

Serum Glycoproteins in Diabetic and Non-Diabetic Patients With and Without Cataract

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Purpose: To investigate the changes in serum glycoproteins from type 2 diabetic and non-diabetic patients with and without cataract.

Material and Method: A total of 85 subjects were selected for this study and divided into four groups. The first group consisted of 21 healthy subjects, the second group consisted of 21 diabetic patients with no chronic complications, the third group consisted of 20 diabetic patients with cataract and the fourth group had 23 non-diabetic patients with cataract. The patients with and without cataract were selected on clinical grounds from the Ziauddin University and Jinnah Postgraduate Medical Centre in Karachi, Pakistan.

Result: Diabetic patients with and without cataract had significantly higher levels of plasma glucose, glycated hemoglobin, glycated plasma proteins and serum fructosamine. In addition to these parameters, the levels of hexosamine, sialic acid and serum total protein were higher in diabetic compared to non diabetic subjects with and without cataract. Analysis of the protein fractions showed that alpha-1- and -2 globulins were higher in diabetic patients without complications compared to healthy subjects. Serum alpha-1-globulin, alpha-2-globulin, beta globulins and gamma globulins were all significantly higher in diabetic patients with cataract compared to healthy subjects but not serum albumin.

Conclusion: The levels of serum glycoproteins in non-diabetic patients with cataract were not higher than those of healthy subjects thus mechanisms other than hyperglycaemia are responsible for the development of cataract in these patients.

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Diabetes mellitus is a common endocrine disorder characterized by hyperglycemia, metabolic abnormalities and long-term complications afflicting the eyes, kidneys, nerves and blood vessels¹. World-wide projections suggest that more than 220 million people will have diabetes by the year 2010 and the majority of these will suffer from type 2 diabetes mellitus². In Pakistan, diabetes mellitus is a major health problem affecting more than 16% of people over the age of 25 years in some areas with a further 10% suffering from impaired glucose tolerance³.

Cataract is a serious consequence of long-term diabetes and according to the WHO, affects about half of the 45 million blind people world-wide⁴. Cataract is characterized by opacification of the eye lens affecting mainly the nuclear, cortical, and posterior subcapsular regions. Pathological studies of cataractous lenses have revealed that cataracts are composed of protein aggregates that precipitate in the lens of the eye. The insoluble aggregates obstruct the passage of light through the lens preventing it from reaching the photoreceptors in the retina⁵. The lens crystallins may be divided into α -, β - and γ -crystallins which are stable, water soluble proteins accounting for about 90% of the total protein content^{6,7}. These ubiquitous crystallins are expressed early in life and must remain transparent throughout life despite the high protein concentration in the lens and the continued exposure to intra and extracellular oxygen derived free radicals⁵.

Diabetic cataract occurs much earlier than senile cataract and causes opacification of the lens and eventual loss of vision. It has been suggested that increased non-enzymatic glycosylation (glycation) of lens crystallins may cause conformational changes resulting in exposure of thiol groups to oxidation and cross-link formation⁸. Furthermore, the lens crystallins have virtually no turnover and are ideal candidates for accumulation of glycation-derived cross-links. Thus increased cross-linking of lens crystallins may cause aggregation producing the high molecular weight material responsible for opacification. Increase glycation of serum proteins could cause an increase in the serum of circulating advanced glycation endproducts (AGEs). These AGEs are responsible for glycation-induced cross-linking of structural proteins

and believed to underlie the pathogenesis of diabetic complications⁹. Previous studies have also shown an increase in enzymatically glycosylated proteins such as alpha-2 glycoprotein fractions were increased in diabetic patients¹⁰.

The aim of this study was to investigate changes in serum glycoproteins (both enzymatic and non-enzymatic) in diabetic and non-diabetic patients with and without cataract.

