus *Pseudimares* Kimmins (Neuropterida: Neuroptera: Myrmeleontidae). *Arthropod Systematics & Phylogeny* 75: 535–554.

Badano D., Miller R. & Stange L.A. 2018. Rediscovery and revision of the antlion genus *Ripalda* Navás within a phylogeny of Nemoleontini (Neuroptera, Myrmeleontidae). *Invertebrate Systematics* 32: 933–949. https://doi.org/10.1071/IS18022

Banks N. 1909. New genera and species of tropical Myrmeleonidae. *Journal of the New York Entomological Society* 17: 1–4.

Banks N. 1913. Synopses and descriptions of exotic Neuroptera. *Transactions of the American Entomological Society* 39: 201–242.

Bao R. & Wang X.L. 2006. Two new species of *Myrmeleon* Linnaeus, 1767 (Neuroptera: Myrmeleontidae) from China, with a key to Chinese species. *Proceedings of the Entomological Society of Washington* 108: 125–130.

Bao R., Shen Z.R. & Wang X.L. 2007. A review of the species of *Hagenomyia* from China (Neuroptera: Myrmeleontidae). *Annales de la Société Entomologique de France (N.S.)* 43: 45–48. https://doi.org/10.1080/00379271.2007.10697492

Bao R., Wang X.L. & Liu J.Z. 2009. A review of the species of *Myrmeleon* Linnaeus, 1767 (Neuroptera: Myrmeleontidae) from mainland China, with the description of a new species. *Entomological News, Philadelphia* 120: 18–24. https://doi.org/10.3157/021.120.0108

Brauer F. 1868. Zwei neue Myrmeleon-Arten. Verhandlungen der Kaiserlich-Königlichen Zoologisch-Botanischen Gesellschaft in Wien 18: 189–190.

Brullé G.A. 1839. Néoroptéres. *In*: Webb P.B. & Berthelot S. (eds) *Histoire Naturelle des Iles Canaries*. *Tome 2. Part 2*. Béthune, Paris.

De Geer C. 1773. Des Fourmilions exotiques. *In: Mémoires pour servir à l'Histoire des Insects. Vol. 3*. Stockholm, Pierre Hesselberg.

Esben-Petersen P. 1913. H. Sauter's Formosa-Ausbeute. Planipennia II, Megaloptera und Mecoptera. *Entomologische Mitteilungen* 2: 222–228, 257–265.

Esben-Petersen P. 1918. Results of Dr. E. Mjöberg's Swedish Scientific Expeditions to Australia 1910–1913. 18. Neuroptera and Mecoptera. *Arkiv för Zoologi* 11: 1–37.

Esben-Petersen P. 1931. Myrmeleontiden aus Süd-Indien. *Revue Suisse de Zoologie* 38: 445–448. https://doi.org/10.5962/bhl.part.117649

Gerstaecker A. 1884 [1885]. Vier Decaden von Neuropteren aus der Familie Megaloptera Burm. *Mitt[h] eilungen aus dem Naturwissenschaftlichen Verein für Neu-Vorpommern ünd Rugen* 16: 1–49.

Ghosh S.K. 1983. Notes on the biogeography of Neuroptera: Planipennia from certain areas of the North West Himalayan and Northern Peninsular sectors of India. *Records of the Zoological Survey of India* 80: 291–300.

Ghosh S.K. 1984. Contribution to the taxonomical studies of Neuroptera (Suborder Planipennia) from eastern India. 1. Family Myrmeleontidae. *Records of the Zoological Survey of India, Miscellaneous Publications, Occasional Paper* 52: 1–63.

Ghosh S.K. 1990. Description of an unknown female along with two new locality records of the family Myrmeleontidae (Neuroptera) from Lakshadweep Islands, India. *Records of the Zoological Survey of India* 87: 259–261.

Ghosh S.K. 2000. Neuroptera fauna of North-East India. *Records of the Zoological Survey of India, Occasional Paper*: 184, i–xiii + 1–179.

