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Neuroprotective Effect of Vinpocetine against Lead Acetate-Instigated Neurotoxicity in Rats by Evaluation Tumor Necrosis Factor-Alpha, Interleukin-1Beta and Interleukin-10

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Abstract

Lead toxicity elicits neurological damage which is a well-known disorder that has been considered to be a major cause for multiple condition such as behavioral defect; mental retardation; and nerve insufficient activity.

This research is designed to estimate potential protective effect of vinpocetine on neurotoxicity stimulated by lead acetate in rats.

Eighteen adult rats of both sexes were randomly enrolled into three groups. Each group includes 6 rats as followings: Group I- Rats were given 0.3ml normal saline solution orally; then intraperitoneal injection of $100\mu l$ of the normal saline was given 1h later; this group was considered as control. Group II- Rats were given an intraperitoneal injection of 20mg/kg lead acetate for 5 days. Group III- Rats were orally given 3mg/kg vinpocetine, which was given 1hr before [(the IP injection of Pb every 24 hours at a dose of 20mg/kg) for 5 days and continued for 10 days]. On 11^{th} day of the study, the brain of each animal has been surgically cut-out to make homogenate preparation to estimate tumor necrosis factor-alpha (TNF- α), interleukin-1 beta (IL- 1β), and interleukin-10 (IL-10) levels.

Lead significantly elevated TNF- α and IL-1beta; while, it significantly decreased IL-10 levels. Vinpocetine significantly minimized IL-1beta and TNF- α ; furthermore, vinpocetine significantly raise IL-10 levels at (P<0.05).

Vinpocetine may have a neuro-protective activity against lead-stimulated toxicity brain of rats. **Keyword: Lead, Vinpocetine, Rats, Neuroprotective, Cytokines.**

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الكلمات المفتاحية: الرصاص فنبوستين الجرذان حماية عصبية سايتوكينات.

لخلاصة

الضرر العصبي الناجم عن سمية الرصاص هو حالة معروفة جيداً حيث أنها أساس للعديد من الاضطرابات مثل التخلف العقلي ؛ المشاكل السلوكية ؛ تلف الأعصاب تم تصميم هذا العمل للتحقق في النشاط الوقائي للفينبوسيتين على السمية العصبية التي تسببها أسيتات الرصاص في المجروعة : تلف الأعصاب تم المعتقدام ثمانية عشر جرذا بالغًا من كلا الجنسين بشكل عشوائي في ثلاث مجموعات مكونه ٦من جرذان لكل منها: المجموعة الأولى - أعطيت الجرذان ٢٠ مل من محلول ملحي عن طريق الفم. وبعد ساعه تم حقنها ب ١٠٠ ميكرولتر من محلول ملحي داخل الصفاق . تعد هذه المجموعة معلورة المجموعة الثانية: أعطيت الجرذان حقنة داخل الصفاق من أسيتات الرصاص لمدة ٥ أيام بجرعة والاستمرار لمدة ١٠ أيام) ؛ حيث تم إعطاؤه قبل ساعة واحدة من حقن الرصاص داخل الصفاق يومياً بجرعة ٢٠ مجم / كجم لمدة ٥ أيام. في اليوم والاستمرار لمدة ١٠ أيام) ؛ حيث تم إعطاؤه قبل ساعة واحدة من حقن الرصاص داخل الصفاق يومياً بجرعة ٢٠ مجم / كجم لمدة ٥ أيام. في اليوم والاستمرار لمدة ١٠ أيام) والانترلوكين واحد بيتا (1- (31 - 11)) النتائج: سببت اسيتات الرصاص بشكل معنوي بارتفاع الانترلوكين واحد بيتا وعامل نخر الورم الفا وكان له دور معنوي في رفع الانترلوكين عشرة في جناسة نسيج الدماغ . الفنبوستين قلل بشكل معنوي من الانترلوكين واحد بيتا وعامل نخر الورم الفا وكان له دور معنوي في رفع الانترلوكين عشرة في جناسة نسيج الدماغ عند بشكل معنوي من الانتراوكين عشرة في جناسة نسيج الدماغ عند بشكل معنوي المستقبة بواسطة اسيتات الرصاص في الجرذان.

