Phenotypic characterization of *Candida* spp. isolates from chronic periodontitis patients

Cristiane Yumi Koga-Ito¹; Edson Yukio Komiyama²; Clélia Aparecida de Paiva Martins²; Silvana Soléo Ferreira dos Santos³; Ivan Balducci⁴; Antonio Olavo Cardoso Jorge¹

¹PhD in Microbiology and Immunology, Department of Oral Biosciences and Diagnosis, Dental School of São José dos Campos, São Paulo State University, Brazil;

²MSc in Microbiology and Immunology, Department of Oral Biosciences and Diagnosis, Dental School of São José dos Campos, São Paulo State University, Brazil;

³PhD in Oral Biopathology, Basic Institute of Biosciences, University of Taubaté, Brazil;

⁴MSc in Biostatistics, Department of Social Dentistry, Dental School, São José dos Campos, São Paulo State University, Brazil.

Received for publication: September 05, 2007 Accepted: May 30, 2008

Abstract

Aim: Several typing methods for *Candida* spp. have been suggested in the literature in order to distinguish isolates for studies about the virulence or infection routes of these microorganisms and, in particular, for epidemiological purposes. The aim of this study was to establish a comparison between the phenotypic profile of oral *Candida* isolates from periodontitis patients and control individuals. **Methods**: The morphotyping and biotyping of 35 *C. albicans* isolates obtained from chronic periodontitis patients and 48 isolates from control individuals were performed. For morphotyping, the isolates were plated on malt extract agar and incubated for 10 days. Sixteen different morphotypes were observed for *C. albicans*, the most frequently observed being 0000 and 0001. **Results**: Biotype 0000 (complete absence of fringe) was most prevalent among the isolates obtained from periodontitis patients compared to those from control individuals, with statistical significance. Biotyping revealed 5 different biotypes with higher prevalence of the biotype 357 among the isolates from control and periodontitis groups. **Conclusions**: The results obtained by biotyping of the isolates did not permit to differentiate a characteristic model related to periodontal disease, whilst the morphotype 0000 was most frequently isolated from periodontitis patients.

Key words:

periodontitis, Candida albicans, biotyping, morphotyping

Introduction

Candida spp. have been correlated to cases of severe and refractory periodontal infections, particularly in immunocompromised patients or individuals under antimicrobial therapy for long periods¹⁻⁵. Several *Candida* spp. typing methods have been suggested in the literature in order to distinguish isolates for studies about the virulence or infection routes of these microorganisms and, in particular, for epidemiological purposes⁶⁻¹⁰. Among the methods based on phenotypic characteristics, serotyping, biotyping, morphotyping and sensitivity to *killer* toxins are included⁷. There are also methods based on genotypic characteristics, such as immunobloting¹¹ and DNA fingerprinting techniques¹².

Morphotyping is considered an efficient, reproducible and

Correspondence to:

Cristiane Yumi Koga-Ito

Faculdade de Odontologia de São José dos Campos/UNESP Av. Eng. Francisco José Longo, 777, 12245-000 - São José dos Campos, SP, Brasil Phone: +55-12-3947-9033. e-mail: cristianeykito@directnet.com.br low cost method for *C. albicans* characterization^{7,13}. Moreover this method reveals a good discriminatory power. The variation in the morphology of *Candida* colonies was firstly observed by Negroni¹⁴. Later, based on Brown-Thomsen's¹⁶ observations about the morphological variations of colonies due to alterations in the incubation temperature or medium composition, Phongpaichit *et al.*¹⁵ proposed a typing method with codes. This coding system was based on the characteristics of the colonies as well as on their surfaces.

Hunter *et al.*¹⁷, studying the morphotype distribution among 446 *C. albicans* strains, suggested a typing method based in the characteristics of colony surfaces, represented by 4 digits.

The phenotypic switching and *Candida* morphotypes have been associated to their virulence. Different phenotypic expressions in relation to colony growth are related to variations that are considered responsible for the several degrees of virulence^{7,18}. Phenotypic switching occurs frequently and causes changes in colony morphology and cell surface properties, such as alteration in the adherence to epithelial cells¹⁹. The biotyping technique proposed by Odds and Abbott²⁰ has been reported as a method with an adequate discriminatory power mainly when associated with the morphotyping method²¹. Considering the adequate discriminatory power of the combination of morphotyping and biotyping methods, the purpose of this study was to establish a comparison between the phenotypic profile of oral *Candida* isolates from periodontitis patients and control individuals, aiming to correlate specific phenotypic features to the occurrence of periodontal disease.