Hajiesmaeilian A., Shoushtari R.V., Mozaffarian F. & Ebrahimi E. 2020. Tribe Myrmeleontini (Neuroptera: Planipennia: Myrmeleontidae) in Iran. *Zootaxa* 4751: 153–160. https://doi.org/10.11646/zootaxa.4751.1.9

Hassan M.A. & Liu X.Y. 2021. Taxonomic notes on owlflies from Pakistan (Neuroptera: Myrmeleontidae: Ascalaphinae). *Zootaxa* 4970: 401–452. https://doi.org/10.11646/zootaxa.4970.3.1

Hassan M.A., Oswald J.D., Zia A. & Liu X.Y. 2019. Neuropterida (Insecta: Megaloptera, Raphidioptera, Neuroptera) of Pakistan: a catalogue and faunistic review. *Zootaxa* 4686: 497–541. https://doi.org/10.11646/zootaxa.4686.4.3

Hassan M.A., Yuchen Z. & Liu X.Y. 2020a. Taxonomic notes on the antlion genus *Distoleon* Banks (Neuroptera: Myrmeleontidae) from Pakistan. *Zootaxa* 4869: 347–368. https://doi.org/10.11646/zootaxa.4869.3.3

Hassan M.A., Hayashi Y. & Liu X.Y. 2020b. First record of the dobsonfly genus *Protohermes* van der Weele, 1907 from Pakistan (Megaloptera: Corydalidae). *Zootaxa* 4732 (3): 422–434. https://doi.org/10.11646/zootaxa.4732.3.5

Hayashi H., Matsumoto R., Sugawara H. & Liu X.Y. 2020. Two new species of *Baliga* (Neuroptera: Myrmeleontidae: Myrmeleontinae) with the molecular phylogeny of the tribe Myrmeleontini in Japan. *Japanese Journal of Systematic Entomology* 26 (2): 235–251.

Hebert P.D.N., Ratnasingham S. & DeWaard J.R. 2003. Barcoding animal life: Cytochrome c oxidase subunit 1 divergences among closely related species. *Proceedings of the Royal Society London B. Biological Sciences* 270: 1–4. https://doi.org/10.1098/rsbl.2003.0025

Hölzel H. 1972. Die Neuropteren Vorderasiens IV. Myrmeleonidae. Beiträge zur Naturkundlichen Forschung in Südwestdeutschland, Beiheft 1: 3–103.

Hölzel H. 1987. *Myrmeleon hyalinus* Olivier – eine chorologisch-taxonomische Analyse (Neuropteroidea: Planipennia: Myrmeleonidae). *Zeitschrift der Arbeitsgemeinschaft Österreichischer Entomologen* 38: 78–88.

Iqbal M. & Yousuf M. 1992. New record and redescriptions of two species of the genus *Myrmeleon* Linnaeus (Myrmeleontidae: Neuroptera) from Punjab (Pakistan). *Pakistan Entomologist* 14: 95–96.

Iqbal M. & Yousuf M. 1997. Antlions (Myrmeleontidae: Neuroptera) of the Punjab, Pakistan. *Pakistan Journal of Zoology* 29: 127–138.

Kimura M. 1980. A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. *Journal of Molecular Evolution* 16: 111–120. https://doi.org/10.1007/BF01731581

Krivokhatsky V.A., Shapoval N.A. & Shapoval A.P. 2014. Муравьиные львы (Neuroptera, Myrmeleontidae) В орнитологических ловушках на Куршской косе: трехвидовое сообщество с новым для науки видом [= Antlions (Neuroptera, Myrmeleontidae) from ornithological traps on the Curonian spit: a three-species community containing a new species]. *Zoological Journal* [= Зоологический Журнал = Zoologicheskii Zhurnal] 93: 171–178. [In Russian.]

Kuwayama S. 1962. A revisional synopsis of the Neuroptera in Japan. *Pacific Insects* 4: 325–412.

Kumar S., Stecher G. & Tamura K. 2016. MEGA7: molecular evolutionary genetics analysis ver. 7.0 for bigger datasets. *Molecular Biology and Evolution* 33: 1870–1874. https://doi.org/10.1093/molbev/msw054

Latreille P.A. 1810. Considérations générales sur l'Ordre naturel des Animaux composant les Classes des crustacés, des arachnides, et des insectes: avec un Tableau méthodique de leurs Genres, disposés en familles. Schoell, Paris. https://doi.org/10.5962/bhl.title.39620

Machado R.J.P. & Oswald J.D. 2020. Morphological phylogeny and taxonomic revision of the former antlion subtribe Periclystina (Neuroptera: Myrmeleontidae: Dendroleontinae). *Zootaxa* 4796: 1–322. https://doi.org/10.11646/zootaxa.4796.1.1

