# Formulation and Investigation of Lacidipine as a Nanoemulsions Rajaa A. Dahash<sup>\*,1</sup> and Nawal A. Rajab<sup>\*\*</sup>

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### Abstract

Lacidipine (LCDP) is a calcium-channel blocker with low aqueous solubility and bioavailability. Nanoemulsion (NE) is one of the popular methods that has been used to solve the solubility problems of many drugs. LCDP was formulated as a NE utilizing triacetin as an oil phase, tween 80 and tween 60 as surfactants and ethanol as a co-surfactant. Nine formulas were prepared, and different tests performed to ensure the stability of the NEs, such as thermodynamic stability, particle size, polydispersity index, zeta potential, dilution test, conductivity test, drug content, viscosity and *in-vitro* drug release. Results of characterization showed that LCDP NE (F-5) using triacetin, tween80, ethanol and DDW in a ratio of (10:60:30) was selected as the best formula, since it has excellent thermodynamic stability with a particle size of 13.42, low PDI 0.234, zeta potential (-14.5mV), good dilution without drug precipitation, efficient electrical conductivity 0.241ms/cm, higher percent of drug content (99.14%) with acceptable viscosity, and complete release of the drug after (30 min.) with significantly higher (P<0.05) dissolution rate in comparison with pure drug powder.

The selected formula (F-5) subjected to further investigations as drug and excipient compatibility study by Fourier transform infrared spectroscopy (FTIR) and high performance liquid chromatography (HPLC) and Atomic force microscope (AFM).

The outcomes of the (FTIR) explain that the distinctive peaks for LCDP were displayed the same functional group's band with very slight shifting, which suggests the presence of hydrogen bonding. This indicates that there was no interaction between LCDP and other NE components, while the HPLC demonstrated that was no change in retention time and no extra peaks reported. Therefore, these excipients were found to be compatible with LCDP.

In conclusion, the NE was found to be an efficient method to enhance the solubility and dissolution rate of drugs that have poor water solubility (lipophilic drugs).

Keyword: Lacidipine, , Triacetin, Tween 80, Tween60 , Nanoemulsion .

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#### الخلاصة

ان العديد من المركبات الصيدلانية الفعالة لديها مشاكل في الذوبان حتى الأن، والتي اصبحت عقبة رئيسية تقيد استخدامها في المستحضرات الصيدلانية. ان عقار اللاسيدبين هو مانع لقنوات الكالسيوم له ذوبان مائي وتوافر حيوي منخفض جدا.

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### Introduction

Oral delivery of drugs is regarded as the optimal route to achieve therapeutic and prophylactic effects against various diseases, especially chronic conditions. It may have poor bioavailability as a hurdle, leading to challenges for pharmaceutical manufacturers to design delivery system(s) that can provide improved pharmacokinetic profiles and hence therapeutic responses <sup>(1,2)</sup>.

The term nanoemulsion (NE) refers to a thermodynamically stable isotropically clear dispersion of two immiscible liquids, such as oil and water, stabilized by an interfacial film of surfactant molecules. Their size varies from 10 to 1000 nm.  $^{(3,4)}$ .

Lacidipine (LCDP) is а dihydropyridine calcium-channel blocker developed for oral administration .It used in the treatment of hypertension and atherosclerosis and possessed an antioxidant effect <sup>(5)</sup>. Chemical of LCDP is (E)-4-[2-[3-(1,1name Dimethylethoxy) -3-oxo-1-propenyl] phenyl]-1,4-dihydro -2,6-dimethyl -3,5-pyridine dicarboxylic acid diethyl ester, The molecular weight (455.5) and pka <sup>v</sup>.5 , log P (octanol/water) is 5.°, the powder is a white to pale yellow powder, melt at  $17^{\circ}C^{(6,7)}$ , and the chemical structure is shown in figure(1)  $^{(8)}$ .



# Figure 1. The chemical structure of LCDP $_{\scriptscriptstyle (8)}$

LCDP poorly absorbed from the gastrointestinal tract after oral doses and undergoes extensive first-pass metabolism the bioavailability has been reported to be 2 to 10%. The rate limiting step for drug absorption in this class is dissolution <sup>(9)</sup>.

This study aims to prepare LCDP as a nanoemulsion to improve its dissolution rate.

## **Materials and Methods**

#### Materials

LCDP was obtained from Baoji Guokang Bio-Technology Co., Ltd, China, triacetin was purchased from Hyper-Chem LTD CO, China, tween 20 obtained from HiMedia Chemicals, India, tween 60 was obtained from Avonchem, England, tween 80 was purchased from Riedel-De-Haen, Germany, Ethanol was purchased from Sigma-Aldrich, Germany, Hydrochloric acid was purchased from Thomas Baker, India . All other chemicals were of analytical grade.

## Methods

Characterization of LCDP

Differential scanning calorimetry analysis (DSC)

The DSC study was performed for the pure drug to evaluate the thermotropic properties and thermal behavior of the LCDP.

