COMPARATIVE ANATOMICAL AND HISTOLOGICAL STUDY OF THE RABBITS (*Oryctolagus cuniculus*) ADRENAL GLAND AT THIRTY AND NINETY DAYS AGE.

Ahmed Abdulla Hussein*

Thamir Abood Abass**

* College of veterinary medicine - University of Diyala. Republic of Iraq **Center of Nano-Technology and advance materials - University of Technology.

ABSTRACT

Twelve animals regardless of the sex divided equally into two age periods (30, 90 days age). The adrenal glands were collected and fixed in 10% neutral buffered formalin for cortical tissue and Orth's solution for medullary tissue. These specimens were conducted for histomorphologic investigation applied in both age periods. The glands of rabbits were paired white to creamy in color, oval to crescentic in shape. It was embedded in fatty tissue, lying cranial to the cranial pole of kidney within the retroperitoneal cavity similar to these of other mammals. The left adrenal gland was far off from the left kidney as a compared with the right gland. Histological part included fixation of the adrenal glands and then processed by routine paraffin embedding. The sections of 6 µm thicknesses were stained by H&E, PAS and Van Gieson stains. The adrenal capsule observed as one layer in the two age periods. The zona glomerulosa appeared as arches of cells at sub capsular region, while the zona fasiculata and reticular zone were developed with age. Medulla composed of central vein surrounding by chromaffin cells.

Key words: adrenal gland, rabbit, cortex, medulla, sinusoid.

INTRODUCTION

Rabbits are small mammals in the family *Leporidae* of the order *Lagomorpha*, found in several parts of the world. There are eight different genera in the family classified as rabbits, such as the European rabbit (*Oryctolagus cuniculus*); cottontail rabbits (*genus Sylvilagus*) and Amami rabbit (*Pentalagus furnessi*) and others. The male is called a buck and the female is a doe; a young rabbit is a kitten or kit (Http://en.wikipedia. org/wiki/Rabbit). Their breeding season extends from January to August, when a succession of litters, usually of three to seven young can be produced by the does at 30 day intervals. Doe reach sexual maturity from as early as four months old (McKillop *et al.*, 1993). The mammals' adrenal

Received for publication Sept. 17, 2014.

Accepted for publication Dec. 17, 2014.

gland which investigated after birth was composed of two distinct functional compartments, i.e., the cortex and the medulla. It is the vital endocrine gland that occupies the central role in the regulatory mechanisms of the body metabolism. The parenchyma of the adrenal gland of birds and some animals constituted mainly of three components namely the cortical tissue, chromaffin tissue and vascular sinusoids (Hassan, 1975; Fahmy *et al.*, 2008; Papadkar & Dhamani 2012).

MATERIALS AND METHODS

Twelve healthy rabbits of two different ages were used in this study; those are 30, 90 day's age. The euthanasia was conducted intramuscular injection of ketamine HCl 60 mg/kg and xylazine 6 mg/kg, and after that the abdominal cavity was opened and both right and left adrenal glands were exposed. 10% buffer formalin was injected through abdominal aorta and left for 48 hours for through fixation of the organ. Anatomical study included measuring the weight and length of each animal in different age periods. The current work includes: Position weight, size and dimensions of Adrenal gland. Histologically: other samples washed with distilled water and put in suitable container with Orth's solution as a fixative containing potassium dichromate, sodium sulfate, formalin and water for 24 hours then specimens were transferred to 2.5 % aqueous solution of potassium dichromate for 48 hours after that they wash by putting them under running water overnight and adding of 5 ml of acetic acid to it will improve this fixative for the demonstration of chromaffin granules in the medullary tissues (Luna, 1968). Another half of specimens were fixed with neutral buffer formalin to the cortical tissue of the adrenal gland. The staining method is done for differentiate different slide components. Hematoxylin & Eosin stain, Periodic Acid Schiff (PAS) stain, Van Gieson stain.

