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# SGK1 (glucose transport), dishevelled2 (wnt signaling), LC3/p62 (autophagy) and p53 (apoptosis) proteins are unaltered in Lafora disease

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**Graphical Abstract:** Multiple disparate functions have been attributed to the Lafora disease gene products laforin and malin with separate hypotheses of pathogenesis of neurotoxic polyglucosans (Lafora bodies) and neurodegeneration in this disease. We tested four of these hypotheses in Lafora disease mouse models. We were unable to confirm these hypotheses.



**Abstract**: Glycogen forms through the concerted actions of glycogen synthase (GS) which elongates glycogen strands, and glycogen branching enzyme (GBE). Lafora disease (LD) is a fatal neurodegenerative epilepsy that results from neuronal accumulation of hyperphosphorylated glycogen with excessively long strands (called polyglucosans). There is no GBE deficiency in LD. Instead, the disease is caused by loss-of-function mutations in the *EPM2A* or *EPM2B* genes, encoding, respectively, a phosphatase, laforin, and an E3 ubiquitin ligase, malin. A number of experimentally derived hypotheses have been published to explain LD, including: *The SGK1 hypothesis* - Phosphorylated SGK1 (pSGK1) raises cellular glucose uptake and levels, which would activate GS. Based on observing increased pSGK1 in LD mice it was proposed that raised pSGK1 leads to polyglucosan generation through GS hyperactivation. *The Dishevelled2 hypothesis* - Downregulating malin in cell culture was reported to increase levels of dishevelled2, which through the wnt/glycogen synthase kinase-3 pathway would likewise overactivate GS. *The Autophagic defect hypothesis* - Polyglucosans may be natural

byproducts of normal glycogen metabolism. LD mice were reported to be autophagy-defective. LD would arise from failed autophagy leading to failed polyglucosan clearance. Finally, *the p53 hypothesis* - laforin and malin were reported to downregulate p53, their absence leading to increased p53, which would activate apoptosis, leading to the neurodegeneration of LD. In the present work we repeat key experiments that underlie these four hypotheses. We are unable to confirm increased pSGK1, dishevelled2, or p53 in LD mice, nor the reported autophagic defects. Our work does not support the above hypotheses in understanding this unique and severe form of epilepsy.

Keywords: Lafora Disease; polyglucosan; glucose transport; Wnt pathway; autophagy; apoptosis

## Introduction

Glycogen is synthesized by glycogen synthase (GS) and glycogen branching enzyme (GBE). GS activity is downregulated by phosphorylation, mainly by glycogen synthase kinase-3 (GSK3), and allosterically activated by the main intracellular form of glucose, glucose 6-phosphate (G6P). GBE deficiency results in an abnormal form of glycogen (called polyglucosan) which is poorly branched (i.e. has chains that are too long) and which precipitates, accumulates, and in neurons clogs axons causing Adult Polyglucosan Body Disease (which clinically resembles amyotrophic lateral sclerosis). GS overactivity likewise generates polyglucosans. In horses, an overactivating muscle GS gene mutation, and in mice, transgenic overexpression of muscle GS, lead to muscle polyglucosan accumulations. In humans, deficiency of the irreversible glycolytic enzyme muscle phosphofructokinase impairs glycolysis, which raises G6P levels, which hyperactivates GS and leads to muscle polyglucosans and disease<sup>1</sup>.