MATERIALS AND METHODS

Patients over 50 years of age were selected on clinical grounds from Ziauddin University Hospital, Karachi and Jinnah Postgraduate Medical Centre, Karachi. The study included a total of 85 subjects which were divided into four groups. Group one consisted of 21 apparently healthy subjects who had no history of diabetes, cataract or any other major illness, like macro-vascular disease, retinopathy, tuberculosis, rheumatoid arthritis, liver disease or malignancy. Group two consisted of 21 type 2 diabetic patients without any clinical evidence of chronic diabetic complications whereas group three had 20 type 2 diabetic patients with cataract. Finally, group four consisted of 23 non-diabetic patients suffering from cataract. The age, sex, weight, duration of diabetes and treatment received were recorded. Drugs were stopped 48 hours prior to any sample collection. Physical examination including measurement of blood pressure and any family medical history was recorded. Individuals were classified as having diabetes mellitus if they have a fasting plasma glucose concentration ≥ 7.0 or random plasma glucose level ≥ 11.1 mmol/L according to established criteria¹¹. Patients with a history of blurred vision or double vision and spots were examined with a slit lamp to determine the type of cataract. Blood samples were collected from subjects after completing a consent form for each patient and explaining the nature of the study.

Blood glucose level determined by the glucose oxidase method¹². The reagents were obtained from glucose enzymatique PAP 7500 kit of bioMerieux. For the estimation of glycated haemoglobin, haemolysate was prepared by treating blood with a detergent in a buffered medium and removal of the labile fraction.

Haemoglobins are retained by a cationic exchange resin. Haemoglobin A_{1C} is specially eluted after washing away the haemoglobin A_{1A} and haemoglobin A_{1B} fractions and is quantified by direct photometric reading at a wavelength of 415 nm. The kit was obtained from Bio Systems Reagents and Instruments, Spain and based on an established procedure¹³. Fructosamine was detected by the nitro-blue tetrazolium reaction. The kit was obtained from Quimica Clinia Aplicada, Spain. Serum hexoamine was determined by Cessi and Pillego's method¹⁴, total serum protein by the Biuret method of Reinhold¹⁵, sialic acid by Natelson method¹⁶, and glycosylated proteins by the method of Ma *et al*¹⁷. Glycated plasma proteins were hydrolyzed with oxalic acid to release 5-hydroxymethyl furfural which was detected by reaction with thiobarbituric acid. This method gives overall estimation of ketoamine linkages. Serum protein electrophoresis¹⁸ was carried out by Helena Electrophoretic System, using a kit method (Titan III Cat. No. 3023 obtained from Helena Laboratories).

Epi-Info was used for statistical analysis of the data. Epi-Info is a statistical package available from the US Centre for Disease Control and Prevention. The statistical significance of the difference between two mean of various parameters between different groups was evaluated by Student's t test. The difference was

regarded as highly significant if the *P* value was less than 0.001, statistically significant if the *P* value was less than 0.05, and non-significant if the *P* value was greater than 0.05.

RESULTS

The mean age of non-diabetic and diabetic patients with cataract were significantly higher (*P*<0.05) as compared with control subjects (Table 1). Fasting plasma glucose, HbA_{1C}, serum fructosamine, glycosylated plasma protein, serum hexosamine and serum sialic acid levels were significantly higher (*P*<0.05) in all diabetic patients with or without cataract as compared with control subjects (Table 1). These parameters did not change (*P*>0.05) in non-diabetic patients with cataract as compared with control subjects (Table 1). However, they were higher in diabetic patients when compared to non-diabetic patients with cataract (*P*<0.001; Table 1). Total serum protein, alpha-1 and alpha-2 globulins were significantly higher (*P*<0.05) in diabetic patients with or without cataract as compared with control subjects (Table 1). Beta globulin and gamma globulin were significantly higher (*P*<0.05) in non-diabetic patients and diabetic patients with cataract as compared with control subjects (Table 1).

Table 1. The age, weight and concentration of blood analytes in non-diabetic and diabetic patients with and without cataract.