Approximate weighed samples (about 3 mg) were put in sealed aluminium pans and warmed at a scanning rate of  $10^{\circ}$ C/min<sup>(11)</sup>.

## Saturation solubility study of LCDP

Saturated solubility of LCDP was estimated in various oils, surfactants, co-surfactants and dissolution media.

The measurement of solubility was done as follows: The excess amount of LCDP was added to (5 mL) of each selected individual oils, surfactants and co-surfactants contained in stoppered vials separately, then shaken utilizing a water bath shaker at  $25\pm1^{\circ}$ C for 72 hours to prepare a saturated solution. After accomplishing the equilibrium, the mixtures were centrifuged at 3000rpm for 15min, followed by filtration through a 0.45-micrometer millipore filter. Samples were suitably diluted with ethanol and analyzed by UV/Vis spectrophotometer at  $\lambda$  max of LCDP. The measurements were done in triplicate <sup>(12,13)</sup>.

# Construction of pseudo-ternary phase diagrams

The aqueous titration method was utilized to construct the pseudo-ternary phase diagram. Based on the solubility studies, triacetin was selected as an oil phase, tween 80 and tween 60 were selected as surfactant and ethanol were selected as a co-surfactant, and deionized water (DDW) used as an aqueous phase. The oil: surfactant:co-surfactant (Smix) mixed at different ratios ranging from (1:9 to 9:1). Smix ratios was 1:3, 1:2, 1:1, 2:1 and 3:1 for Smix (tween 80/ethanol) and 1:2, 1:1, 2:1 and 3:1 for Smix tween 60/ethanol <sup>(14,15)</sup>.

### Preparation of LCDP nanoemulsion

Different o/w NE formulations (table 1) were prepared using the Smix and oil concentrations according to pseudo-ternary phase diagrams; primary LCDP emulsion was prepared via dissolving (2 mg) of the drug in the selected oil. The magnetic stirrer used to ensure complete mixing then the selected Smix added slowly in a fixed ratio till the clear solution was obtained.

DDW added dropwise to the clear solution with the continuous stirring at (~500 rpm ) at room temperature until the formation

of a clear emulsion. After that, the prepared emulsions were ultrasonicated via utilizing a 20 kHz sonicator for 10 min  $^{(16,17)}$ .

NE-F	Triacetin	Surfactant	Co-	Smix ratio	Smix	DDW
	%W/W		surfactant		% W/W	%W/W
F-1	10	Tween 80	Ethanol	1:3	60	30
F-2	10	Tween 80	Ethanol	1:2	60	30
F-3	10	Tween 80	Ethanol	1:1	60	30
F-4	10	Tween 80	Ethanol	2:1	60	30
F-5	10	Tween 80	Ethanol	3:1	60	30
F-6	10	Tween 60	Ethanol	1:2	60	30
F-7	10	Tween 60	Ethanol	1:1	60	30
F-8	10	Tween 60	Ethanol	2:1	60	30
F-9	10	Tween 60	Ethanol	3:1	60	30

#### LCDP nanoemulsion characterization Visual transparency

Optical observation for NE formulas was intent utilizing good light source for transparency and flow ability <sup>(18)</sup>.

### Thermodynamic study

*I. Centrifugation study*: In this study, formulas were centrifuged at 5000 rpm for 30 min and then checked for instability such as phase separation. The formulations that did not show any signs of instability were chosen for heating-cooling cycle <sup>(19)</sup>.

**II.** Heating-cooling cycles test: Stability of NE depends on the variation of temperature was studied by heating-cooling cycle. Formulations subjected to six cycles between refrigerator temperature 5 °C and at 50 °C storage at each temperature for not less than 48 hours <sup>(20)</sup>.

*III. Freezing–thawing test*: This test was done by exposing the formulations for two different temperatures which are  $(-21^{\circ}C)$  and  $(25^{\circ}C)$ using refrigerator and the time for each temperature not less than 24 hours <sup>(21)</sup>.

### Droplet size measurement

The droplet size of NE was established by analyzing the fluctuations in light scattering due to the Brownian motion of the particle utilizing dynamic light scattering technique (Zetasizer Nano ZS, Malvern, UK)<sup>(22)</sup>.

#### Polydispersity index measurement (PDI)

The estimation of the (PDI) gives information about the uniformity of droplet size within the formulated NE. The lower PDI value (near zero) indicates a monodisperse droplet population <sup>(23)</sup>.

#### Zeta potential measurement (3 – potential)

The droplet charge (zeta potential) of the NEs was determined to utilize dynamic light scattering technique, Zeta potential was believed to be sufficient for ensuring the physical stability of NEs <sup>(24)</sup>.

#### **Dilution** test

The aqueous dilution test was performed, one mL of each NE formulas (F1-F9) diluted to 50 mL,100 mL and 500 mL with distilled water at 25°C with constant stirring at 50 rpm and observed visually for turbidity, clarity and phase separation <sup>(25)</sup>.