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Introduction

Exposition to Lead (Pb) can occur through many routes involving contaminated water, air, food, soil, and other public pathway; and the secure threshold for Pb exposure has not been specified, as there is no accurate amount for toxicity of such element (1). Pb is available in various forms and is considered as a basic ingredient of different organic compounds, which have been directly penetrated to skin, respiratory, and brain; where, toxic effect of central nervous system is considered as a dominant effect of such element (2). Thus; Pb can cause significant public health problems, although its concentration in ecosystem has been decreased after several trials (3). Researchers published that neurological destruction that stimulated by Pb toxicity can urge various disorders like mental defect, Alzheimer's disease (AD), behavioral problems, loss of nerve activity, Parkinson's disease (PD), and probably schizophrenia (4). Pb has the ability to pass through blood-brain barrier (BBB) and can substitute calcium (Ca⁺²) ions. Accordingly, such element can interfere with activity of Ca+2 on cell functions and perturb several biological actions (5). The pro-inflammatory pathway of neurotoxicityinstigated by Pb has not been completely evaluated

Vinpocetine is an alkaloid vincamine derivative. In many countries, vinpocetine has been utilized for the treatment of central nervous system disorders such as stroke and dementia for more than 30 years. Up to date, It is also obtainable in the market as a nutrition supplement to boost memory and cognizance. The safe and marvelous activity of vinpocetine result in discovering the novel remediation and mechanism actions of it in disease pattern and diverse cell types ⁽⁸⁾.

Vinpocetine, has boosting the cognitive function that it has been utilized as a nootropic agent for patients with central nervous system disorder; where, it increases glucose uptake and cerebral blood flow (9). Moreover, it can reduce the peril of strokes and temporary ischemic attacks in chronic cerebrovascular insufficiency patients (10). As well as, vinpocetine is an efficacious antioxidant and then inhibit lipid peroxidation (11). Furthermore, such drug exhibit memory-protective and memoryboosting properties and potent anti-inflammatory (12) activity Besides, vinpocetine phosphodiesterase-1 [(PDE)-1] inhibitor (13) and a blocker of voltage-gated Na⁺ channels ⁽¹⁴⁾. Previously, in vitro studies approved vinpocetine inhibited the blockage of the mitochondrial complexes (II, III, and IV) as well as entirely negated the deduction of pyruvate levels and the assemble of free radical-stimulated by noxious concentrations of amyloid peptides in PC12 cells (15). It is a potential choice for the management of various neurodegenerative diseases that related to

cognitive improvement properties and the antiinflammatory effect of vinpocetine (16).

The current study is designed to estimate probable protective action of vinpocetine against neurotoxicity instigate by Pb in rats through the estimation of tumor necrosis factor-alpha (TNF-alpha), interleukin-10 (IL-10) and, interleukin-1 beta (IL-1 β) levels in brain tissue homogenate.

Materials and Methods

Experimental animals

Eighteen male and female Albino adult rats (weighing 160-250gm) were selected for this study, Rats were obtained from House Animal for College of Pharmacy, Basra University. Commercial pellets and tap water *ad libitum* were dependent in feeding of rats during experiment period.

Materials

Lead acetate powder was purchased from Fluka Chemical, Turkey. Vinpocetine pure powder was purchased from America medic science (USA).

Experimental design

Adult rats were randomly distributed into three equal groups (6 animals for each group) as follows: **Group I**- each rat was given 0.3ml normal saline orally for 10 days,5day before IP injection of normal saline and then 100µl of the normal saline solution injected IP 1hr later for 5day; this group considered as control. **Group II**- each rat was given 0.3ml normal saline orally for 10 days,5day before IP injection of pb acetate which freshly prepared (20 mg/kg/day body wt.) for 5 days ⁽¹⁷⁾. **Group III**- Each rat was orally given 3mg/kg/day vinpocetine (dissolve in normal saline for 5 days by oral gavage before starting Pb injection and continued for 10 days; where it was given 1hr before Pb, which was injected IP every day at a dose 20 mg/kg for 5 days ⁽¹⁸⁾

Twenty four hour after the end of the treatment duration; i.e.at day six, each animal was euthanized by diethyl ether, and then by cervical dislocation. Thereafter, the skull was crushed by surgical scissor and then the brain of each rat has been cutout surgically for homogenate preparation.

Preparation and estimation of homogenate biochemical parameters

The preparation of brain tissue homogenate involved removal of excess blood by rinsing in ice-cold phosphate buffer saline (PBS,pH=7.4), followed by desiccation using filter paper and then measuring the weight of each brain tissue before homogenization was performed. Then each of rats' brain tissue minced to small pieces and put in 15ml plastic test tube containing chilled PBS solution (pH=7.4); where ratio of tissue weight in g to PBS volume in mL is 1:9. Homogenization was performed using cell lab homogenizer in icy condition. Then, the homogenate was centrifuged for approximately 15 minutes at 2000×g. The supernatant liquid was accurately collected and kept

at -20 °C until the time for the evaluation of TNF- α , IL1 β , and IL-10 cytokines levels by automated biochemistry analyzer (Elabscience, USA) ⁽¹⁹⁾.