Material and Methods

This research project has been independently reviewed and approved by the Bioethics Research Committee of the Dental School of São José dos Campos, São Paulo State University, Brazil (Protocol #72/99-PH/CEP).

Candida albicans isolates from chronic periodontitis (n=35) and control individuals (n=48) were included in this study. These strains were previously isolated and belonged to the strain collection of the Microbiology Laboratory at the Dental School of São José dos Campos, São Paulo State University.

Isolates from chronic periodontitis were obtained from 88 individuals aged 25 and 62 years (mean age of 41.33 \pm 5.54 years), with at least two 5-mm deep periodontal sites and diagnosed clinically as chronic periodontitis patients. Control group isolates were obtained from 68 individuals aged 25 to 55 years (mean age of 34.45 \pm 7.93 years) diagnosed as periodontally healthy patients.

Candida isolates were morphotyped according to the methodology proposed by Hunter et al.7 and Pongpaichit et al 15. Briefly, the isolates were plated on Sabouraud dextrose agar and incubated at 25°C for 48 h. Then, yeast suspensions were prepared in sterile distilled water adjusted to the turbidity of tube #3 of McFarland scale. Using sterile swabs, the suspensions were plated on the surface of malt extract agar (Difco, Detroit, MI, USA). Plates were maintained at room temperature in a light-proof environment for 10 days. After this period, the macromorphological aspects of the fringes and surface of the colonies were evaluated. The results of morphotyping were recorded using 4-digit codes. Biotyping methods were performed according to Odds and Abbot²², with a combination of tests of tolerance (to pH 1.4 and NaCl), resistance (5-fluorocytosine, safranine and boric acid), enzymatic activity (proteinase) and growth in presence of urea, sorbose and citrate. The isolates were plated on Sabouraud dextrose agar and incubated at 37°C for 24 h. Then, saline suspensions adjusted to the turbidity of tube #3 of McFarland scale were obtained. Aliquots of 100 μ L of these saline suspensions were deposited in the wells of a Steers' inoculator and plated in the test culture media and the positive control (Sabouraud agar). All tests were performed in triplicate. Plates were incubated at 37°C for different periods of time (3 to 4 days for control, safranine, urea, citrate, boric acid and sodium chloride; 6 to 7 days to

resistance to pH 1.4 and tests of sorbose, proteinase and 5fluorocytosine). Positive tests were considered when the strains grew at pH 1.4, 5-fluorocytosine, sodium chloride, boric acid, urea, sorbose and sodium citrate, as well as for the strains that formed colonies with diameter grater than 2 mm in presence of safranine.

The results of biotyping were recorded using 3-digit codes, according to Odds and $Abbott^{22}$. Values of 1, 2, 3 and 4 were attributed to the positive tests and 0 to the negative tests.

Statistical Analysis

The data regarding the occurrence of the morphotypes and biotypes were compared between the periodontitis and control groups by two proportion Z test at 5% significance level.

Results

The morphotype 0000 was the most frequently observed (50%) among the oral isolates from control individuals. The morphotype 0001 was observed in 12.5% of the isolates. The morphotypes 1232, 7240, 0004, 2331, 1230 and 7520 were also observed (Table 1).

Among the isolates from chronic periodontitis patients, the morphotype 0000 was also the most frequently found (77.14%) differing significantly from the control group (p=0.007) (Figure 1). Other 7 morphotypes were observed among the isolates of this group (Table 1).

Biotypes 357 and 757 were the most frequently observed in both groups without statistically significant differences (p=0.522) (Table 2).