RESULTS AND DISCUSSION

Anatomically at thirty and ninety days old rabbits, the right and left adrenal glands regardless of their gender of the local rabbit were located in the abdominal cavity cranial to the cranial pole of the kidney. Both glands were white to creamy color positioned retroperitoneal and embedded in a mass of fat (Fig. 1, 2). The shape of adrenal gland appeared crescent in the left adrenal while the right gland is an oval in the shape (Fig. 3). The cross section of the adrenal gland at all ages showed an outer pallor part which is the cortex and an inner dark brown part which is the medulla (Fig. 4). Histologically: the adrenal glands were enclosed in the capsule which showed two layers (outer fibrous and inner cellular) at thirty and ninety days of age (Fig. 5).

Zona glomerulosa

At thirty days of age, the zona glomerulosa was well differentiated and contains many sinusoids. The cells were polyhedral in shape with relatively large oval to rounded nuclei with relatively small amount of cytoplasm, the cells arranged as arches under capsule, the trabeculae appear thin and it showed many mitotic figures (Fig.6). At ninety day age the zona glomerulosa the cells have large rounded nuclei and less amount of cytoplasm. The sinusoids were well observed beside the cellular proliferation of mitotic division among the cells (Fig.7).

Zona fasiculata

It was wide and extended to occupy a large area of the cortex compared with the other two zones of cortex. It extended between upper zona glomerulosa and lower zona reticularis. The cells of this zone appeared foamy in appearance due to the presence of vacuoles in their cytoplasm. The zone composed of cellular cords, the cells showed foamy picture due to the presence of vacuoles in their cytoplasm. The examination showed high proliferation with containing small rounded basophilic nuclei, the cells contain oval or rounded darkly stained basophilic nuclei and arranged as a cords of cuboidal or low columnar cells (Fig. 8). Zona fasiculata at ninety days old rabbits was well developed Histological picture was similar to that of the thirty day of age (Fig. 9).

Zona Reticularis

It composed of large polyhedral cells with oval or rounded basophilic nuclei and showed different degrees of mitotic division and the presence of many sinusoids in 30 days. Clear obvious boundaries and clear demarcation among its cells (Fig 10). There were no clear demarcations between zona reticularis and medulla but the cells of both reticular zone and medulla overlap with each other due to development by migration of chromaffin cells to the center of the adrenal gland (Fig. 11).

Medulla

It occupied the central part of the adrenal gland and composed of the medullary cells named chromaffin cells. Its characteristic feature was the presence of the central vein. Chromaffin cells arranged in cellular cords or clusters surrounding the central vein. These cells possessed either epinephrine or nor epinephrine secretory granules. The chromaffin cells stained brown when fixed with potassium dichromate salts (Fig.12).while the histological observation in ninety days age and the constant development of adrenal gland and the chromaffin cells have a large central nuclei and granular cytoplasm (Fig.13).



Fig. 1: Adrenal glands in situ of 30 days old-rabbits, respectively. It showed right adrenal gland (R.A), left Adrenal gland (L.A), right kidney (R.K) and left kidney (L.K)



Fig. 2: Adrenal glands in situ of 90 days old-rabbits, respectively. It showed right adrenal gland (R.A), left Adrenal gland (L.A), right kidney (R.K) and left kidney (L.K).



Fig.3: *The shape of adrenal gland. It showed a crescent shape in the left adrenal while the right gland is an oval in shape.*



Fig.4: Cross section of the adrenal gland in rabbit showed capsule (cap), Cortex(C) and medulla (M).