Machado R.J.P., Gillung J.P., Winterton S.L., Garzón-Orduña I.J., Lemmon A.R., Lemmon E.M. & Oswald J.D. 2019. Owlflies are derived antlions: anchored phylogenomics supports a new phylogeny and classification of Myrmeleontidae (Neuroptera). *Systematic Entomology* 44: 418–450. https://doi.org/10.1111/syen.12334

Mansell M.W. 1996. Predation strategies and evolution in antlions (Insecta: Neuroptera: Myrmeleontidae). *In*: Canard M., Aspöck H. & Mansell M.W. (eds) *Pure and Applied Research in Neuropterology. Proceedings of the Fifth International Symposium on Neuropterology (2-6 May 1994, Cairo, Egypt)*. Privately printed, Toulouse, France.

Markl W. 1954. Vergleichend-morphologische Studien zur Systematik und Klassifikation der Myrmeleoniden (Insecta, Neuroptera). *Verhandlungen der Naturforschenden Gesellschaft in Basel* 65: 178–263.

McLachlan R. 1875. A sketch of our present knowledge of the neuropterous fauna of Japan (excluding Odonata and Trichoptera). *Transactions of the [Royal] Entomological Society of London* 23: 167–190. https://doi.org/10.1111/j.1365-2311.1875.tb01906.x

McLachlan R. 1894. Two new species of Myrmeleonidae from Madagascar. *Annals and Magazine of Natural History*, Series 6, 13: 514–517. https://doi.org/10.1080/00222939408677745

Michel B., Clamens A.L., Bérthoux O., Kergoat G.J. & Condamine F.L. 2017. A first higher-level time calibrated phylogeny of antlions (Neuroptera: Myrmeleontidae). *Molecular Phylogenetics and Evolution* 107: 103–116. https://doi.org/10.1016/j.ympev.2016.10.014

Miller M., Pfeiffer W. & Schwartz T. 2010. Creating the CIPRES Science Gateway for inference of large phylogenetic trees. *Proceedings of the Gateway Computing Environments Workshop (GCE)* 14: 1–8. https://doi.org/10.1109/GCE.2010.5676129

Monserrat V.J. & Acevedo F. 2013. Los mirmeleónidos (hormigas-león) de la Península Ibérica e Islas Baleares (Insecta, Neuropterida, Neuroptera, Myrmeleontidae). *Graellsia* 69: 283–321. https://doi.org/10.3989/graellsia.2013.v69.098

Navás L. 1912a. Insectos neurópteros nuevos o poco conocidos. *Memorias de la Real Academia de Ciencias y Artes de Barcelona* 10: 135–202.

Navás L. 1912b. Myrméléonides nouveau de 1'extréme Orient (Neuroptera). *Revue Russe d'Entomologie* 12: 110–114.

Navás L. 1912c. Myrméléonides nouveaux ou peu connus (Ins. Névr.). *Annales de la Societe Scientifique de Bruxelles* 36: 203–248.

Navás L. 1912d. Notas sobre Mirmeleónidos (Ins. Neur.). Brotéria (Zoológica) 10: 29–75, 85–97.

Navás L. 1912e. New Mirmeleónido (Ins. Neur.) From the Canary Islands. *Magazine of the Royal Academy of Exact Physical and Natural Sciences of Madrid* 10: 672–674.

Navás L. 1913a. Bemerkungen über die Neuropteren der Zoologischen Staatssammlung in München. V. *Mitteilungen der Münchener Entomologischen Gesellschaft* 4: 9–15.

Navás L. 1913b. Algunos Neurópteros de Marruecos. *Memorias de la [Real] Sociedad Española de Historia Natural* 8: 111–122.

Navás L. 1914a. Myrméléonides (Ins. Névr.) nouveaux ou critiques. *Annales de la Société Scientifique de Bruxelles* 38: 229–254.

Navás L. 1914b. Neurópteros sudamericanos. Primera serie. Brotéria (Zoológica) 12: 45–56, 215–234.

Navás L. 1914c. Névroptères de l'Indochine. 1<sup>re</sup> série. *Insecta, Rennes* 4: 133–142.

Navás L. 1915a. Neurópteros nuevos o poco conocidos. Quinta [V] Serie. *Memorias de la Real Academia de Ciencias y Artes de Artes Barcelona*, Series 3 (11): 455–480.