#### Conductance measurement

The o/w NEs are highly conducting since the water is the external phase, whereas w/o NEs are not conducting as they have water in the internal phase <sup>(26)</sup>.

#### Drug content estimation

Accurately 10 ml of each NE formula which contains (2 mg) was diluted with ethanol to the sign (100 mL) in a volumetric flask and subjected to the centrifugation for 15 minutes (3000 rpm), then filtered using 0.45  $\mu$ m filter syringe and suitably diluted. Determination of the contents of LCDP NEs via utilizing UV/Vis spectrophotometer at the selected  $\lambda$  max <sup>(27,28)</sup>.

#### Viscosity measurement

 $\begin{array}{ccc} & \text{Determination} & \text{of} & \text{viscosities} \\ \text{determines whether the system is o/w or w/o} \\ \text{emulsion} \ ^{(29)}. \end{array}$ 

#### *In vitro* drug dissolution study

The in vitro release of LCDP loaded NE occurs using USP dissolution apparatus type-II. Dialysis bag (Molecular cut off 12000Da) was utilized. ten ml of each formula which contain 2mg of LCDP was put in the bag, and this bag was immersed in 500 ml of dissolution medium. The rotation speed was 50 rpm, and the dissolution medium was 0.1N HCl with 1% tween 20 at 37  $\pm$  0.5 °C <sup>(30)</sup>. Samples (5 ml) were withdrawn at a regular time intervals (5,10,15, 20,30, 40,50 and 60 min) from the dissolution medium and the samples then filtered by using through a 0.45 µm filter svringe and were analyzed by UV/Vis spectrophotometer at the  $\lambda$  max. of the drug <sup>(31)</sup>. Selection of the optimum formula

The choice of the optimum formula was accomplished, and this achieved according to the globule size analysis, PDI, zeta potential

measurements, electrical conductivity, viscosity, drug content and in vitro release studies. *Evaluation of the selected LCDP optimum* 

# Evaluation of the selected LCDP optimum formula

# Drug and excipient compatibility study by FTIR

To demonstrate any possible interaction between the drug and the utilized excipients in the selected formula. Samples were mixed with potassium bromide and pressed in the form of a disc; FTIR spectroscopy analyzed the disc from 4000-400 cm<sup>-1 (32)</sup>.

#### Validation of the HLPC method

HPLC method used in the to investigation and determine the possible interactions between oil, drug and other excipients. A waters HPLC system used which was equipped with a SPA-20A detector. The system was controlled through Breez software. The mobile phase which consisted of acetonitrile: water (65:35% v/v) with a flow rate of 1mL/min at ambient temperature and the injection volume was 10 µL.

The detective wave length was set at 239 nm. The mobile phase was filtered through  $(0.45 \mu m)$  in millipore solvent filtration apparatus before use (33).

#### Atomic Force Microscopy (AFM) Study

The AFM is capable of scanning the surfaces in controlled environmental conditions and can measure the particle size of nanoparticles accurately. AFM confirmed the size and surface morphology of LCDP nanoparticles after drying of the selected formulations. Droplets of optimized formulas were deposited on freshly cleaved mica and dried 15 minutes in the oven by the droplet evaporation technique. Particle size, 3Ddimension graph, and a histogram of particle size distribution were obtained <sup>(34)</sup>.

## **Results and Discussion**

# Differential scanning calorimetry analysis (DSC)

Pure LCDP powder showed a characteristic endothermic peak at (185.80 °C °C), such sharp endothermic peak signifies that drug used was in a pure crystalline state and it was near the reported one  $^{(35)}$ , as shown in figure 2.



Figure 2. Differential scanning calorimetry thermogram of LCDP.

#### Saturation solubility study of LCDP

As demonstrated in the table (2), higher solubility of LCDP was in triacetin while the lower solubility was in liquid paraffin. To ensure the drug is solubilized form, triacetin was utilized in the formulations, no precipitation of drug will occur since lipophilic drugs can be easily present in a solubilized form  $(^{36,37})$ . Regarding surfactants, tween 80 and tween 60 were chosen as a surfactant to obtain a one-phase clear solubilizing capacity for LCDP  $(^{39})$ .

Oil	Solubility(mg/ml)	SD	
Coconut oil	32.139	±0.65	
Corn oil	15.224	±0.27	
Grape seed oil	4.165	±0.32	
Lavender oil	6.175	±0.48	
Liquid paraffin	1.281	±0.89	
Oleic acid	58.126	±0.95	
Olive oil	16.366	±0.65	
Sunflower oil	6.156	±0.85	
Triacetin	63.137	±0.78	
	Surfactant		
Propylene glycol	18.221	±0.53	
Tween 80	41.156	±0.78	
Tween 60	39.187	±0.85	
Tween 20	35.043	±0.89	
Co-surfactant			
Ethanol	65.164	± 0.72	
Methanol	52.189	±0.96	
<b>PEG200</b>	46.232	±0.26	
<b>PEG400</b>	38.562	±0.87	

#### Table 2. Saturation solubility study of LCDP

# Construction of pseudo-ternary phase diagrams

Figures (3 and 4) showed the pseudoternary phase diagram for the o/w NEs using triacetin as an oil phase, tween 80 and tween60 as a surfactant and ethanol as a co-surfactant.