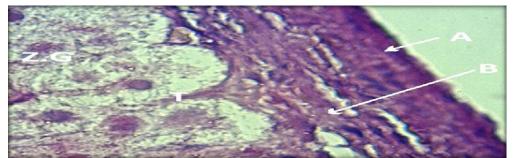


Fig.5:*Capsule of 30 days -old rabbits. It showed two Capsule cellular (B)* and fibrous (A), zona glomerulosa cells (Z.G), trabecula (T), PAS,X 1000

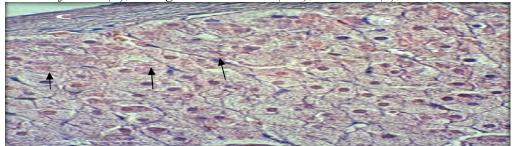


Fig.6: Zona glomerulosa of 30 days -old rabbits. It showed zona glomerulosa cells (arrows), capsule(c) , H&E,X400

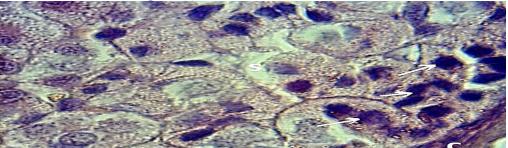


Fig.7:Zona glomerulosa of 90 days -old rabbits. It showed Mitotic division (white arrows), capsule (c), H&E,X1000

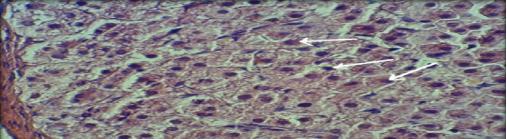


Fig.8:Zona fasiculata of 30 days -old rabbits. It showed cords of basophilic cells (white arrows), capsule(red arrow), H&E, X400

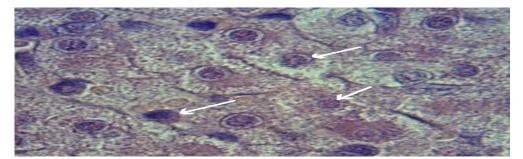


Fig.9:Zona fasiculata of 90 days -old rabbits. It showed cords of basophilic cells (white arrows), H&E, X1000



Fig.10:Zona reticularis of 30 days -old rabbits. It showed the network of basophilic nuclei cells (white arrows), H&E X400

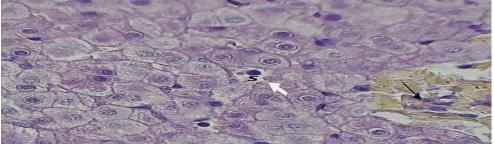


Fig.11:Zona reticularis of 90 days -old rabbits. It showed the network of basophilic nuclei cells (white arrow), and medulla(black arrows), PASX1000

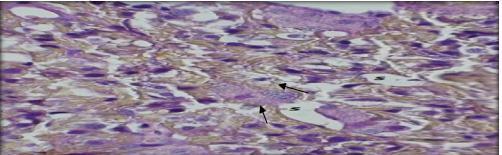


Fig.12:Medulla of 30 days -old rabbits. It showed the chromaffin cells (arrows), H&E, X1000

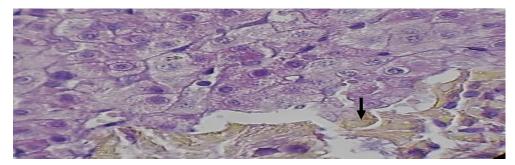


Fig.13:Medulla of 90 days -old rabbits. It showed the chromaffin cells (Arrow), PASX400, H&E, X1000

Anatomical study

Both glands were positioned retroperitoneal and embedded in a mass of fat located in the abdominal cavity cranial to the cranial pole of the kidney. The distance between the right adrenal gland and its related right kidney was lesser than the distance present between the left gland and the left kidney. It maybe to shortage of renal or adrenal artery. While the shape, color, weight character were previously observed in the bats (Papadkar& Dhamani, 2012) and rats too (Ezumi *et al.*, 2007). Gilbert, 2004), in cats (Westropp *et al.*, 2003; Tomich, 1965).