Navás L. 1915b. Some Neuroptera from the United States. *Bulletin of the Brooklyn Entomological Society* 10: 50–54.

Navás L. 1916. Neuroptera nova africana. VII series. *Memorie dell' Accademia Pontifica dei Nuovi Lincei*, Series 2, 2: 51–58.

Navás L. 1919. Comunicaciones entomológicas. 3. Insectos exóticos. *Revista de la Academia de Ciencias Exactas Fisico-Quimicas y Naturales de Zaragoza*, Series 1, 4: 287–306.

Navás L. 1920. Sur des Névroptéres nouveaux ou critiques. Deuxiéme (II) série. *Annales de la Société Scientifique de Bruxelles* 39: 189–203.

Navás L. 1919 [1921]. Comunicaciones entomológicas. 4. Insectos exóticos nuevos, críticos o poco conocidos. *Revista de la Academia de Ciencias Exactas Fisico-Quimicas y Naturales de Zaragoza*, Series 1, 6: 61–81.

Navás L. 1923a. Insecta orientalia. I Series. *Memorie dell'Accademia Pontifica dei Nuovi Lincei, Rome*, Series 2, 6: 29–41.

Navás L. 1923b. Quelques Myrméléonides (Ins. Névr.) d'Afrique. *Annales de la Société Scientifique de Bruxelles* 43: 143–147.

Navás L. 1925. Insectos exóticos nuevos o poco conocidos. Segunda [II] serie. Memorias de la Real Academia de Ciencias y Artes de Barcelona, Series 3 19: 181–200.

Navás L. 1927. Insectos del Museo de París. 4.a serie. Brotéria (Zoológica), 24: 5-33.

Navás L. 1930a. Comunicaciones entomológicas. 12. Insectos de la India. 2.a serie. *Revista de la [Real] Academia de Ciencias Exactas Fisico-Quimicas y Naturales de Zaragoza (1)* 13: 29–48.

Navás L. 1930b. *Neurocolinus* nom. nov. for Colinus Navás, 1925 and *Chenbergus* nom. nov. for *Brachycentrus* Taschenberg, 1865. *Boletín de la Sociedad Entomologica de España* 13: 42–43.

Navás L. 1931. Décadas de insectos nuevos. Decada 1. *Revista de la Real Academia de Ciencias Exactas Fisicas y Naturales de Madrid* 26: 60–69.

Navás L. 1934a. Insectos del Museo de Hamburgo. 2.a serie. *Memorias de la Real Academia de Ciencias y Artes de Barcelona*, Series 3, 23: 499–508.

Navás L. 1934b. Insectos suramericanos. Novena serie. Revista de la Real Academia de Ciencias Exactas Fisicas y Naturales de Madrid 31: 155–184.

Navás L. 1936a. Décadas de insectos nuevos. Década 28. Brotéria, Ciências Naturais 32: 161-170.

Navás L. 1936b. Insectes du Congo Belge. Série IX. *Revue de Zoologie et de Botanique africaines* 28: 333–368.

Navás L. 1936c. Mission Scientifique de l'Omo. Tome III. Fascicule 19. Neuroptera, Embioptera, Plecoptera, Ephemeroptera et Trichoptera. *Memoires du Museum National d'Histoire Naturelle, Paris*, New Series 4: 101–128.

Nguyen L.T., Schmidt H.A., von Haeseler A. & Minh B.Q. 2015. IQ-TREE: A fast and effective stochastic algorithm for estimating maximum-likelihood phylogenies. *Molecular Biology and Evolution* 32: 268–274. https://doi.org/10.1093/molbev/msu300

Ohm P. 1965. *Myrmeleon noackinov*. sp., eine neue Myrmeleontiden-Art von der Balkan Halbinsel (Neuroptera). *Fragmenta Balcanica, Musei Macedonici Scientiarum Naturalium* 5: 107–114.

Okamoto H. 1910. Die Myrmeleoniden Japans. Wiener Entomologische Zeitung 29: 275–300.

Olivier G.A. 1811. Encyclopedie méthodique. Histoire naturelle. Vol. 8 (Insectes). Paris: 175–179.