Figure 3.pseudo-ternary phase diagram o/w emulsion diagram using triacetin, tween 80: ethanol in different ratios.



Figure 4. Triangular co-ordinate o/w emulsion diagram using triacetin, tween60: ethanol in different ratios.

# *LCDP NE characterization thermodynamic study*

NEs are thermodynamically stable systems, formed of particular concentrations of oil, Smix and DDW with no phase separation and no cracking or creaming. Small droplet size prevents any flocculation, enabling the system to remain dispersed with no separation <sup>(40)</sup>, as shown in table (3).

Table 3. Results of thermodynamic stabilitystudies For LCDP nanoemulsions.

F-code	Centrifugation test	Heating- cooling cycles	Freeze- thawing cycles
NE-1	Pass	Pass	Pass
NE-2	Pass	Pass	Pass
NE-3	Pass	Pass	Pass
NE-4	Pass	Pass	Pass
NE-5	Pass	Pass	Pass
NE-6	Pass	Pass	Pass
NE-7	Pass	Pass	Pass
NE-8	Pass	Pass	Pass
NE-9	Pass	Pass	Pass

#### Droplet size

Table (4) showed the results of droplet size measurement. The results illustrated that when the concentration of surfactant increased the particles size reduced since this high surfactant concentration decreases surface tension and stabilizes newly developed surfaces during homogenization and production of smaller particles <sup>(41)</sup>. Figures 5 and 6 show the droplet size measurement of the LCDP NEs formulas 1-9.

Table 4. Particle size measurement forLCDP nanoemulsion.

F-	Mean	F-	Mean
Code	particle size	Code	particle size
	(nm)		(nm)
F-1	345.7	F-6	304.3
F-2	258.8	F-7	220.6
F-3	205.5	F-8	15.05
F-4	13.42	F-9	13.47
F-5	13.42		



Figure 5 . Particle size distribution of lacidipine NEs formulas (1, 2, 3,4,5 and 6) respectively



Figure 6. Particle size distribution of LCDP NEs formulas (7, 8 and 9) respectively.

#### Polydispersity index measurement (PDI)

PDI refers to the quality of the dispersion; this index represents uniformity and homogeneity of the particles in the NEs , as revealed in table (5) <sup>(35)</sup>.

Table 5.The Polydispersity index of LCDPnanoemulsions

F-Code	PDI	F-Code	PDI
F-1	0.581	F-6	0.261
F-2	0.269	F-7	0.362
F-3	0.370	F-8	0.410
F-4	0.381	F-9	0.186
	0.234		

#### Zeta potential measurement (3 – potential)

Zeta potential governs the degree of repulsion between adjacent, similarly charged, dispersed particles. When the nonionic surfactants adsorb onto the nanoscale droplets, they lowering the zeta potentials and preserve stability <sup>(42)</sup>. Table (6) and figures 7 and 8 show the values of the zeta potential of formulas 1-9 **Table6. Zeta potential of LCDP nanoemulsions** 

F- code	Zeta potential (mV)	F- code	Zeta potential (mV)
F-1	-3.84	F-6	-3.04
F-2	-5.4	F-7	-10.8
F-3	-9.62	F-8	-11.8
F-4	-6.45	F-9	-3.84
F-5	-14.5		



Figure 7. Zeta potential values of lacidipine NEs formulas (1,2,3 and 4) respectively.



Figure 8. Zeta potential values of LCDP nanoemulsions formulas(5,6,7,8and 9) respectively .

#### Dilution test

Dilution test confirmed the high physical stability of the LCDP NE under dilution with water. In less than 1 minute, all NE formulas (F1-F9) showed clear and fine bluish NE indicating o/w type, proved that they are maintaining the nanosized character and could be diluted in GI fluids without drug precipitation <sup>(43)</sup>.

### Conductance measurement

Electrical conductivity had a potent relation with the type or nature of the external phase of NE. Higher values for electrical conductivity indicate higher conductivity of water <sup>(44)</sup>. The results in a table (7) showed higher conductivity values.

Table 7. Conductivity measurement ofLCDP nanoemulsions.

F-	σ(ms/cm)	F-Code	σ(ms/cm)
Code			
F-1	0.267	F-6	0.123
F-2	0.143	F-7	0.156
F-3	0.154	F-8	0.0691
F-4	0.176	F-9	0.105
F-5	0.241		

#### Drug content estimation

All NEs formulas agreed with the requirements of the British Pharmacopeia range (95%-105%) as shown in table (8), which indicated that high content uniformity and revealed the adequacy of the preparation method <sup>(45)</sup>.