Statistical analyses

Data were explicated as mean \pm standard error (SEM). ANOVA –post hoc test was utilized for estimating the significant difference among groups. Differences were statistically considerable for P value less than 0.05 (P<0.05).

Results

Effect of vinpocetine against lead acetate on tumor necrosis factor-alpha (TNF- α) in rats' brain tissue homogenate.

Table 1 and figure 1 showed that rats injected with 20 mg/kg of Pb acetate IP every day for 5 days lead to a significant elevation in the level of TNF- α in homogenate tissue of brain compared to those level in control rats. The TNF- α level in brain tissue homogenate were respectively, 307.5 ± 17.1 and 83.3 ± 5.3 .

Furthermore, there were significant reduction in TNF- α level in homogenate tissue of brain in group of rats treated with 3mg/kg vinpocetine prior to 20 mg/kg of Pb acetate compared to the corresponding level in group of rats injected IP with 20 mg/kg of Pb acetate every day for 5 days . The TNF- α level in brain tissue homogenate were respectively, 173.3 ± 4.4 and 307.5 ± 17.1 .

Effect of vinpocetine against lead acetate on interleukin-1beta (IL-1 β) in rats' brain tissue homogenate.

Table 1 and figure 2 showed that rats injected with Pb acetate every 24 hours at a dose

20mg/kg for 5 days IP led to significant rising in the level of IL-1 β in homogenate tissue of brain compared to control rats. The level of such cytokine in homogenate tissue of brain were respectively, 125.5 ± 6.6 and 61.3 ± 4.9 .

Moreover, table 1, and figure 2 showed that, there were significant reduction in IL-1 β level in homogenate tissue of brain for the groups of rats treated with 3mg/kg vinpocetine prior to 20 mg/kg Pb acetate compared to corresponding levels in group of rats injected IP with 20 mg/kg of Pb acetate every day for 5 days. The level of IL-1 β in homogenate tissue of brain were respectively, 84 ± 3.9 and 125.5 ± 6.6 .

Effect of vinpocetine against lead acetate on interleukin-10 (IL-10) in rats' brain tissue homogenate.

Table 1 and figure 3 showed that rats injected with 20 mg/kg of Pb acetate every day for 5 days IP, there was a significant reduction in the level of IL-10 in brain tissue homogenate compared to those levels in control rats. The level of IL-10 in homogenate tissue of brain were respectively, 40 ± 2.8 and 205 ± 7.6 .

Furthermore, there were significant raising in the level of IL-10 in homogenate tissue of brain in rats treated with 3mg/kg vinpocetine prior to 20 mg/kg lead acetate compared to corresponding levels in group of rats injected with 20 mg/kg of Pb acetate every day for 5 days IP. The level of IL-10 in homogenate tissue of brain were respectively, 63.8 ± 3.9 and 40 ± 2.8 .

Table 1. Effect of vinpocetine on TNF- α , IL-1 β , and IL-10 levels in brain tissue homogenate of rats after IP injection of lead acetate

Treatment Groups n=6	Treatment Type	TNF-alpha (pg/ml) (Mean±SEM)	IL-1Beta (pg/ml) (Mean±SEM)	IL-10 (pg/ml) (Mean±SEM)
I	Negative control/ normal saline	83.3±5.3	61.3±4.9	205±7.6
II	lead acetate (20 mg/kg)	307.5±17.1*a	125.5±6.6*a	40±2.8*a
Ш	3mg/kg vinpocetine prior to 20 mg/kg of lead acetate	173.3±4.4 ^b	84±3.9 b	63.8±3.9 b

^{*} Mean significant-different compared to control rats at P<0.05.

Different letters mean there are significant different in the same column at P<0.05.

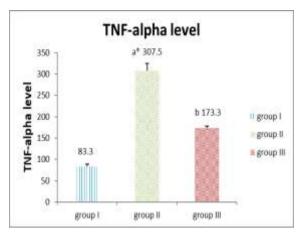


Figure 1. Effect of vinpocetine on TNF- α levels in brain tissue homogenate after IP injection of lead acetate in rats.

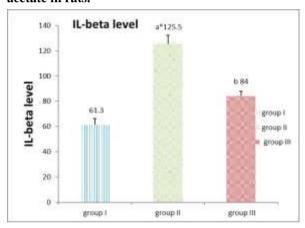


Figure 2. Effect of vinpocetine on IL-1 β levels in brain tissue homogenate after IP injection of lead acetate in rats.