Histological study

Capsule similar findings were previously found in the *Rattus norvegicus* (Ezumi *et al.*, 2007)) and rabbits (Fahmy *et al.*, 2008). The thickness of the capsule at 30 days age gradually became two layers outer (fibrous) and inner cellular. These observations were found in the adrenal of rat (El Sayed *et al.*, 1990); goat (Dimova, 2003) and rabbit (Fahmy *et al.*, 2008). The Zona glomerulosa of the both ages was characterized by aggregation of small polyhedral cells as arches beneath the capsule. This histological result also observed and documented previously by (El Sayed *et al.*, 1990) in the adrenal of albino rats, (Rosol *et al.*, 2001) in the adrenal of mice, (Hafez *et al.*, 2005), in the adrenal of male albino rats and (Fahmy *et al.*, 2008) in the adrenal of the rabbit. The mitotic figures in this zone in all ages also observed by Lombarado & Cortesini (1990); Chen *et al.* (1998); Yılmaz & Girgin (2005) and Mughal *et al.* (2004) in the adrenal glomerulosa of rat.

These cells were possessed large amount of clear cytoplasm with small rounded basophilic nuclei these histological observations were found previously too by Clark *et al.* (2005) in the adrenal glands of the Dolphin. In Zona fasiculata showed an important aspect through its development. It was wide and extended to occupy a large area of the cortex compared with the other two zones of cortex.

It extended between upper zona glomerulosa and lower zona reticularis. Zona fasiculata cells appeared foamy cells in appearance due to the presence of vacuoles in their cytoplasm.

These vacuoles present as a result of dissolving of fat droplets with xylene through histological procedure of slide preparation. The cells contain oval or rounded darkly stained basophilic nuclei and arranged as a cords of cuboidal or low columnar cells. These histological observations were previously mentioned by Hafez et al. (2005) in adrenal gland of the albino rats Rosol et al. (2001); Bielohuby et al., 2007) in mice, (Yılmaz and Girgin, 2005) in the porcupine and rat, respectively by Aughey & Frye (2011) and (Hafez et al. (2005) albino rat. Zona Reticularis innermost zone of the adrenal cortex undergoes the similar morphological changes that were taking placed in the zona fasiculata at one day undifferentiated zone. The cells of zona reticularis were arranged as net-work and contained large central nucleus. This observation was mentioned too by Junqueira and Carneiro (1980) and Fahmy et al. (2008) in the adrenal reticularis of the rabbit and Hafez et al. (2005)) in the adrenal of the albino rat. This result agrees with those of Junqueira and Carneiro (1980), Mughal et al. (2004) in the rat; (El shennawy and Aboelwafa (2011) in albino rat. Medulla occupied the central part of the adrenal gland and composed of the medullary cells named chromaffin cells. Its characteristic feature mentioned similarly by Yılmaz and Girgin(2005) in the porcupine was the presence of the central vein, this histological feature was observed by many other investigators Coupland & MacDougall, 1966; Lemos et al., 2006; Mescher, 2010; Yılmaz and Girgin, 2005. This result was mentioned by El Sayed et al. (1990) in albino rat, Papadkar & Dhamani (2012) in Indian male fruit bat Lemos et al. (2006) in Rhesus Macaque and Papadkar & Dhamani (2012) in human embryo and Yılmaz & Girgin, (2005); (Chavhan et al.(2011). This result mentioned by Mescher, (2010); Aughey & Frye (2011); Junqueira and Carneiro (1980), Wheater et al. (1987) and Junqueira et al. (1998) in humans and some animals.