Lafora disease (LD) is an epilepsy that onsets in teenagers, progresses to intractability, and gradually to death within 10 years<sup>2</sup>. Pathologically, LD is characterized by gradual accumulation in neuronal somatodendritic compartments of masses of polyglucosans (Lafora bodies; LB) that have the particularity of being hyperphosphorylated<sup>3</sup>. LB are most likely pathogenic: it was shown that preventing their formation through knockout (ko) or downregulation of GS in LD mouse models eliminates LB and rescues the disease<sup>4-6</sup>. LD is caused by loss-of-function mutations of the EPM2A or EPM2B genes encoding the interacting laforin and malin enzymes respectively<sup>7, 8</sup>. Laforin is a glycogen phosphatase, and malin a ubiquitin E3 ligase whose ubiquitination targets, at least in cell culture experiments, include  $GS^{3, 7-11}$ . Multiple hypotheses have been advanced to explain LD, including: (I) Laforin deficiency leads to glycogen hyperphosphorylation, which, through unknown mechanisms, results in conversion of normal glycogen to polyglucosan<sup>3</sup>. (II) Malin deficiency

leads to reduced ubiquitin-mediated proteasomal degradation of GS, and the resultant increased GS drives polyglucosan generation<sup>9, 11</sup>. (III) Laforin or malin deficiency lead, through unknown mechanisms, to increased phosphorylated serum/glucocorticoid-induced kinase-1 (pSGK1), which increases glucose uptake, potentially hyperactivating GS to drive polyglucosan formation<sup>12</sup>. (IV) Malin deficiency upregulates dishevelled2, which upregulates the wnt pathway, which downregulates GSK3, which would upregulate GS, leading to  $polyglucosans^{13}$ . (V) Polyglucosans may be byproducts of normal glycogen metabolism, which would need to be cleared, e.g. by autophagy. Laforin or malin deficiency downregulates autophagy, which would prevent clearance of polyglucosans and allow their accumulation<sup>14-16</sup>. (VI) Laforin or malin deficiency upregulates p53, which is proapoptotic and would lead to neurodegeneration<sup>17</sup>.

The first two hypotheses garnered the greatest attention because of the direct mechanistic connections between the disease gene products and corresponding substrates, glycogen for laforin's phosphatase activity and GS for malin ubiquitination. This led to substantial new glycogen metabolism knowledge, including the specific locations of phosphorylation in glycogen<sup>18, 19</sup>. However, both hypotheses were undermined by recent results. It was shown that expressing a laforin mutant lacking the laforin phosphatase activity in laforin ko mice rescues the disease, including eliminating LB<sup>20</sup>, suggesting that laforin's glycogen phosphatase function is dispensable, and some other laforin function is critical in LD. Secondly, it was shown that laforin or malin ko mice do not have increased amounts of soluble GS or GS activity<sup>3, 21, 22</sup>, bringing into question the relevance of the malin action on GS seen in cell culture experiments. Given these new weaknesses of the leading hypotheses we decided to take a fresh look at the other four, and as a first step repeated experiments that had been fundamental to generating each of them, and performed new experiments. In each case we were unable to replicate critical experiments, or obtain new results contradicting the proposed hypothesis, which we present below.

### Results

Hypothesis III (SGK1) - Authors of this hypothesis showed that in cell culture pSGK1 increases following shRNA knockdown of laforin or malin. They then tested whether pSGK1 is increased in laforin ko LD mice and reported this to be the case. Specifically, they show a Western blot of SGK1 and pSGK1 and quantify the pSGK1/SGK1 ratio, which is seen to be twofold increased in the ko mice compared to wild-type  $(wt)^{12}$ . We reproduce their figure of this result in Figure 1A and note that: (1) They studied only laforin (and not malin) ko mice. (2) While there is variance in the measurements of the ko results, no variance is shown in the wt result (n=2 for each genotype). (3) They analyzed only skeletal muscle (and not brain). (4) They do not state the age of the mice, which is important given the progressive nature of the disease (for a finding to be pathogenic it should be present prior to disease onset). We performed SGK1 and pSGK1 Western blots on extracts from skeletal muscle and brain from 1 mo laforin<sup>23</sup> and malin<sup>24</sup> ko mice (LB are just starting to form at this age). We used the same antibodies as the authors of the hypothesis and the same laforin ko mice. We find no differences between wt and the ko animals (Figure 1B).