F- code	% Drug	SD
	content	
NE-1	99.14	±0.81
NE-2	96.29	±0.21
NE-3	98.03	±0.56
NE-4	96.15	±0.32
NE-5	99.14	±0.68
NE-6	99.26	±0.42
NE-7	99.16	±0.26
NE-8	96.03	±0.97
NE-9	97.48	±0.46

Table 8.Drug content of LCDP formulations (mean ±SD, n=3).

#### Viscosity measurement

From figure (9), it was demonstrated as the concentration of the surfactant increased, the viscosity increased this may be due to entrapping of the water molecules in cross-linking surfactants chains and also highest surfactant concentration would make the dispersion medium more rigid, as well as formulas that contain tween 60 has a higher viscosity than that contain tween 80 since tween 60 has a higher molecular weight than tween 80  $(^{46,47})$ .

The results also showed that the viscosity decreased as the rotation speed increased (shear rate) indicating the pseudoplastic (shear thinning liquids) flow of the preparation <sup>(48)</sup>.



Figure 9. Viscosities of LCDP nanoemulsions

#### In vitro drug dissolution study

The release of the drug from all NEs formulations was found nearly 100% at the end of 60 min.; higher the dissolution, faster the absorption, and hence quicker and higher the drug action can be obtained by smaller the particle size of a drug in the dosage forms (49). Figure (10) demonstrated that the release of LCDP from the formulas that contain tween 80 as a surfactant was higher than that contain tween 60 which could be explained by the smaller droplet size of formulas containing tween 80 as compared to that formula which contains tween 60 leading to a higher rate of dissolution. The higher HLB value of tween 80. which is 15 enhanced the continuous distribution and solubilization of the incorporated lipophilic drug within the system (50,51)



Figure 10.A comparative dissolution profile of lacidipine NEs (F- 2, F-6 and pure LCDP) in 500ml of 0.1 N HCl (with 1% tween 20) dissolution medium at 37  $^{\circ}$ C

#### Selection of the optimum formula

After studying the characterization of prepared LCDP NEs (F1-F9), it was found that (F-5) is selected as the best formula that is characterized by a low particle size (13.42), low polydispersity index (PDI) (0.234), zeta potential (-14.5), good spreadability on the filter paper, efficient electrical conductivity (0.241 ms/cm), good pH value (5.9), good percent of light transmittance (99.10), accepted viscosity, higher drug content percent (99.14) and higher dissolution rate. The optimized formula would be subjected to further studies. *Evaluation of the selected Lacidipine optimum formula* 

# Drug and excipient compatibility study by FTIR

FTIR is an extremely powerful technique in discovering and evaluating any possible chemical interaction between LCDP and any excipient during NEs preparation. FTIR spectra of pure LCDP powder showed characteristic peaks which are: 3348.78 cm<sup>-1</sup> due to (N–H) stretching vibration, 3109.65–2976.59 cm<sup>-1</sup> corresponding to (=C–H) stretching, 2930.31 cm<sup>-1</sup> due to aliphatic (C–H) stretching, 1674.87 cm<sup>-1</sup> for ester (C=O) stretching, 1495.53 cm<sup>-1</sup> and 1451.17 cm<sup>-1</sup> due to aromatic –C=C stretching, 1372.10 cm<sup>-1</sup> corresponding to aliphatic C–H bending, 745.35 cm<sup>-1</sup> for disubstitued ortho benzene stretching and 982.55 cm<sup>-1</sup> for C–N stretching. The FTIR spectrum of pure LCDP and selected formula (F-5) displayed the same functional groups band with very slight shifting, which suggests the presence of hydrogen bonding <sup>(52,53)</sup>. Figures 11 and 12 showed the FTIR spectra of the LCDP and the selected formula (F-5) respectively.



Figure11.The FTIR spectrum of LCDP.



Figure 12.FTIR spectrum of the selected formula (F-5).

#### Validation of the HLPC method

The chromatograms of Pure drug LCDP and LCDP in the selected formula (F-5) there was no change in retention time, and no extra peaks reported. Therefore, these

excipients were found to be compatible with LCDP  $^{(54)}$ . Figures (13 and 14) showed the Chromatograms of LCDP, triacetin, tween80 ethanol and F5 respectively.



Figure 13. Chromatograms of pure LCDP and triacetin, respectively.



Continue figure 13. Chromatograms of pure Lacidipine and triacetin, respectively.



Figure 14. Chromatograms of tween80 and F-5 respectively

### Atomic Force Microscopy (AFM) Study

AFM is capable of scanning and measures the properties and characteristic of the surfaces. With the high accuracy of the AFM, it is possible to determine the dimensions of nanoparticles with high reliability. The morphological analysis and particle size of formula F5 performed by AFM were close to spherical in shape and smooth surface <sup>(55,56)</sup>, as shown in figure 15.