Discussion

Candida genus yeasts are considered opportunistic microorganisms^{23,24} and may lead to severe periodontal infections in immunocompromised patients or individuals under antimicrobial therapy for long periods¹. Under specific situations, such as in immunodepressed patients²⁵, superinfection by Candida can be refractory to conventional periodontal treatments. In fact, C. albicans presents virulence factors that can have an important role in the pathogenesis of periodontal disease such as the ability of penetrating the epithelium, inhibiting polymorphonuclear cells and causing lysis of monocytes^{8,26}. Also, Lu et al.²⁷ have described the hyphal invasion capacity of C. albicans to inhibit the production of antimicrobial peptides by oral epithelium. Candida species have been isolated from the subgingival microbiota of the gingival tissues of patients with periodontal abscesses1, periodontitis28,29, and patients with chronic periodontitis under therapy with antibiotics¹⁷.

Morphotyping has been employed in epidemiological and virulence studies. Ribeiro *et al.*³⁰ studying children with Down's syndrome showed that the *C. albicans* isolates from these patients induced more formation of fringes compared to the isolates obtained from control individuals, and the authors suggested a correlation with increased virulence. In fact, Hunter *et al.*¹⁷ had previously suggested a correlation

Table 1 - Distribution of the morphotypes observedamong Candida albicans isolates from control individualsand periodontitis patients.

CONTROL GROUP (n=48)			
Morphotype	n	%	
0000	24	50.0	
7340	1	2.1	
7540	1	2.1	
5340	2	4.1	
0001	6	12.5	
1040	1	2.1	
1240	2	4.2	
7520	1	2.1	
7320	1	2.1	
7330	1	2.1	
7531	1	2.1	
0002	2	4.1	
0006	2	4.1	
1240	1	2.1	
7321	1	2.1	
3340	1	2.1	
PERIO	DONTITIS GROU	P (n=35)	
0000	27	77.14	
1232	1	2.86	
7240	2	5.70	
0004	1	2.86	
2331	1	2.86	
1230	1	2.86	
7520	1	2.86	
7540	1	2.86	
100 - z = 2.86; p = 0.0 90 - (C1, 95% 7.30 t) 80 - 70 - 60 - 50 - 40 - 30 - 20 - 10 - 10 - (C1, 95% 7.30 t))07<0.05 o 46.98)	z = 0.64; p = 0.522>0.05 (Cl, 95% -13.23 to 26.09%)	

Fig. 1 - Statistical comparison of the morphotype 0000 and biotype 357 occurrence in the studied groups.

Control Periodontitis

Biotype 357

Control Periodontitis

Morphotype 0000

between invasive infections and isolates with discontinued fringes. The increased virulence of isolates with fringes was also related by Spadari *et al.*³¹. These authors observed a higher degree of adherence in these isolates compared to strains without fringes. In the present study, this tendency was not observed, as similar proportion of isolates in the groups showed discontinued fringes. Four isolates from the

Table 2 - Distribution of biotypes observed amongCandida albicans isolates from control individuals andperiodontitis patients

CONTROL GROUP (n=48)			
Biotype	n	%	
357	36	75.0	
757	9	18.7	
157	1	2.1	
313	1	2.1	
353	1	2.1	
PERIODONTITIS GROUP (n=35)			
357	24	68.6	
757	5	14.2	
777	3	8.6	
377	2	5.8	
577	1	2.8	

control group (8.33%) and 3 from the periodontitis group (8.57%) presented discontinued fringes. These results suggest that no correlation between colonies with discontinued fringes and periodontitis occurrence could be done.

The morphotype 0000 (complete absence of fringes) was the most frequently isolated from the oral cavity of control and periodontitis patients, and this result corroborates the absence of correlation between the presence of discontinuous fringes, which is associated to filamentation, and periodontitis occurrence. Previous studies^{6,17,32} also observed the higher incidence of this morphotype among isolates from the oral cavity. This result may suggest that the mycelial form formation might not be an essential feature of Candida in the periodontal milieu, and other factors (i.e. proteolytic enzymes and toxins production) might be more important contributors to the pathogenesis of this disease. For more detailed data, further studies on these features could be useful. Also, studies on genotyping could provide important results. In fact, several authors have pointed out the importance of combining phenotyping and genotyping methods^{6,9,33,34}.

Hunter *et al.*⁷ have demonstrated the occurrence of the morphotypes 7540 and 7340 among isolates from several sites of the body. These morphotypes were also observed among the isolates of the present study. The great variability of morphotypes in the control group (16 models) has been reported elsewhere¹⁷. Ribeiro *et al.*²⁹ did not observe the occurrence of morphotype 0000 and found a high prevalence of morphotype 5530 in Down's syndrome patients.