**Figure 1.** SGK1 and pSGK1 are not increased in LD mice. (A) Results from Singh et al. reporting increased pSGK1 in laforin knockout mice. (B) Current results do not confirm elevated pSGK1. Anti-SGK1 and anti– phospho (Thr-256) SGK1 were purchased from Millipore, and the secondary antibody is from Santa Cruz Biotechnology. WT, wild-type; Mko, malin knockout; Lko, laforin knockout. One month-old brain and muscle tissue lysates were used, respectively. The two samples from the same genotype are from two different animals. The Western blot experiment was repeated at least three times. All Western blot procedures are as described in Turnbull et al, Annals of Neurology, 2010; 68:925-933. Briefly, 40  $\mu$ g of total proteins was applied to 10% SDS-PAGE, and transferred onto nitrocellulose membrane. After blocking with 5% milk in 1× Tris-buffered saline and

Tween 20, the membrane was probed with primary antibody at  $4^{\circ}$ C for overnight, followed by secondary antibody conjugated with horseradish peroxidase as recommended by the manufacturer. Proteins were visualized using a chemiluminescent detection kit.

*Hypothesis IV (Dishevelled2)* - Noting interaction between malin and dishevelled2 in a yeast 2-hybrid screen, authors of this hypothesis overexpressed malin in cell culture and showed that this results in ubiquitination and degradation of dishevelled2<sup>13</sup>. We tested whether dishevelled2 is reduced in malin ko animals and found this not to be the case (Figure 2).



**Figure 2.** Dishevelled2 (Dvl2) is not increased in LD mice, at both 1 month and 10 months of age. Lanes 1 and 10 in the left panel confirm specificity of the antibody (Dvl2 antibody from Cell Signaling); lane 1, non-activated Dvl2 (MDA-MB-123 cells grown in RPMI-1640 and 5% FBS); lane 10, activated Dvl2 (same cells grown in Wnt-3a conditioned media for 3 hours. Bao et al: PloS One 2012; 7(11):e48670). One month-old and 10 month-old muscle tissues, from wild-type, Mko and Lko mice, were used, respectively.

*Hypothesis V (Autophagy)* - When autophagy is active, the LC3 protein is lipidated (LC3-II) and associated with autophagosomes. LC3-II is the most commonly used autophagy marker, its increase indicating increased autophagy and decrease the opposite. Two groups reported decreased LC3-II in cell lines from laforin ko mice and LD patients<sup>14, 16</sup>. One reported reduced LC3-II in laforin ko and malin ko mouse brains. A second autophagy marker, p62, was also affected. Surprisingly, additional experiments showed that the apparent autophagy defect in laforin ko mice was mTOR-dependent, while in malin ko mice it was not <sup>15</sup>. We performed LC3 and p62 Western blots in skeletal muscle and brain from laforin and malin ko mice and observe no changes in either (Figure 3).



Figure 3. LC3-II and p62 are unchanged in LD mice (anti-LC3 antibody from Novus Biologicals, and anti-p62 from Sigma). One month-old brain and muscle tissue lysates from wild-type, Mko and Lko mice, were used, respectively.

*Hypothesis VI (p53)* – Based on cell culture experiments, and observations in laforin and malin ko mouse, the authors concluded that laforin or malin deficiency lead to increased levels of p53, which leads to apoptosis, which could explain the neurodegeneration of  $LD^{17}$ . We cannot confirm increased p53 levels in laforin or malin ko mice (Figure 4).



**Figure 4.** p53 protein level is unchanged in LD mice (anti-p53 antibody from Santa Cruz Biotechnology). Western blot results for anti-p53 antibody were obtained from total protein and soluble protein of wild-type, Mko and Lko mice, respectively.