Figure 15 The AFM image of LCDP NE (F5) where the scanning area is 2µm\*2µm **Conclusion** 

After discussing the previously obtained data, it is easy to deduce the following points:

- 1. All the NE formulas prepared with triacetin as an oil phase, tween 80 and tween 60 as a surfactant and ethanol as a co-surfactant with different Smix ratios provided a significant increase (P<0.05) in the dissolution rate compared to pure drug powder.
- 2. The formula (NE-5) with triacetin oil and mix (tween80: ethanol) in a ratio of (3:1) was selected as an optimum formula.
- **3.** The compatibility studies (FTIR and HPLC) for the selected formula revealed no specific interactions between LCDP and other excipients.

### References

- Mekhilef SF, A. Hussein A. Novel combination for self-nanoemulsifying drug delivery system of candesartan cilexetil. Iraqi J Pharm Sci. 2018; 27(2):123–34.
- 2. Sivapriya V, ponnarmadha S, Azeezand NA, Sudarshanadeepa V . Novel nanocarriers for ethnopharmacological formulations . Int J App Pharm.2018;10 (4) : 26-30 .
- **3.** Siya M, Sinai Kunde D, Bhilegaonkar S, Godbole AM, Pankaj Gajre M. Biopharmaceutical classification system: A brief account. Int J Res Methodol. 2015; 1(1):20–46.
- **4.** Taher MN, Hussein AA. Formulation and evaluation of domperidone nanoemulsions for oral rout. Iraqi J Pharm Sci. 2015; 24(2):77–90.
- 5. Fares AR, Elmeshad AN, Kassem MAA. Enhancement of dissolution and oral bioavailability of lacidipine via pluronic P123 / F127 mixed polymeric micelles: formulation, optimization using central composite design and in vivo bioavailability study. Drug Delivery. 2018; 25(1):132–42.
- 6. Sandeep V, Narendar D, Arjun N, Kishan V. Lacidipine loaded solid lipid

nanoparticles for oral delivery: preparation, characterization and in vivo evaluation. Int. J. Pharm. Sci. Nanotechnol.2016; 9(6):3524–30.

- 7. Druzbicki K, Mielcarek J, Kiwilsza A, Toupet L, Collet E, Pajzderska A, et al. Computationally assisted (solid-state density functional theory) structural (Xray) and vibrational spectroscopy (FT-IR, FT-RS, TDs-THz) characterization of the cardiovascular drug lacidipine. Cryst Growth Des. 2015; 15(6):2817–30.
- 8. Sweetman SC. Martindale: the complete drug reference. 38th ed. London: Pharmaceutical press; 2014.Cardiovascular drugs. Volume 2.Monograph on drugs and ancillary substances .1417.
- **9.** Bhatt P, Madhav S. A detailed review on nanoemulsion drug delivery system. Int J Pharm Sci Res. 2011; 2(9):2292–8.
- **10.** Patel ZR, Patel HK, Trivedi HJ, Patel KN, Nayak BS. Enhancement of solubility and dissolution properties of lacidipine by solid dispersion. International journal of pharmaceutics and drug analysis. 2015; 3(5): 165-170.
- 11.Swain S, Patra CN, Rao ME. Pharmaceutical drug delivery systems and vehicles. New Delhi: Woodhead Publishing India Pvt. Ltd; 2016.Chapter 5. An overview of liquisolid technology. 157.
- **12.** Ali HH, Hussein AA. Oral nanoemulsions of candesartan cilexetil: formulation, characterization and in vitro drug release studies. AAPS. 2017; 3(1).
- **13.** Bhosale R, Bhandwalkar O, Duduskar A, Jadhav R, Pawar P. Water-soluble chitosan mediated voriconazole microemulsion as sustained carrier for ophthalmic application: Evaluations. Open Pharm Sci J. 2016; 3(1):215–34.
- **14.**Su R, Yang L, Wang Y, Yu S, Guo Y, Deng J, et al. Formulation, development, and optimization of a novel octyldodecanol-based nanoemulsion for transdermal delivery of ceramide IIIB. Int J Nanomedicine. 2017;12: 5203–21. 11
- **15.**Beg S, Jena SS, Patra CN, Rizwan M, Swain S, Sruti J, et al. Development of solid self-nanoemulsifying granules (SSNEGs) of ondansetron hydrochloride with enhanced bioavailability potential. Colloids Surfaces B Biointerfaces. 2013; 101:414–23.
- **16.** Miastkowska MA, Banach M, Pulit-Prociak J, Sikora ES, Głogowska A, Zielina M. Statistical analysis of optimal

ultrasound emulsification parameters in thistle-oil nanoemulsions. J. Surfactants Deterg. 2017; 20(1):233–46.