REFERENCES

- Aughey, E. and F.L. Frye. 2011 . Comparative veterinary histology with clinical correlates. Manson publishing, Veterinary press, 2nd ed.pp156.
- Bielohuby, M., N. Herrbach, R. wanke, C. Maser-Gluth, F. Beuschlein, E. Wolf And A. Hoeflich. 2007. Growth analysis of mouse adrenal gland from wearing to adulthood: time and gender dependent alterations of cell size and number in the cortical compartment. *Am. J. Physiol. Endocrinology. Metabolic disease*, 293: 139 – 146.
- Chavhan, P.R., A.A.Dhamani, and S.D. Misar. 2011. Histoarchitectural changes in the adrenal gland of female bat (*taphozous kachensis*) during estrus and pregnancy. *Journal of Cell and Tissue Research*, 11(2) 2857-2863.
- Chen, L.G., Kao, C.H. and Yang, V.C. 1998. Late effects in mouse thyroid and adrenal gland after 60 co ∞-irradiation of the brain: An ultra-structural study. *Scanning Microscopy*, 12(3): 475 486.
- Clark, L.S., D.C. Pfeiffer and D.F. Cowan. 2005. Morphology and histology of the atlantic bottlenose dolphin (*Tursiops truncatus*) adrenal gland with

emphasis on the medulla. *Anatomy Histology Embryology*, 34(2): 132-40.

- Coupland, R.E. and J.D. MacDougall. 1966. Adrenaline formation in noradrenaline storing chromaffin cells invitro induced by corticosteron in rabbit. *Journal of Endocrinology*, 36: 317-318.
- Dimova, T. 2003. Histological features of goat adrenal gland during puberity. comptes rendusde l'Academie bulgare des Scinces., 56(4):350-352.
- ElSayed, S., S.N., Ibrahim, M.K. Tawfik and Y. ElArian, 1990. Histological, Histochemical and morphometric changes of the contra lateral adrenal gland after unilateral adrenalectomy. *Sci. Med. J. Cai. Med. Synd.*, 2(4):20-26.
- Elshennawy, W. W. and H.R. Aboelwafa. 2011. Structural and ultra-structural alterations in mammalian adrenal cortex under influence of steroidogenesis inhibitor drug. *Journal of American Science*, 7(8)50-51.
- Ezumi, M. F., S.S. Amrah, C. G. farid, and S. S. Mohsin. 2007. Morphological Characteristics of the adrenal of (*Rattus norvegicus*) Arevisit by Scanning Electron Microscopy. *Annals of Microscopy*, 7: 33-35.
- Fahmy, H. F., A. A. Abd El Samad, and M. M. Omar. 2008. A Comparative Study of the Effect of Budesonide versus Prednisolone on Adrenal Gland of the Male Rabbit. *Egyptian Journal of Histology*, 31(2): 188 – 197.
- Gilbert, J. A. 2004. Necessary conditions to maintain Rat adrenal chromaffin cells in primary culture Cell biology of chromaffin cells. R. Borges & L. Gandia Eds. Institute Teofilo Hernando, Spain.
- Hafez, M. S., N. A. Makhlouf and H. A. Saleh. 2005. Histological changes of the contra lateral adrenal cortex following unilateral adrenalectomy in male albino rat. *Journal of Histology*, 28(2): 251-262.
- Hassan, A. 1975. Histological and histochemical studies on the adrenal glands of geese and ducks. *Acta Vet. Acad. Sci. Hung.*, 25(1): 19-26. <u>Http://en.wikipedia</u>. org/wiki/Rabbit.
- Junqueira, L. C. and J. Carneiro. 1980. Basic histology. 3rd ed. Los. Altos. California, pp: 421-429.
- Junqueira, L. C., J. Carneiro and R.O. Kelly. 1998. Basic histology, 9th ed. Mc Graw and Hill, New York. PP: 387-395.
- Lemos, D., J. Downs and H. Urbanski. 2006. Twentyfour-hour Rhythmic Gene Expression in the Rheseus Macaque Adrenal Gland. *Molicular Endocrinology*, 20(5): 1164-1176.
- Lombarado, L. D. and C. Cortesini. 1990. The Zona glomerulosa of the adrenal gland in rats with portacaval shunt. An ultra-structural and morphometric study. *Int. J. Exp. path.*, 71 : 639-646.
- Luna, G. 1968. Manual of histological staining methods of the armed forces institute of pathology, 3rd ed. McGraw Hill Book Company, New York.

