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Original scientific paper

# Reversed phase parallel artificial membrane permeation assay for log *P* measurement

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#### **Abstract**

A reversed phase parallel artificial membrane permeation assay (RP-PAMPA) was newly invented for log P measurement. An oil/water/oil sandwich was constructed using a conventional PAMPA instrument. 1 % agarose was used to improve the physical stability of the water phase. A linear correlation between log P and the apparent permeability was observed in the -0.24 < log P < 2.85 region ( $R^2 = 0.98$ ). RP-PAMPA was also applied to pKa measurement.

## **Keywords**

octanol, partition, parallel artificial membrane permeation assay,  $pK_a$ , drug

#### Introduction

High throughput physicochemical profiling of a drug is still challenging in early drug discovery. Various methods have been proposed for octanol – water partition coefficient (log P), solubility, and  $pK_a$  measurements [1]. The parallel artificial membrane permeation assay (PAMPA) has been widely used in drug discovery as PAMPA is compatible with high throughput screening (HTS) [2,3]. In normal phase (NP-) PAMPA methods, a lipid phase is immobilized on a filter (usually a 96 well filter plate) and the permeability of a drug across the lipid membrane is measured. Previously, Faller et al. applied NP-PAMPA to  $\log P$  measurement [4]. The apparent permeability ( $P_{\rm app}$ ) across the octanol impregnated filter membrane was found to correlate with logP. However, the  $P_{\rm app}$  —  $\log P$  curve showed a bell-shaped relationship with a plateau around  $\log P = 1$ . Therefore, it was impossible to estimate the  $\log P$  values around  $\log P = 1$ . A bell-shaped relationship between  $P_{\rm app}$  and lipophilicity of drugs is usually observed in NP-PAMPA [5].

The purpose of the present study was to overcome the drawback of NP-PAMPA for  $\log P$  measurement. A reversed phase PAMPA (RP-PAMPA) method for  $\log P$  measurement was newly invented. In RP-PAMPA, an oil/water/oil sandwich was constructed using a conventional PAMPA instrument (Figure 1). In addition, RP-PAMPA was applied for  $pK_a$  measurement, especially for low solubility compounds.

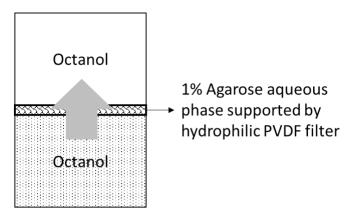


Figure 1. Schematic configuration of RP-PAMPA

#### **Materials and Methods**

#### Materials

Octanol, pentoxifylline, 1-naphthol, dipyridamole, and acid blue 9 were purchased from Tokyo chemical industry (Tokyo, Japan). Agar powder, agarose S, agarose H, prednisone, sulfamethoxazole, carbamazepine, caffeine, chlormphenicol, ethanol, trisodium citrate, sodium dihydrogenphosphate, disodium hydrogenphosphate, sodium hydroxide solution, propranolol hydrochloride, warfarin sodium, piroxicam, and ketoprofen were purchased from Wako pure chemicals (Tokyo, Japan). Phenacetin was purchased from Yamamoto Corporation (Osaka, Japan). The other reagents were of analytical grade.

## $P_{app}$ measurement

The schematic configuration of RP-PAMPA is shown in Figure 1. In RP-PAMPA, the water phase (water membrane) was immobilized on the hydrophilic filter with the aid of agarose. Agarose S was dissolved in hot water or a buffer at 1.0 % and then poured into a hydrophilic filter (Multi Screen-HV, pore size 0.48  $\mu$ m, low protein binding, Millipore). A model drug was dissolved in octanol at 10 mM and added to the donor plate (downside, 300  $\mu$ L). The filter plate was then put on the donor plate. The filter plate was filled with 200  $\mu$ L of octanol. After 16 hour incubation at room temperature (25 ± 1 °C), both octanol phases were diluted tenfold by ethanol and the drug concentrations in the donor and accepter sides were measured by UV spectroscopy. For p $K_a$  measurement sodium - phosphate and sodium - citric acid buffers (100 mM of anion species) were used to construct the water membrane.  $P_{app}$  was calculated as previously reported [6]:

$$P_{\text{app}} = \frac{-\ln(1 - C_{\text{A}} / C_{\text{equillibrium}})}{A \times (1 / V_{\text{D}} + 1 / V_{\text{A}}) \times t} \tag{1}$$

$$C_{\text{equilibrium}} = (C_{\text{D}} \times V_{\text{D}} + C_{\text{A}} \times V_{\text{A}}) / (V_{\text{D}} + V_{\text{A}})$$
(2)

where  $C_A$  and  $C_D$  are the drug concentrations in the donor and acceptor phases at time t, respectively.  $V_D$  and  $V_A$  are the volumes of the donor and acceptor phases, respectively. A is the membrane surface area (0.28 cm<sup>2</sup>).