Conversion of glycogen to toxic polyglucosans in diseases associated with GBE deficiency the or phosphofructokinase deficiency is readily explained by classical glycogen metabolism. This is not so in LD, where heretofore veiled and until recently unsuspected facets of glycogen metabolism need to be uncovered. Identification of the LD genes and enzymatic activities of their products, glycogen dephosphorylation (laforin) and GS ubiquitination (malin), were critical steps forward and gave rise to the 'glycogen leading hyperphosphorylation' and 'GS hyperactivation' hypotheses of LD. As mentioned, recent observations weakened these hypotheses and led us in this work to revisit four others. We are unable to confirm in LD mouse models the disturbance in dishevelled2 suggested by cell culture overexpression experiments and are unable to replicate results critical to the SGK1, autophagy, and increased p53-apoptosis hypotheses. A possible explanation for the difference between our mouse data with previous results is murine background. Our malin ko mice<sup>24</sup> were generated separately from others<sup>15</sup>. The laforin ko mice, on

the other hand, are the same in all LD labs<sup>23</sup>, although they have been available in the different labs for over a decade and might have diverged in genetic background. The 'defective autophagy' hypothesis is additionally indirectly contradicted by the following. One of the groups that advanced this hypothesis found a disturbance in LC3 in cell lines but themselves could not confirm this in mouse brain<sup>16</sup>. Secondly, a recent experiment showed that what disturbances there are in markers of autophagy in LD mice disappear with GS downregulation. In this experiment, malin ko mice were bred with GS ko mice to eventually generate malin ko animals heterozygous for GS (GS activity is reduced by 50% in these mice) or completely lacking GS. In both situations, LB were dramatically reduced, the LD neurological phenotype was completely rescued, and what autophagic marker dysregulations there were corrected<sup>4</sup>, confirming that autophagic disturbance in LD, if or when present, is secondary to the disturbance in glycogen metabolism, and is not a primary cause of LD<sup>4</sup>.

But what then are the early pathogenic steps linking laforin or malin deficiency to polyglucosan formation and There are additional LD hypotheses invoking LD? laforin/malin actions via the neurodevelopmental protein neuronatin<sup>25</sup> and others, all of which will need to be revisited. However, in our view, the original two major hypotheses should continue to be explored. After all, laforin is a confirmed glycogen phosphatase, glycogen in both laforin and malin ko mice is hyperphosphorylated, and this hyperphosphorylation correlates with polyglucosan accumulation and LB formation <sup>3, 18, 19, 21, 24</sup>. As for malin, it is a confirmed ubiquitin E3 ligase whose substrates include GS, hyperactivity of which is a long-established cause of polyglucosan formation in phosphofructokinase deficiency and in various natural or engineered models of polyglucosan diseases<sup>1</sup>. We anticipate that future experiments will mitigate the recent observations weakening these gene function-based hypotheses, and will, in ways we cannot presently see, bring them together to explain the toxic glycogen structural transformation underlying LD.

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#### References

1. Hedberg-Oldfors, C. & Oldfors, A. (2015). Polyglucosan storage myopathies. Molecular aspects of medicine.

2. Minassian, B. A. (2001). Lafora's disease: towards a clinical, pathologic, and molecular synthesis. Pediatr Neurol, 25:21-9.

3. Tagliabracci, V. S., Girard, J. M., Segvich, D., Meyer, C., Turnbull, J., Zhao, X., Minassian, B. A., Depaoli-Roach, A. A. & Roach, P. J. (2008). Abnormal metabolism of glycogen phosphate as a cause for Lafora disease. The Journal of biological chemistry, 283:33816-25.

4. Duran, J., Gruart, A., Garcia-Rocha, M., Delgado-Garcia, J. M. & Guinovart, J. J. (2014). Glycogen accumulation underlies neurodegeneration and autophagy impairment in Lafora disease. Human molecular genetics, 23:3147-56.