The biotyping method proposed by Odds and Abbott²⁰ has been considered as a method with adequate discriminatory power. However, in the present study, it was not able to distinguish between periodontitis and control isolates. Biotyping permitted the identification of 5 different biotypes among the isolates from the oral cavity of control individuals. The biotypes 357 and 757 were the most frequently observed (75% e 18.8%, respectively). Among the isolates from periodontitis patients, the biotypes 357, 757, 777, 377 and 577, and the model 357 (68.6%) were the most frequently observed. In the present study, most isolates assimilated urea and citrate and did not assimilate sorbose. Also, these isolates were tolerant to salt and developed in presence of boric acid and safranine. Neely *et al.*³⁵ also verified a high prevalence of isolates with these features among *Candida* isolates from children.

In conclusion, the results obtained by biotyping of the isolates did not allow distinguishing a characteristic model related to periodontal disease, whilst the morphotype 0000 was most frequently isolated from periodontitis patients.

References

- 1. Hevoluo H, Hakkarainen K, Paunio K. Changes in the prevalence of subgingival enteric rods, staphylococci and yeasts after treatment with penicillin and erythromycin. Oral Microbiol Immunol 1993;8:75-9.
- 2. Slots J, Feik D, Rams TE. Age and sex relationshisp of superinfecting microorganisms in periodontitis patients. Oral Microbiol. Immunol 1990; 5:305-8.
- Oliveira LF, Jorge AOC, Santos S S. In vitro minocycline activity on superinfecting microorganisms isolated from chronic periodontitis patients. Braz Oral Res 2006; 20:202-6.
- 4. Dáhlen G. Microbiological diagnostics in oral diseases. Acta Odontol Scand 2006; 64: 164-8.
- Ito CY, de Paiva Martins CA, Loberto JC, dos Santos SS, Jorge AOC. In vitro antifungal susceptibility of Candida spp. isolated from patients with chronic periodontitis and from control patients. Braz Oral Res 2004; 18: 80-4.
- Candido RC. Candida albicans : marcadores epidemiológicos em amostras isoladas de diferentes materiais biológicos. São Paulo, 1991, 167p. (Ph. D. Thesis. Escola Paulista de Medicina. EPM).
- 7. Hunter PR. A critical review of typing methods for Candida albicans and their applications. Crit Rev Microbiol 1991; 17:417-34.
- Pfaller M.A. Epidemiological typing methods of mycoses. Clin Infect Disc 1992; 14(suppl. 1): S4-10.
- Maffei CM, Paula CR, Mazzocato CS, Franceschini S. Phenotype and genotype of Candida albicans strains isolated from pregnant women with recurrent vaginitis. Mycopathologia 1997; 137:87-94.
- Manfredi M, McCullough MJ, Al-Karaawi ZM, Vescosi P, Porter SR. In vitro evaluation of virulence attributes of Candida spp. Isolated from patients affected by diabetes mellitus. Oral Microbiol Immunol 2006; 21:181-9.
- Lee W, Burnie J, Matthews R. Fingerprinting Candida albicans. J Immunol Meth 1986; 93: 177-82.
- Scherer S, Stevens DA. Aplication of DNA typing methods to epidemiology and taxonomy of Candida species. J Clin Microbiol 1987; 25: 675-9.
- 13. Giammanco G M, Lopes M.M, Coimbra RS, Pignato S, Grimont PAD, Grimont F et al. Value of morphotyping for the characterization of Candida albicans clinical isolates. Mem Instit Oswaldo Cruz.2005; 100: 483-90.
- Negroni P. Variacion hacia el tipo R de Mycotorula albicans. Rev Soc Arg Biol 1935; 11: 449.
- Phongpaichit S, Mackenzie DWR, Fraser C. Strain differentiation of Candida albicans by morphotyping. Epidemiol Infect 1987; 99: 421-8.
- Brown-Thomsen J. Variability in Candida albicans (Robin) Berkhout. I. Studies in morphology and biological activity. Hereditas 1968; 60: 355-98.
- Hunter PR, Fraser CAM. Application of a numerical index of discriminatory power to a comparison of four physiochemical typing methods for Candida albicans. J ClinMicrobiol 1989;

27: 2156-60.