- **17.** O'Sullivan J, Murray B, Flynn C, Norton I. Comparison of batch and continuous ultrasonic emulsification processes. J Food Eng. 2015; 167:114–21.
- 18. Yadav V, Jadhav P, Kanase K, Bodhe A, Dombe S. Preparation and evaluation of microemulsion containing antihypertensive drug. Int. J. Appl. Pharm.2018; 10(5).
- **19.**Parmar N, Singla N, Amin S, Kohli K. Study of cosurfactant effect on nanoemulsifying area and development of lercanidipine loaded (SNEDDS) self nanoemulsifying drug delivery system. Colloids Surfaces B Biointerfaces .2011; 86(2):327–38.
- **20.** Acharya U. Formulation and evaluation of nano emulsion-based system for transdermal delivery of antipsoriatic drug. World J Pharm Pharm Sci. 2017; 6(7):732–48.
- **21.** Yasser M, Gad S, El-Sayed M, Ghorab M. The effect of converting liquid valsartan SNEDDS into solid SNEDDS using different solid carriers on its performance. International Journal of Biological & Pharmaceutical Research. 2013; 4(12):1015–26.
- **22.** Alshahrani SM. Anti-inflammatory studies of ostrich oil based nanoemulsion. Journal of Oleo Science.2019; 208(3):203–8.
- **23.**Siddique AB, Ebrahim H, Mohyeldin M, Qusa M, Batarseh Y, Fayyad A, et al. Novel liquid-liquid extraction and selfemulsion methods for simplified isolation of extra-virgin olive oil phenolics with emphasis on (-)-oleocanthal and its oral anti-breast cancer activity. PLoS One. 2019; 14(4).
- **24.** Oliveira AEMFM, Duarte JL, Cruz RAS, Da Conceição EC, Carvalho JCT, Fernandes CP. Utilization of dynamic light scattering to evaluate Pterodon emarginatus oleoresin-based nanoemulsion formation by non-heating and solvent-free method. Brazilian J Pharmacogn. 2017; 27(3):401–6.
- **25.** Maraie NK, Almajidi YQ. Application of nanoemulsion technology for preparation and evaluation of intranasal mucoadhesive nano- in-situ gel for ondansetron HCl. Journal of Global Pharma Technology. 2018; 10(03): 431- 42.
- **26.**De Azevedo Ribeiro RC, Barreto SMAG, Ostrosky EA, Da Rocha-Filho PA, Veríssimo LM, Ferrari M. Production and

characterization of cosmetic nanoemulsions containing opuntia ficusindica (L.) Mill extract as moisturizing agent. Molecules. 2015;20(2):2492–509.

- **27.** Ezealisiji KM, Mbah CJ, Osadebe P, Krause R. Pharmacokinetics studies of mirtazapine loaded nanoemulsion and its evaluation as transdermal delivery system. Chem Pharm Res .2017; 9(3):74–84.
- **28.**K. Gurpret, S. K. Singh. Review of nanoemulsion formulation and characterization techniques. Indian J Pharm Sci. 2018; 80(5):781–9.
- **29.**Subramanian N, Sharavanan SP, Chandrasekar P, Balakumar A, Moulik SP. Lacidipine self-nanoemulsifying drug delivery system for the enhancement of oral bioavailability. Arch Pharm Res. 2016; 39(4):481–91.
- **30.** Basalious EB, Shawky N, Badr-Eldin SM. SNEDDS containing bioenhancers for improvement of dissolution and oral absorption of lacidipine. I: Development and optimization. Int J Pharm. 2010; 391(1–2):203–11.
- **31.** Basalious EB, Shawky N, Badr-Eldin SM. SNEDDS containing bioenhancers for improvement of dissolution and oral absorption of lacidipine. I: Development and optimization. Int J Pharm. 2010; 391(1–2):203–11.
- **32.** Kumari S, Kumaraswamy R V, Choudhary RC, Sharma SS, Pal A, Raliya R, et al. Thymol nanoemulsion exhibits potential antibacterial activity against bacterial pustule disease and growth promotory effect on soybean. Sci Rep. 2018 ;(October 2017):1–12.
- **33.** Channabasavaraj KP, Nagaraju PT, T SKP, Reddy PS. Reverse phase HPLC method for determination of lacidipine in pharmaceutical preparations. Int. J. Pharm. Sci. Rev. Res. 2010; 5(2): 111-4.
- **34.** Ocwieja M, Morga M, Adamczyk Z. Selfassembled silver nanoparticles monolayers on mica- AFM, SEM, and electrokinetic characteristics. J Nanopart Res. 2013; 15: 1460.
- **35.** Mukharya A, Mansuri N, Chaudhary S, Misra A. Solid-state characterization of lacidipine/PVP K 29/32 solid dispersion primed by solvent co-evaporation. Int J Pharm Investig. 2012; 2(2):90.
- **36.** Shivasaraun U V, Sureshkumar R, Karthika C, Nethravathi P. Flavonoids as adjuvant in psoralen-based phytochemotherapy in the management of

vitiligo/leukoderma. Asian J. Pharm. 2019;13 (2):85–92.