Oswald J.D. 2020. Lacewing Digital Library. Available from:

https://lacewing.tamu.edu/SpeciesCatalog/Main (accessed 13 Jul. 2020)

Oswald J.D. & Penny N.D. 1991. Genus-group names of the Neuroptera, Megaloptera and Raphidioptera of the world. *Occasional Papers of the California Academy of Sciences* 147: 1–94. https://doi.org/10.5962/bhl.part.3428

Pantaleoni R.A. & Badano D. 2012. *Myrmeleon punicanus* n. sp., a new pit-building antlion (Neuroptera Myrmeleontidae) from Sicily and Pantelleria. *Bulletin of Insectology* 65: 139–148.

Rambur J.P. 1842. *Histoire naturelle des Insectes*. *Nevropteres*. Librairie encyclopédique de Roret. Fain et Thunot, Paris.

Rambaut A. 2009. FigTree ver. 1.3. 1. Computer program distributed by the author. [WWW document]. URL http://tree.bio.ed.ac.uk/software/figtree/.GoogleScholar [accessed on 4 January 2011].

Ronquist F., Teslenko M., van der Mark P., Ayres D.L., Darling A., Höhna S., Larget B., Liu L., Suchard M.A. & Huelsenbeck J.P. 2012. MRBAYES 3.2: Efficient Bayesian phylogenetic inference and model selection across a large model space. *Systematic Biology* 61: 539–542. https://doi.org/10.1093/sysbio/sys029

Röhricht W. 1998. Distribution of *Myrmeleon (Morter) bore* (Tjeder, 1941). *In*: Panelius S.P. (ed.) Neuropterology 1997. *Proceedings of the Sixth International Symposium on Neuropterology* (13–16 July 1997, Helsinki, Finland). *Acta Zoologica Fennica* 209: 221–225.

Saitou N. & Nei M. 1987. The neighbor-joining method: A new method for reconstructing phylogenetic trees. *Molecular Biology and Evolution* 4: 406–425.

https://doi.org/10.1093/oxfordjournals.molbev.a040454

Sekimoto S. 2014. Review of Japanese Myrmeleontidae (Neuroptera). *Insecta Matsumurana* (N.S.) 70: 1–87.

Simon C., Frati F., Beckenbach A., Crespi B., Liu H. & Flook P. 1994. Evolution, weighting, and phylogenetic utility of mitochondrial gene sequences and a compilation of conserved polymerase chain reaction primers. *Annals of the Entomological Society of America* 87: 651–701. https://doi.org/10.1093/aesa/87.6.651

Stange L.A. 1994. Reclassification of the New World antlion genera formerly included in the tribe Brachynemurini (Neuroptera: Myrmeleontidae). *Insecta Mundi* 8: 67–119.

Stange L.A. 2004. A systematic catalog, bibliography and classification of the world antlions (Insecta: Neuroptera: Myrmeleontidae). *Memoirs of the American Entomological Institute* 74: 1–565.

Stange L.A. & Wang H.Y. 1998. *Guide Book to Insects in Taiwan (18)*. Neuroptera, Megaloptera, Raphidioptera: 180–241. Shuxin Press, Taipei.

Tillier P., Giacomino M. & Colombo R. 2013. Atlas de répartition des fourmilions en France (Neuroptera: Myrmeleontidae). *Revue de l'Association Roussillonnaise d'Entomologie* 22: 1–51.

Tjeder B. 1941. A new species of Myrmeleontidae from Scandinavia. Preliminary description. *Opuscula Entomologica* 6: 73–74.

Wang X.L., Zhan Q.B. & Wang A.Q. 2018. Fauna Sinica Insecta Vol. 68 Neuroptera Myrmeleontoidea. Science Press, Beijing. [In Chinese].

Winterton S.L., Hardy N.B. & Wiegmann B.M. 2010. On wings of lace: phylogeny and Bayesian divergence time estimates of Neuropterida (Insecta) based on morphological and molecular data. *Systematic Entomology* 35: 349–378. https://doi.org/10.1111/j.1365-3113.2010.00521.x

Yang C.K. 1999. Myrmeleontidae. *In*: Huang B.K. (ed.) *Fauna of Insects Fujian Province of China. Vol. 3*. Fujian Science and Technology Press, Fuzhou.

Yang D., Liu X.Y. & Yang X.K. 2018. Species Catalogue of China. Vol. 2. Animals, Insecta (II), Neuropterida. Science Press, Beijing.

Zhan Q.B., Ábrahám L. & Wang X.L. 2011. A new record species of *Myrmeleon Linnaeus* from China (Neuroptera, Myrmeleontidae). *Acta Zootaxonomica Sinica* 36: 994–996.

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