Parameters	Control Subjects (21)	Diabetic Patients without any complications (21)	Non- diabetic patients with cataract (20)	Diabetic patients with cataract (23)
Age (years)	53.81 ± 1.20	54.71 ± 1.40	59.83 ± 1.69 ^a	57.50 ± 1.58
Sex (F/M)	10/11	10/11	10/13	10/10
Weight (Kg)	64.30 ± 1.57	64.24 ± 1.62	65.22 ± 1.45	67.78 ± 1.55 ^a
Duration of diabetes (years)	-	9.29 ± 0.50	-	9.00 ± 1.00
Fasting plasma glucose (mmol/L)	5.04 ± 0.13	7.83 ± 0.32 ^a	5.34 ± 0.18	9.32 ± 0.34 ^{ab}
% Glycosylated haemoglobin (HbA _{1C})	4.98 ± 0.11	9.30 ± 0.37 ^a	5.04 ± 0.09	8.80 ± 0.34 ^{ab}
Serum fructosamine (mmol/L)	2.25 ± 0.08	3.72 ± 0.17 ^a	1.98 ± 0.07	3.05 ± 0.21 ^{ab}
Glycosylated plasma protein (absorbance / g)	6.20 ± 0.12	7.90 ± 0.30 ^a	5.85 ± 0.12	8.90 ± 0.34 ^{ab}
Hexosamine (mg/ dL)	67.86 ± 3.12	102.94 ± 3.63 ^a	77.52 ± 3.31	118.80 ± 3.43 ^{ab}
Sialic acid (mg/ dL)	35.36 ± 1.34	49.66 ± 1.78	39.22 ± 1.38	50.49 ± 1.76 ^{ab}

Total serum protein (gm%)	7.32 ± 0.12	7.94 ± 0.17 ^a	6.94 ± 0.20	7.97 ± 0.12 ^{ab}
Serum albumin (gm%)	4.01 ± 0.10	4.03 ± 0.11	3.01 ± 0.09	3.57 ± 0.12 ^b
Alpha-1 globulin (gm%)	0.16 ± 0.02	0.38 ± 0.06 ^a	0.17 ± 0.02	0.36 ± 0.09 ^{ab}
Alpha-2 globulin (gm%)	0.77 ± 0.03	0.96 ± 0.05 ^a	0.87 ± 0.05	1.54 ± 0.56 ^a
Beta globulin (gm%)	1.00 ± 0.03	0.92 ± 0.06	1.08 ± 0.07 ^a	1.14 ± 0.07 ^a
Gamma globulin (gm%)	1.48 ± 0.07	1.67 ± 0.09	1.89 ± 0.12 ^a	2.01 ± 0.11 ^a

a. Significant as compared with control subjects

b. Significant as compared with non diabetic patients with cataract

The correlation between fasting plasma glucose and HbA_{1C} in control subjects was $r = 0.284$, between fasting plasma glucose and HbA_{1C} in diabetic patients without cataract was $r = 0.478$, between fasting plasma glucose and HbA_{1C} in non-diabetic patients with cataract was $r = 0.267$ and between fasting plasma glucose and HbA_{1C} in diabetic patients with cataract was $r = 0.467$. The data of diabetic patients are currently available but main difference is that the values of non-diabetic patients with the same complication in the same age group were within normal limits and these parameters are not responsible for the complications.

DISCUSSION

Diabetes mellitus and its complications constitute an important health problem in both developing and developed countries. Cataract remains the commonest cause of blindness world-wide. In the present study, patients were selected with cataract and the possibility of other complications was excluded by the absence of any sign and symptoms on physical examination. Changes in protein concentration and increased enzymatic glycation of various proteins in diabetic patients have been correlated with hyperglycemia, which in turn causes early functional alterations in different tissues. In the present study non-diabetic patients with cataract have normal values except beta and gamma globulin which are increased in non-diabetic patients with cataract. Glycaemic related analytes increased in all diabetic patients with and without cataract and the levels did not change in non-diabetic patients with cataract and control subjects (Table 1). The presence of cataract in non-diabetic patients did not affect the glycaemic related analytes, and increase in diabetic patients without complications, reflect that the diabetes was

uncontrolled. They may be developing changes at the subclinical level which later on present as a complication.