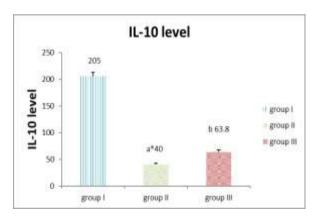


Figure 3. Effect of vinpocetine on IL-10 levels in brain tissue homogenate after IP injection of lead acetate in rats.

Discussion

The present study pointed out on markers of inflammation, [(TNF- α , IL-1b), and the anti-inflammation marker (IL-10)] levels after exposure to Pb; and this study furthermore inspected neuroprotective action of vinpocetine on above markers.

In developing animals, Pb can cross bloodbrain barrier (BBB) and deactivate the basic structural components by damaging the brain glial Moreover, such element stimulated devastation that mainly-occur in most area of brain that include cerebellum, cerebral cortex, and hippocampus, that may consequently cause morphological change in the brain (20). The destructive effect of lead may be related to its creation of ROS or its confliction with calcium inactivation of protein kinase C (PKCs), which may have a critical role in signal transduction, differentiation and cell development Furthermore, lead vies with calcium for prevalent binding sites and is integrated into neurotransmission systems of calcium (22).

The major findings of previous studies that approved raising cytokine creation and axonal destruction with astrocytic activation by Pb effect in immature rat brain ⁽²³⁾. Furthermore, researchers mentioned that Pb can cause increment in level of inflammatory cytokines ⁽⁷⁾.

Glial cells have basic role in local inflammatory processes by creation cytokines such as TNF- α , IL-1 β , IL-6. Furthermore, the neuroinflammation was controlled by activation of glial cells which participated in destructive and progression of several disorders. In Alzheimer's disease, inflammatory and oxidative induction effect that were created by chronically glial cells activation which result destruction of neurons $^{(24)}$.

In this study, IP injection of Pb acetate every 24 hours (20mg/kg) for five days to rats in group II, significantly increase level of inflammatory markers such as TNF- α and IL- β levels of these cytokines, and a significant reduction in IL-10 level each when compared with control group rats at (P<0.05). Results of this work are agreed with previously-mentioned studies (7 , 23).

Furthermore, this study showed the preventive effect of vinpocetine on inflammatory pathway of brain through suppressing the TNF-α and IL-1\beta elevation; where, oral-administration of vinpocetine for 5 days prior to IP injection of lead acetate for 5 days and vinpocetine continued for 10 days in group III significantly decrease levels of inflammatory markers such as cytokines TNF-α and IL-β levels, but with significant increase in IL-10 level when compared with those levels in rats IP injected with Pb acetate at (P<0.05). Researchers reported that, in the CNS; where, the PDE inhibitor (vinpocetine) can down-regulate the following inflammatory cytokines [TNF-α, IL-1, and IL-6], however, it can up-regulate the suppressor cytokines such as IL-10 (25) due to the effects of lipopolysaccharides.

Conclusion

Vinpocetine may have a neuro-protective action against lead-instigate neurotoxicity in rats.

Acknowledgements

This is the first study to estimate in *vivo* neuro-protective action of vinpocetine on leadinstigate neurotoxicity in rats.

Competing interests

There are no competing interests to declare.

References

- 1. Ahmed M, Meki A, and AbdRaboh N .

 Neurotoxic effect of lead on rats:

 Relationship to Apoptosis.

 International Journal of Health Sciences
 2013;(7): 192-199
- 2. Ahmed E. Abdel Moneim & Mohamed A. Dkhil & Saleh Al-Quraishy. Effects of Flaxseed Oil on Lead Acetate-Induced Neurotoxicity in Rats. Biol Trace Elem Res 2011:144:904–913.
- 3. Rojas-Castañeda JC, Vigueras-Villaseñor RM, Rojas P, Chávez-Saldaña M, Gutiérrez-Pérez O, Montes S, Ríos C: Alterations induced by chronic lead exposure on the cells of the circadian pacemaker of developing rats. Int J Exp Pathol 2011;4: 243-50.
- **4.** Liu J, Han D, Li Y, Zheng L, Gu C, Piao Z, Au W, Xu Z, Huo X . Lead Affects Apoptosis and Related Gene XIAP and Smac Expression in the Hippocampus of Developing Rats. Neurochem Res 2010; 35: 473-479.
- 5. Sanders T, Liu Y, Buchner V, and Tchounwou P: Neurotoxic Effects and Biomarkers of Lead Exposure: A Review. Rev Environ Health 2009; 24: 15–45.
- 6. Chibowska K, Bosiacka I, Falkowska A, Gutowska I, Goschorska M, Chlubek D . Review Effect of Lead (Pb) on Inflammatory Processes in the Brain. Int. J. Mol. Sci 2016;17: 2140.
- 7. Farkhondeh T, Boskabady M, Koohi, M, Sadeghi-Hashjin G, Moin M. The effect of lead exposure on selected blood inflammatory biomarkers in guinea pigs. Cardiovasc. Hematol. Disord. Drug Targets 2013; 13: 45–49.
- **8.** Zhanga Li J. An update on vinpocetine: New discoveries and clinical implications . <u>European Journal of Pharmacology</u> . 2018; <u>819</u>: 30-34
- 9. Vas A, Gulyas B, Szabo Z, Bonoczk P, Csiba L, Kiss B, et al. Clinical and non-clinical investigations using positron emission tomography, near-infrared spectroscopy and transcranial Doppler methods on the neuroprotective drug vinpocetine: a summary of evidence. J Neurol Sci 2002; 203-204:259–262.
- **10.** Valikovics A. Investigation of the effect of vinpocetine on cerebral blood flow and cognitive functions. Ideology Sz 2007;60:301–310.