- McKillop, G., S. V. Ginella, C. J. Wilson, A. J. Hanlon and B. D. Pugh, 1993. The effects of power failure on European wild rabbits at electric fences. *Journal of Applied Animal Behaviour Sciences.*, 35: 277-290.
- Mescher, A. L. 2010. Junqueira's Basic Histology. 12th ed. Chapter 20, McGraw-Hill Companies.
- Mughal, I. A., A. S. Qureshi and M. S. Tahir. 2004. Some Histological observations on postnatal growth of rat adrenal gland with advancing age. *International Journal of Agriculture & Biology*, 6(2): 413-417.
- Papadkar, J. N. and A. A. Dhamani. 2012. Seasonal changes in the adrenal gland of the Indian male fruit bat (*Pteropus Giganteus*) during the reproductive cycle. *Life Science Leaflets*, 8: 11-17.
- Rosol, T. J., J. T.Yarrington, J. Latendresse, and C. C. Capen. 2001. Adrenal Gland: Structure, Function, and Mechanisms of Toxicity. *Toxicology* and Pathology, 29(1): 41- 48.
- Sant, S. 2008. Embryology for medical students. 2nd edition, Ch. 14, p: 272
- Tomich, P.Q. 1965. Weight Variation in adrenal gland of the Mongoose in Hawaii, Pacific science, XIX.
- Westropp, J. L., K. A. Welk and C. A. Buffington. 2003. Small adrenal gland in cats with feline interstitial cystitis, *Journal of Urology*, 170: 2494-2497.
- Wheater, P. R., H. G. Burkitt and V. G. Daniles. 1987. Functional histology, a text and color atlas, 2nd ed. Churchill Livingstone, London. pp: 267-271.
- Yılmaz, S. and A. Girgin. 2005. Light and electron microscopic observations on the structure of the porcupine (*Hystrix cristata*) adrenal gland. *Vet. Arch.*, 75 (3): 265-272.

دراسة نسجية تشريحية مقارنة للغدة الكظرية في الإرنب عند الثلاثين والتسعين يوماً من العمر.

ثامر عبود الدباغ **

احمد عبدالله حسين

* مدرس مساعد - جامعة ديالي- كلية الطب البيطري **أستاذ مساعد - الجامعة التكنولوجية - معهد التقانات الاحيائية - www.ahmedess838@yahoo.com

المستخلص

قسم اثنا عشر أرنباً بغض النظر عن الجنس بالتساوي إلى فترتين عمريتين هي 90 ، 30 يوما. جمعت الغدد الكظرية وثبتت في 10% محلول الفورمالين المتعادل للنسيج القشري ومحلول الاورث لنسيج اللب هذه العينات نقلت للاستقصاء النسجي الشكلي الذي طبق في كلتا الفترتين العمريتين. الغددالكظرية في الارنب هي زوج من الغدد البيضاء الى كريمي اللون ، بيضوية إلى هلالية الشكل، مطمورة في النسيج الدهني ، تقع أماميا للقطب الأمامي للكلية ضمن تجويف خارج الخلب كما في باقي الثدييات. الغدة اليسرى بعيدة عن الكلية اليسرى بالمقارنة مع الغدة اليمنى. الجزء النسجي اشتمل على تثبيت الغدة الكطرية بعد ذلك تمت معاملتها بالطمر الروتيني بشمع البرافين. المقاطع النسجي التي سمكها 6 مايكرون صبغت بصبغات الهيموتوكسلين والايوسين وصبغة ملون شيف حمض البيروديك وصبغة فان كيسن. محفظة الغدة الكظرية شوهدت كطبقة واحدة في الفترتين العمريتين . كاقواس من الخلايا تحت منطقة محفظة الغدة بينما النطاق الشبكي تطورا مع العمر. لب الغدة الكظرية يتالف من وريد من وريد من النطاق الحريمي والنطاق الشبكي تلورا مع العمر.

الكلمات المفتاحية : الغده الكظرية ، الأرنب ، قشرة الكظرية ، لب الكظرية ، الجيبانيات