The relationship among HbA_{1C}, blood glucose concentrations and late complications has been established over the last 30 years¹⁹⁻²¹. Serum fructosamine and glycated plasma protein concentrations have close correlation with HbA_{1C} because they reflect glycaemic control within the last 2 to 3 weeks and HbA_{1C} reflects glycaemic control for the last 4 to 6 weeks^{22,23}. In the present study, serum fructosamine and glycated plasma proteins in diabetic patients also have a close correlation with HbA_{1C}. The degree of glycation of plasma proteins, as an alternative index of control and as reflection of possible structural alterations of tissue proteins leading to complications was associated with the diabetic state²². Stratton *et al*²³ suggested that in patients with type 2 diabetes, the risk of diabetic complications was strongly associated with previous hyperglycemia. Any reduction in HbA_{1C} is likely to reduce the risk of complications, with the lowest risk being in those with HbA_{1C} in the normal range. In the present study, the coefficient of variation of HbA_{1C} was higher in diabetic patients with cataract as compared with non-diabetic patients with cataract. It seems that there could be different mechanisms for the development of cataract in diabetic and non-diabetic patients. Serum hexosamine and sialic acid levels were significantly increased in all diabetic patients and were non-significant in non-diabetic patients with cataract as compared with control subjects (Table 1). Other workers have made similar observations^{24,25}. Hangloo *et al*²⁶ found that age and sex had no influence on serum sialic acid levels. As sialic acid is incorporated into carbohydrate chains of glycoproteins and glycolipids in serum and tissues, the degree of incorporation of sialic acid has been reported to affect transvascular permeability and

accumulation of lipid in the arterial wall. Sialic acid is conjugated with constituents of acute phase reactants, which are highly concentrated on surface of endothelial cells²⁶. One likely hypothesis might be that the relationship between clinical condition and sialic acid concentration is due to the activity of a current inflammatory atherosclerotic process and/or to a direct damage to vascular endothelium causing sialic acid into the circulation²⁷. Serum hexosamine levels rise due to hexosamine biosynthetic pathway, which is involved in the pathogenesis of insulin resistance in patients with type 2 diabetes mellitus^{24,28,29}. In the present study the values of hexosamine and sialic acid in non-diabetic patients with cataract were in normal limits, but the values were increased in diabetic patients with cataract and without complications.

CONCLUSION

The uniform increase in fasting plasma glucose, glycosylated hemoglobin (HbA_{1c}), serum fructosemine, glycosylated plasma protein, serum hexosamine and serum sialic acid levels in diabetic patients indicates that the process of glycosylation depends upon hyperglycemia. The parameters do not rise in non-diabetic patients, and hence some other underlying mechanism may be responsible for the development of complications in these patients.