- **11.** Zaitone SA, Abo-Elmatty DM, Elshazly SM. Piracetam and vinpocetine ameliorate rotenone-induced Parkinsonism in rats. Indian J Pharmacol 2012; 44(6):774–779.
- 12. JeonKI,XiangbinXuX,AizawaT,LimJH,Jo noH,KwonDS,AbeJ,Berk BC,Jian-DongLiJD, ChenYanC . Vinpocetine inhibits NF-κB– dependent inflammation via an IKK-dependent but PDE-independent mechanism. Proc Natl Acad Sci U S A 2010;107(21):9795–9800.
- 13. Van Staveren WCG, Markerink-van Ittersum M, Steinbusch HW, de Vente J. The effects of phosphodiesterase inhibition on cyclic GMP and cyclic AMP accumulation in the hippocampus of the rat. Brain Res 2001; 888:275–286.
- 14. Sitges M, Galvan E, Nekrassov V. Vinpocetine blockade of sodium channels inhibits the rise in sodium and calciuminduced by 4 aminopyridines in synaptosomes. Neurochem Int 2005;46:533–540.
- **15.** Pereira C, Agostinho P, Oliveira CR. Vinpocetine attenuates the metabolic dysfunction induced by amyloid betapeptides in PC12 cells. Free Radic Res 2000; 33(5):497–506.
- **16.** Patyar S, Prakash A, Modi M, Medhi B. Role of vinpocetine in cerebrovascular diseases. Pharmacol Rep 2011;63:618–628.
- 17. Nowak P, Szczerbak G, Nitka D, Kostrzewa RM, Sitkiewicz T, Brus R . Effect of prenatal lead exposure on nigrostriatal neurotransmission and hydroxyl radical formation in rat neostriatum: dopaminergic—nitrergic interaction. Toxicology 2008; 246:83–89
- **18.** Rania I. Nadeem1 & Hebatalla I. Ahmed2 & Bahia M. El-Sayeh. Protective effect of vinpocetine against neurotoxicity of manganese in adult male rats. Springer Nature 2018; 391(7):729-742
- **19.** Manal A. I. Al-Geam and Nada N. Al-Shawi. Effects of vitamin E and Q10 supplementation against doxorubicin-induced neurotoxicity in Rats. Iraqi J Pharm Sci, 2018; 27(2): 24-31 24.
- **20.** Jaya Prasanthi RP, Hariprasad Reddy G, Bhuvaneswari Devi C, Rajarami Reddy G. Zinc and calcium reduce lead-induced perturbations in the aminergic system of developing the brain. Biometals 2005:18:615–626.
- **21.** Costa LG. Signal transduction in environmental neurotoxicity. Annu Rev Pharmacol Toxicol 1998; 38:21–43.
- 22. Devi CB, Reddy GH, Prasanthi RP, Chetty CS, Reddy GR. Developmental lead exposure alters mitochondrial

- monoamine oxidase and synaptosomal catecholamine levels in rat brain. Int J Dev Neurosci 2005;23:375–381
- 23. Carey J, Allshire, A., and Van Pelt, F. Immune modulation by cadmium and lead in the acute reporter antigen-popliteal lymph node assay. Toxicol 2006; Sci. 91, 113–122.
- **24.** Struzynska L Bouta B, Koza k, Sulkowski G. Inflammation-Like Glial
- Response in Lead-Exposed Immature Rat Brain. Toxicological Sciences 2007;95(1), 156–162.
- **25.** Yoshikawa M, Suzumura A, Tamaru T, Takayanagi T, Sawada M. Effects of phosphodiesterase inhibitors on cytokine production by microglia. Mult Scler 1999; 5(2):126–133.



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