- **37.**Balamohan P, Anjali CH, Ravindran A. Nanoemulsion: Synthesis, characterization and its applications. J Bionanoscience. 2013; 7(4):323–33.
- **38.** Chatterjee B, Hamed Almurisi S, Ahmed Mahdi Dukhan A, Mandal UK, Sengupta P. Controversies with self-emulsifying drug delivery system from pharmacokinetic point of view. Drug Deliv. 2016; 23(9):3639–52.
- **39.** Kesan K, Minyak F, Terhadap K, Mikroemulsi S. Study on the effect of oil phase and co-surfactant on microemulsion systems. Malaysian J Anal Sci. 2018; 21(6):1409–16.
- **40.** Hanifah M, Jufri M. Formulation and stability testing of nanoemulsion lotion containing centella asiatica extract. J Young Pharm. 2018; 10(4):404–8.
- **41.** Amaral-Machado L, Egito EST Do, De Souza Araujo AA, Alencar EDN, Rutckeviski R, Xavier FH, et al. Therapeutic bullfrog oil-based nanoemulsion for oral application: Development, characterization and stability. Acta Pharm. 2018; 69(1):33–48.
- **42.**Swain S, Patra CN, Rao ME. Pharmaceutical drug delivery systems and vehicles. New Delhi: Woodhead Publishing India Pvt. Ltd; 2016. Chapter 1. Self-emulsifying drug delivery systems. 27-28.
- **43.** Al-sakini SJ, Maraie NK. Optimization and in vitro evaluation of the release of class ii drug from its nanocubosomal dispersion. Int J Appl Pharm. 2019;11(2):86–90.
- **44.**Maalouf M, Sun CN, Pyle B, Emery M, Haugen GM, Hamrock SJ, et al. Factors enabling high mobility of protons and water in perfluorosulfonate membranes under low hydration conditions. Int J Hydrogen Energy. 2014; 39(6): 2795–800.
- **45.**British Pharmacopoeia Commission. British Pharmacopoeia. London: The Stationery Office; 2009. Volume I and II. Monographs: medicinal and pharmaceutical substances. 3375-3379.
- **46.** Elfiyani R, Amalia A, Pratama SY. Effect of using the combination of tween 80 and ethanol on the forming and physical

stability of microemulsion of eucalyptus oil as antibacterial. J Young Pharm. 2017; 9(1):118–21.

- **47.** Arora R, Aggarwal G, Harikumar SL, Kaur K. Nanoemulsion based hydrogel for enhanced transdermal delivery of ketoprofen. Adv Pharm. 2014; 2014:1–12.
- **48.**Sinko PJ, Sinch Y. Martin's physical pharmacy and pharmaceutical sciences. 6<sup>th</sup> ed. Philadelphia: Lippincott Williams & Wilkins, a Wolters Kluwer business; 2011. Chapter 20. Rheology. 469-492.
- **49.** Attwood D, Florence AT. Physical pharmacy. London: Pharmaceutical Press; 2008. Chapter 1. Solids. 1-10.
- **50.** Felton L. Remington: Essentials of pharmaceutics. London: Pharmaceutical Press; 2012. Chapter 8. Dissolution. 63-80.
- **51.**Bhagat C, Singh SK, Verma PRP, Singh N, Verma S, Ahsan MN. Crystalline and amorphous carvedilol-loaded nanoemulsions: Formulation optimisation using response surface methodology. J Exp Nanosci. 2013; 8(7–8):971–92.
- **52.** Dinda SC, Panda SK. Formulation and invitro/in-vivo assessment of enhanced bioavailability of lacidipine using nano pure technique. Albanian J Pharm Sci. 2014; 1(1):20–5.
- **53.**Sandeep V, Narendar D, Arjun N, Kishan V. Lacidipine loaded solid lipid nanoparticles for oral delivery: preparation, characterization and in vivo evaluation. Int. J. Pharm. Sci. Nanotechnol. 2016; 9(6):3524–30.
- **54.**Bozdağ-Pehlivan S, Subaşi B, Vural I, Ünlü N, Çapan Y. Evaluation of drugexcipient interaction in the formulation of celecoxib tablets. Acta Pol Pharm - Drug Res. 2011; 68(3):423–33.
- **55.**Singh N, Verma SM, Singh SK, Verma PRP. Antibacterial action of lipidic nanoemulsions using atomic force microscopy and scanning electron microscopy on Escherichia coli. J Exp Nanosci. 2015; 10(5):381–91.
- **56.** Hassmoro NF, Abdullah S, Rusop M. Atomic force microscopy characterization of latex nanoparticles synthesized by slow drying process of nano-emulsion polymerization. Procedia Eng . 2013; 56:755–9.



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