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REFERENCE

1. **Nathan DM.** Long-term complications of diabetes mellitus. *New Engl J Med.* 1993; 328: 1676-85.
2. **Alberti KGMM, Zimmet PZ.** Definition, diagnosis and classification of diabetes mellitus and its complication. Provisional report of a WHO consultations. *Diabetic Med.* 1998; 15: 539-53.
3. **Shera AS, Rafique G, Khawaja IA, et al.** Pakistan national diabetes survey: prevalence of glucose intolerance and associated factors in Shikarpur, Sindh province. *Diabetic Medicine.* 1995; 12: 1116-21.
4. The world health report 1998: life in the 21st century: a vision for all, World Health Organization, 1998.
5. **Bloemendal H, Jong W de, Jaenicke R, et al.** Ageing and vision: structure, stability and function of lens crystallins. *Progress in Biophysics and Molecular Biology* 2004; 86: 407-85.
6. **Horwitz J.** The function of α -crystallin in vision. *Seminars in cellular and developmental Biology.* 2000; 11: 53-60.
7. **MacRae T.** Structure and function of small heat shock/ α -crystallin proteins: Established concepts and emerging ideas. *Cellular and Molecular Life Sciences.* 2000; 57: 899-913.
8. **Kyselova Z, Stefak M, Bauer V.** Pharmacological prevention of diabetic cataract. *J Diabetes and its Complications.* 2004; 18: 129-40.
9. **Ahmed N.** Advanced glycation endproducts- role in pathology of diabetic complications. *Diabetes research and clinical practice.* 2005; 67: 3-21.
10. **Rahman MA, Zafar G, Shera AS.** Changes in glycosylated proteins in long-term complications of diabetes mellitus. *Biomed and Pharmacother* 1990; 44: 229-34.
11. **Gabir MM, Roumain J, Hanson RL, et al.** The 1997 American diabetes association and 1999 world health organization criteria for hyperglycemia in the diagnosis and prediction of diabetes. *Diabetes Care.* 2000; 23: 1108-12.
12. **Tietz NW:** Clinical guide to laboratory tests, 3rd ed., WB. Saunders. Company, Philadelphia, PA. 1995; 268-73.
13. **Rochman H.** Hemoglobin A_{1c} and diabetes mellitus. *Annals of clinical and laboratory science.* 1980; 10: 111-5.
14. **Cessi C, Pilliego F.** The determination of amino sugars in the presence of amino acids and glucose. *Biochemical J.* 1960; 77: 508-10.
15. **Varley H, Gowenlock AH, Bell M.** Practical clinical biochemistry, 5th ed, William Heinemann Medical Books Ltd, London. 1980; 545-7.
16. **Natelson S.** Microtechniques of clinical chemistry for the routine laboratory, 2nd ed, Thomas Springfield, Illinois. 1961; 378-80.
17. **Ma A, Naughton MA, Cameron DP.** Glycosylated plasma proteins: a simple method for the elimination of interference by glucose in its estimation. *Clinica Chimica Acta* 1981; 115: 111-7.
18. **Ralli EP, Barbosa X, Beck EM, et al.** Serum electrophoretic patterns in normal and diabetic subjects. *Metabolism.* 1957; 6: 331.
19. **McLellan AC, Thornalley PJ, Benn J, et al.** Glyoxalase system in clinical diabetes mellitus and correlation with diabetic complications. *Clinical Science.* 1994; 87: 21-9.
20. **Thornalley PJ.** The glyoxalase system: new development towards functional characterization of a metabolic pathway fundamental to biological life. *Biochemical J.* 1990; 269: 1-11.
21. **Brownlee M, Vlassara H, Cerami A.** Advanced glycosylation endproducts in tissue and the biochemical basis of complications. *New Engl J Med.* 1998; 318: 1315-21.
22. **Skeie S, Thue G, Sandberg S.** Interpretation of hemoglobin A_{1c} (HbA_{1c}) values among diabetic patients: Implication for quality specifications for HbA_{1c}. *Clinical Chemistry.* 2001; 47: 1212-7.
23. **Stratton IM, Adler AI, Neil HA, et al.** Association of glycaemia with macrovascular and microvascular complication of type 2 diabetes (UKPDS 35). *BMJ* 2000; 321: 405-12.
24. **Crook M.** The determination of plasma or serum sialic acid. *Clinical Biochemistry.* 1993; 26: 31-8.

25. **Marsall S, Bacote V, Traxinger RR:** Discovery of a metabolic pathway mediating glucose induced desensitization of the glucose transport system: Role of hexosamine biosynthesis in the induction of insulin resistance. *J Biological Chemistry*. 1991; 266: 4706-12.
26. **Hangloo VK, Kaul I, Zargar HU.** Serum sialic acid levels in healthy individuals. *J Postgraduate Medicine*. 1990; 36: 140-2.
27. **Lindberg G, Eklund GA, Gullberg B, et al.** Serum sialic acid concentration and cardiovascular mortality. *Br Medical J*. 1991; 302: 533-4.
28. **Span PN, Pouwels NJM, Olthaar AJ, et al.** Assay for hexosamine pathway intermediates (Uridine diphosphate-N-acetyl amino sugars) in small samples of human muscle tissue. *Clinical Chemistry*. 2001; 47: 944-6.
29. **Hawkin M:** Role of the glucosamine pathway in fat induced insulin resistance. *J Clinical Investigation*. 1997; 99: 2173-82.