- O'Connell B, Coleman DC, Bennett D, Sullivan D, Mc Cann SR et al. An epidemiological study of Candida species infection in cancer patients using genetic fingerprinting and morphotyping. J Hosp Infect 1995; 31: 211-7.
- Mattews RC. Pathogenicity determinants of Candida albicans: potencial targets for immunotherapy. Microbiol 1994; 140: 1505-11.
- Odds FC, Abbott AB. Modification and extension of tests for differentiation of Candida albicans species and strains. Sabouraudia 1983; 21: 79-81.
- Otero L, Vásquez L, Palacio V, Vásquez S, Carreño F, Méndez FJ. Comparison of seven phenotyping methods for Candida albicans. Eur J Epidemiol 1995; 11: 221-224.
- 22. Odds FC, Abbott AB. A simple system for the presumptive identification of Candida albicans and differentiation of strains within the species. Sabouradia 1980; 18: 301-317.
- Arendorf TM, Walker DM. Candida albicans: Its association with dentures, plaque and oral mucosa. J Dasa 1980; 35: 563-569.
- Leung KW, Dassanayake RS, Yau JYY, Jin LJ, Yam WC, Samaranayake L I. Oral colonization, phenotypic, and genotypic profiles of Candida species in irradiated, dentate, xerostomic nasopharyngeal carcinoma survivors. J Clin Microbiol 2000; 38:2219-26.
- Brawner DL, Cutler JE. Oral Candida albicans isolates from nonhospitalized normal carriers, immunocompetent hospitalized patients, and immunocompromised patients with or without acquired immunodeficiency syndrome. J Clin Microbiol 1989; 27: 1335-41.
- 26. Barret-Bee KY, Wilson RG, Ryley JF. A comparison of phosphoslipase activity cellular adherence and pathogenicity of yeasts. J Gen Microbiol 1985; 131: 1217-21.
- Lu Q, Jayatilake JAMS, Samaranayake LP, Jin L. Hyphal invasion of Candida albicans inhibits the expression of human â-defensins in experimental oral candidiasis. J Investig Dermatol 2006; 126:2049-56.
- Del Castilho L, Bikandi J, Nieto A, Quindós G, Sentandreu R, Pontón J. Comparison of morphortypic and genotypic methods for strain delineation in Candida. Mycoses 1997; 40: 445-59.
- 29. Slots J, Rams TE, Listgarten MA. Yeasts, enteric rods and pseudomonas in the subgingival flora of severe adult periodontitis. Oral Microbiol Immunol 1988; 3: 47-52.
- Ribeiro EL, Scroferneker ML, Cavalilaes MS, Campos CC, Nagato GM, Souza NA et al. Phenotypic aspects of oral strains of Candida albicans in children with Down's Syndrome. Braz J Biol 2006; 66: 939-44.
- 31. Spadari E, Arosio V, Malighetti V, Bellotti MG, Zambelini A, Spadari F, et al. Morphotipi de Candida albicans e adesività in vitro a cellulle della mucosa orale di pazienti HIV-positivi e com AIDS dopo esposizione delle blastopore al fluconazolo. Parte II. Minerva Stomatol. 1998; 47: 293-7.
- 32. Barreto de Oliveira MT. Leveduras isoladas da mucosa bucal de portadores sadios, pacientes com Sida e neoplasias: produção de exoenzimas e tipagem das amostras de Candida albicans. São Paulo, 1993, 107p. (Master's degree dissertation. Institute of Biomedical Sciences, USP).
- 33. Betremieux P, Chevrier S, Quindós G, Sullivan D, Polonelli L, Guiguen C. Use of DNA fingerprinting and biotyping methods to study a Candida spp. outbreak in neonatal intensive can unit. Pediatr Infect Dis J 1994; 13: 899-905.
- Maffei CML, Paula CR, Mazoocato TS, Franceschini S. Phenotype and genotype of Candida albicans strains isolated from pregnant women with recurrent vaginitis. Mycopathol 1997; 137: 87-94.
- Neely AN, Odds FC, Basatia BK, Holder IA. Characterization of Candida isolates from pediatric burn patients. J Clin Microbiol 1988; 26: 1645-9.