### **Results**

## Construction of RP-PAMPA

We first investigated the stability of the water phase (water membrane) constructed on the hydrophilic

filter with the aid of agarose. 1.0 % concentration was selected to enable pipetting of the hot sol phase while maintaining the physical strength of the agarose gel. It was found that at least 30  $\mu$ L of 1 % agarose was required to provide sufficient physical strength for  $P_{app}$  measurement. The ager powder was found to form a less stable water phase compared to Agarose S and H. In a preliminary study, it was found that more than 10 hours were required to achieve a steady – state flux across the water phase (data not shown). Therefore, the Agarose S 30  $\mu$ L water membrane and 16 hour incubation time were employed in the following studies.

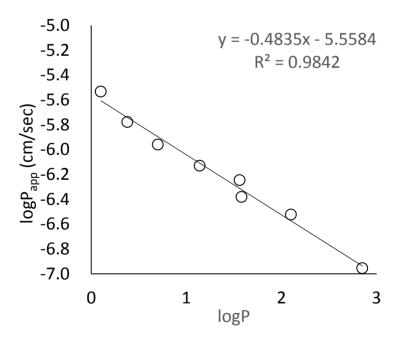
# $Log P_{app} - log P relationship$

The log  $P_{\rm app}$  and log P data are summarized in Table 1 [1,7,8]. Figure 2 shows the correlation between log  $P_{\rm app}$  and log P. A linear correlation was observed in the -0.24 < log P < 2.85 region (R<sup>2</sup> = 0.98). The slope of the log-log plot was -0.48.

**Table 1**.  $P_{app}$  and  $\log P$ 

Table 217 app arta 1087		
Drug	log P (literature) <sup>a</sup>	$\log P_{app}$ (cm s <sup>-1</sup> , mean ± S.D., N = 6)
phenacetin	1.58	-6.38 ± 0.04
caffeine	0.10	-5.53 ± 0.02
carbamazepine	2.1	-6.52 ± 0.04
prednisone	1.56	-6.25 ± 0.01
chloramphenicol	1.14	-6.13 ± 0.01
sulfamethoxazole	0.70	-5.96 ± 0.02
1-naphthol	2.85	-6.95 ± 0.03
pentoxifylline	0.38	-5.78 ± 0.02

<sup>&</sup>lt;sup>a</sup> Refs. [1,7,8]

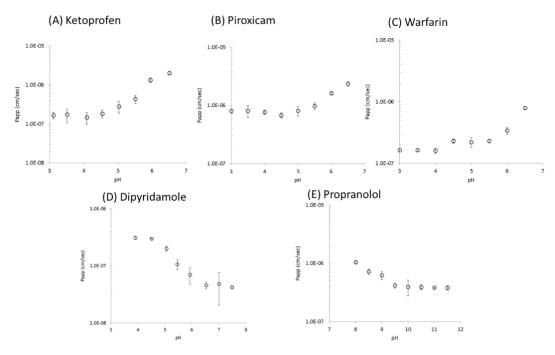


**Figure 2**. Log  $P - \log P_{app}$  relationship.

## $pH - P_{app}$ profile

The pH -  $P_{app}$  profiles were shown in Figure 3 and Table 2. The p $K_a$  values were obtained as the intersection of the slope and the horizontal lines. Estimated and literature p $K_a$  values are shown in Table 3 [1,9].





**Figure 3**. pH –  $P_{app}$  relationship. Mean  $\pm$  S.D. N = 3.

**Table 2.**  $P_{app}$  of dissociable drugs at each pH<sup>a</sup>

Compound	рН	$P_{\rm app}$ (10 <sup>-6</sup> cm sec <sup>-1</sup> )	Compound	рН	$P_{\rm app}$ (10 <sup>-6</sup> cm sec <sup>-1</sup> )
Ketoprofen	3.1	$0.17 \pm 0.03$	Dipyridamole	3.9	$0.31 \pm 0.02$
$(\log P = 3.2)$	3.5	0.17 ± 0.07	$(\log P = 3.9)$	4.5	$0.30 \pm 0.01$
	4.1	0.15 ± 0.05		5.1	$0.20 \pm 0.02$
	4.5	$0.18 \pm 0.04$		5.5	$0.11 \pm 0.02$
	5.0	$0.28 \pm 0.09$		5.9	$0.07 \pm 0.02$
	5.5	$0.44 \pm 0.10$		6.5	0.05 ± 0.01
	6.0	$1.33 \pm 0.18$		7.0	$0.05 \pm 0.03$
	6.5	$2.00 \pm 0.23$		7.5	$0.04 \pm 0.00$
Piroxicam	3.0	$0.81 \pm 0.07$	Propranolol	8.0	1.05 ± 0.06
$(\log P = 2.0)$	3.5	0.81 ± 0.19	$(\log P = 2.9)$	8.5	$0.72 \pm 0.08$
	4.0	0.77 ± 0.07		9.0	$0.63 \pm 0.10$
	4.5	$0.68 \pm 0.07$		9.5	$0.42 \pm 0.04$
	5.0	0.81 ± 0.15		10.0	$0.40 \pm 0.12$
	5.5	$0.98 \pm 0.13$		10.5	$0.39 \pm 0.04$
	6.0	1.62 ± 0.08		11.0	$0.38 \pm 0.01$
	6.5	2.37 ± 0.24		11.5	$0.38 \pm 0.03$
Warfarin	3.0	0.16 ± 0.01			
$(\log P = 3.1)$	3.5	0.16 ± 0.01			
	4.0	0.16 ± 0.02			
	4.5	$0.23 \pm 0.02$			
	5.0	$0.22 \pm 0.04$			
	5.5	$0.23 \pm 0.02$			
	6.0	$0.34 \pm 0.04$			
	6.5	$0.80 \pm 0.05$			