5. Turnbull, J., DePaoli-Roach, A. A., Zhao, X., Cortez, M. A., Pencea, N., Tiberia, E., Piliguian, M., Roach, P. J., Wang, P., Ackerley, C. A. & Minassian, B. A. (2011). PTG depletion removes Lafora bodies and rescues the fatal epilepsy of Lafora disease. PLoS Genet, 7:e1002037.

6. Turnbull, J., Epp, J. R., Goldsmith, D., Zhao, X., Pencea, N., Wang, P., Frankland, P. W., Ackerley, C. A. & Minassian, B. A. (2014). PTG protein depletion rescues malin-deficient Lafora disease in mouse. Annals of neurology, 75:442-6.

7. Minassian, B. A., Lee, J. R., Herbrick, J. A., Huizenga, J., Soder, S., Mungall, A. J., Dunham, I., Gardner, R., Fong, C. Y., Carpenter, S., Jardim, L., Satishchandra, P., Andermann, E., Snead, O. C., 3rd, Lopes-Cendes, I., Tsui, L. C., Delgado-Escueta, A. V., Rouleau, G. A. & Scherer, S. W. (1998). Mutations in a gene encoding a novel protein tyrosine phosphatase cause progressive myoclonus epilepsy. Nat Genet, 20:171-4.

8. Chan, E. M., Young, E. J., Ianzano, L., Munteanu, I., Zhao, X., Christopoulos, C. C., Avanzini, G., Elia, M., Ackerley, C. A., Jovic, N. J., Bohlega, S., Andermann, E., Rouleau, G. A., Delgado-Escueta, A. V., Minassian, B. A. & Scherer, S. W. (2003). Mutations in NHLRC1 cause progressive myoclonus epilepsy. Nat Genet, 35:125-7.

9. Vilchez, D., Ros, S., Cifuentes, D., Pujadas, L., Valles, J., Garcia-Fojeda, B., Criado-Garcia, O., Fernandez-Sanchez, E., Medrano-Fernandez, I., Dominguez, J., Garcia-Rocha, M., Soriano, E., Rodriguez de Cordoba, S. & Guinovart, J. J. (2007). Mechanism suppressing glycogen synthesis in neurons and its demise in progressive myoclonus epilepsy. Nature neuroscience, 10:1407-13.

10. Worby, C. A., Gentry, M. S. & Dixon, J. E. (2006). Laforin, a dual specificity phosphatase that dephosphorylates complex carbohydrates. J Biol Chem, 281:30412-8.

11. Worby, C. A., Gentry, M. S. & Dixon, J. E. (2008). Malin decreases glycogen accumulation by promoting the degradation of protein targeting to glycogen (PTG). J Biol Chem, 283:4069-76.

12. Singh, P. K., Singh, S. & Ganesh, S. (2013). Activation of serum/glucocorticoid-induced kinase 1 (SGK1) underlies increased glycogen levels, mTOR activation, and autophagy defects in Lafora disease. Mol Biol Cell, 24:3776-86.

13. Sharma, J., Mulherkar, S., Mukherjee, D. & Jana, N. R. (2012). Malin regulates Wnt signaling pathway through degradation of dishevelled2. J Biol Chem, 287:6830-9.

14. Aguado, C., Sarkar, S., Korolchuk, V. I., Criado, O., Vernia, S., Boya, P., Sanz, P., de Cordoba, S. R., Knecht, E. & Rubinsztein, D. C. (2010). Laforin, the most common protein mutated in Lafora disease, regulates autophagy. Human molecular genetics, 19:2867-76.

15. Criado, O., Aguado, C., Gayarre, J., Duran-Trio, L., Garcia-Cabrero, A. M., Vernia, S., San Millan, B., Heredia, M., Roma-Mateo, C., Mouron, S., Juana-Lopez, L., Dominguez, M., Navarro, C., Serratosa, J. M., Sanchez, M., Sanz, P., Bovolenta, P., Knecht, E. & Rodriguez de Cordoba, S. (2012). Lafora bodies and neurological defects in malindeficient mice correlate with impaired autophagy. Human molecular genetics, 21:1521-33.