 $<sup>^{\</sup>rm a}$  Mean  $\pm$  S.D. N = 3. Measured at 25 °C. The buffer concentration was 100 mM.

**Table 3**.  $pK_a$  values

Drug	This study <sup>a</sup>	Literature <sup>b</sup>
Ketoprofen	4.9	4.0
Piroxicam	4.8	4.7
Warfarin	5.7	5.0
Dipyridamole	5.9	6.1
Propranolol	9.4	9.5

<sup>&</sup>lt;sup>a</sup> Measured at 25 °C. The buffer concentration was 100 mM.

#### **Discussion**

In this study, RP-PAMPA for log P measurement was investigated for the first time. 1.0 % agarose was used to improve the physical stability of the water membrane. The mesh size of agarose is significantly larger than the size of drug molecules so that it does not affect the diffusion coefficient of drugs [10]. By using RP-PAMPA,  $\log P$  in the -0.24 <  $\log P$  < 2.85 range can be accurately measured. The measurable range can be expanded by using a more sensitive quantitation method such as LC-MS. The slope of the  $P_{\rm app} - \log P$  relationship was 0.45, which is significantly smaller than 1. If  $P_{\rm app}$  follows the solubility – partition theory for membrane permeation, i.e.,  $P_{\rm app} = PD/h$  where D is the diffusion coefficient and h is the thickness of the membrane, the slope of the log-log plot should be unity [11]. The reason for this deviation is not clear.

Previously, Kwon et al. reported a poly(dimethylsiloxane)(PDMS) permeation assay, which might be regarded as a kind of reversed phase membrane permeation assay [12]. However, the configuration of the PDMS permeation assay was largely different from the one used in the present study that is usually referred as PAMPA. In the PDMS permeation assay, a side-by-side single diffusion chamber was employed. A PDMS membrane was put between two chambers filled with aqueous bulk fluids. In addition, PDMS disks were added to both the donor and acceptor sides as dosing and sampling (extracting) phases, respectively. The aqueous phases were stirred by magnetic stirrers. In the PDMS permeation assay, a good correlation was observed between  $\log P_{\rm app}$  and  $\log P$  in the range of  $\log P > 3$  even though PDMS was used instead of octanol as the oil phase.

For low solubility drugs, it has been difficult to measure  $pK_a$  by using conventional methods such as pH titration. The pH – solubility profile can be used to estimate  $pK_a$  for low solubility drugs [13,14]. However, this method may not be accurate due to aggregate formation, low detection limit, etc. In RP-PAMPA, a drug is solubilized in the organic solvent phase. Therefore, it would become possible to measure  $pK_a$  for low solubility drugs by using RP-PAMPA. As the pH of the water membrane was changed in RP-PAMPA, the pH –  $P_{app}$  relationship should become a mirror image of that for NP-PAMPA. However, the pH –  $P_{app}$  relationship deviated from the Henderson – Hasselbalch equation. Therefore, the  $pK_a$  values of the model drugs were estimated as the intersection of the slope and the horizontal lines. The  $pK_a$  values of acidic drugs were underestimated by the RP-PAMPA method. The reason for this deviation is not clear. One possible reason may be that the incubation time of 16 hours might not be sufficient to achieve a steady state at pH >  $pK_a$  for acids. The  $pK_a$  of diclofenac has been reported to be ca. 4.0 in most cases in the literature. However,  $pK_a$  of 5.7 was obtained from the pH-solubility profile [15]. The  $P_{app}$  value at a pH where a drug molecule is undissociable (intrinsic water permeability,  $P_{w,int}$ ) also correlated with log P. However,  $P_{w,int}$  deviated from the log  $P_{app}$  — log P line for the undissociable drugs about 0.3 log unit. The difference of the water membrane (pure water vs. a buffer) could be a reason for the discrepancy. The present RP-PAMPA method

<sup>&</sup>lt;sup>b</sup> Refs. [1], [9].

needs to be improved for  $pK_a$  measurement in the future.

In conclusion, in the present study, RP-PAMPA for  $\log P$  measurement was constructed for the first time. 1.0 % agarose can be used to stabilize the water membrane. RP-PAMPA was applied to  $\log P$  and  $pK_a$  measurements. As PAMPA is compatible with the current HTS instrument, RP-PAMPA will be a useful tool in drug discovery.

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