16. Puri, R., Suzuki, T., Yamakawa, K. & Ganesh, S. (2012). Dysfunctions in endosomal-lysosomal and autophagy pathways underlie neuropathology in a mouse model for Lafora disease. Human molecular genetics, 21:175-84.

17. Upadhyay, M., Gupta, S., Bhadauriya, P. & Ganesh, S. (2015). Lafora disease proteins laforin and malin negatively regulate the HIPK2-p53 cell death pathway. Biochemical and biophysical research communications, 464:106-11.

18. Nitschke, F., Wang, P., Schmieder, P., Girard, J. M., Awrey, D. E., Wang, T., Israelian, J., Zhao, X., Turnbull, J., Heydenreich, M., Kleinpeter, E., Steup, M. & Minassian, B. A. (2013). Hyperphosphorylation of glucosyl C6 carbons and altered structure of glycogen in the neurodegenerative epilepsy Lafora disease. Cell Metab, 17:756-67.

19. Tagliabracci, V. S., Heiss, C., Karthik, C., Contreras, C. J., Glushka, J., Ishihara, M., Azadi, P., Hurley, T. D., DePaoli-Roach, A. A. & Roach, P. J. (2011). Phosphate incorporation during glycogen synthesis and Lafora disease. Cell Metab, 13:274-82.

20. Gayarre, J., Duran-Trio, L., Criado Garcia, O., Aguado, C., Juana-Lopez, L., Crespo, I., Knecht, E., Bovolenta, P. & Rodriguez de Cordoba, S. (2014). The phosphatase activity of laforin is dispensable to rescue Epm2a-/- mice from Lafora disease. Brain, 137:806-18.

21. DePaoli-Roach, A. A., Tagliabracci, V. S., Segvich, D. M., Meyer, C. M., Irimia, J. M. & Roach, P. J. (2010). Genetic depletion of the malin E3 ubiquitin ligase in mice leads to lafora bodies and the accumulation of insoluble laforin. The Journal of biological chemistry, 285:25372-81.

22. Tagliabracci, V. S., Turnbull, J., Wang, W., Girard, J. M., Zhao, X., Skurat, A. V., Delgado-Escueta, A. V., Minassian, B. A., Depaoli-Roach, A. A. & Roach, P. J. (2007). Laforin is a glycogen phosphatase, deficiency of which leads to elevated phosphorylation of glycogen in vivo. Proceedings of the National Academy of Sciences of the United States of America, 104:19262-6.

23. Ganesh, S., Delgado-Escueta, A. V., Sakamoto, T., Avila, M. R., Machado-Salas, J., Hoshii, Y., Akagi, T., Gomi, H., Suzuki, T., Amano, K., Agarwala, K. L., Hasegawa, Y., Bai, D. S., Ishihara, T., Hashikawa, T., Itohara, S., Cornford, E. M., Niki, H. & Yamakawa, K. (2002). Targeted disruption of the Epm2a gene causes formation of Lafora inclusion bodies, neurodegeneration, ataxia, myoclonus epilepsy and impaired behavioral response in mice. Human molecular genetics, 11:1251-62.

24. Turnbull, J., Wang, P., Girard, J. M., Ruggieri, A., Wang, T. J., Draginov, A. G., Kameka, A. P., Pencea, N., Zhao, X., Ackerley, C. A. & Minassian, B. A. (2010). Glycogen hyperphosphorylation underlies lafora body formation. Annals of neurology, 68:925-33.

25. Sharma, J., Mukherjee, D., Rao, S. N., Iyengar, S., Shankar, S. K., Satishchandra, P. & Jana, N. R. (2013). Neuronatin-mediated aberrant calcium signaling and endoplasmic reticulum stress underlie neuropathology in Lafora disease. The Journal of biological chemistry, 288:9482-90.