

Review

## The Erythrocytic Hypothesis of Brain Energy Crisis in Sporadic Alzheimer Disease: Possible Consequences and Supporting Evidence

Elena Kosenko <sup>1,\*</sup><sup>(D)</sup>, Lyudmila Tikhonova <sup>1</sup><sup>(D)</sup>, Gubidat Alilova <sup>1</sup><sup>(D)</sup>, Amparo Urios <sup>2</sup> and Carmina Montoliu <sup>2,3</sup>

- <sup>1</sup> Institute of Theoretical and Experimental Biophysics of Russian Academy of Sciences, Pushchino 142290, Russia; ljudasik09@rambler.ru (L.T.); hells2012@yandex.ru (G.A.)
- <sup>2</sup> Hospital Clinico Research Foundation, INCLIVA Health Research Institute, 46010 Valencia, Spain; aurios@cipf.es (A.U.); carmina.montoliu@uv.es (C.M.)
- <sup>3</sup> Pathology Department, Faculty of Medicine, University of Valencia, 46010 Valencia, Spain
- \* Correspondence: eakos@rambler.ru or gieraki@mail.ru; Tel.: +7-4967-73-91-68

Received: 16 December 2019; Accepted: 10 January 2020; Published: 12 January 2020



Abstract: Alzheimer's disease (AD) is a fatal form of dementia of unknown etiology. Although amyloid plaque accumulation in the brain has been the subject of intensive research in disease pathogenesis and anti-amyloid drug development; the continued failures of the clinical trials suggest that amyloids are not a key cause of AD and new approaches to AD investigation and treatment are needed. We propose a new hypothesis of AD development based on metabolic abnormalities in circulating red blood cells (RBCs) that slow down oxygen release from RBCs into brain tissue which in turn leads to hypoxia-induced brain energy crisis; loss of neurons; and progressive atrophy preceding cognitive dysfunction. This review summarizes current evidence for the erythrocytic hypothesis of AD development and provides new insights into the causes of neurodegeneration offering an innovative way to diagnose and treat this systemic disease.

**Keywords:** Alzheimer's disease; amyloid  $\beta$  peptides; brain energy crisis; erythrocytic hypothesis; red blood cells; restoration of energy metabolism

## 1. Introduction

Alzheimer's disease (AD) is a progressive systemic disorder that develops slowly attacking the brain, brain blood vessels, and peripheral tissues [1], as well as red blood cells (RBC) [2,3], platelets [4], and leukocytes [5], and is characterized by progressive deterioration in cognitive abilities including memory impairment, typically in the elderly.

In 95%–98% of cases AD develops sporadically, and according to statistics on this type of dementia, about 50% of people aged 80 have the disorder [6]. Unless a way is found to halt progression of this disease, and if the population continues to grow, it is believed that by 2050 the number of people worldwide with this illness will be close to 115 million people, 7.7 million new patients per year, meaning that someone in the world will develop AD every 4 s [7].

Although during recent decades significant progress has been made in presenting different hypotheses of sporadic AD [8–19], the underlying mechanisms of the disease need further scrutinity.

The "amyloid cascade hypothesis" [20] has greatly influenced research over the last twenty years with considerable effort devoted to designing new anti-amyloid drugs, and although there is still no cure for AD [21,22], this hypothesis remains the most prevalent. Nevertheless, the key points of the hypothesis suggesting a central role for amyloid plaques in disease development and considering



the brain and its age-related changes independently of the blood flow are paradoxical rather than convincing. The main reason is that the idea behind the "amyloid cascade hypothesis" is not consistent with the known axiom stating that even a short pause in glucose and oxygen inflow to the brain causes its damage and chronic deficiency in these substrates can ultimately lead to irreversible brain damage affecting cell viability and activity, and further promoting permanent cognitive impairment [23,24]. Understanding the mechanisms through which aerobic glucose metabolism is disturbed in the brain is clearly of paramount importance.

Many mechanisms are involved in impaired glucose oxidation in the brain affected by AD [25]. It is noteworthy, however, that earlier studies were mostly conducted using autopsy brains from AD patients as the object of examination [26,27], making it difficult to unravel the true mechanisms of the failure in brain energy metabolism, since many factors influence results obtained using postmortem brain tissues, including the agonal stage, brain storage time and temperature protocol, and lactate accumulation [28, 29]. Therefore, no possible correlation between the ascertained causes of the impaired glucose utilization and clinical symptoms of the disease has been identified [26]. By contrast, with functional brain imaging techniques for measuring brain metabolic fluxes in vivo [30,31] it is possible not only to identify the brain regions with significant reductions in glucose metabolism but also to predict memory impairment in people with normal cognitive functions long before the clinical symptoms appear [32–36]. Thus, there is evidence supporting the view that the reduction in blood-carrying energy substrates, particularly glucose and oxygen observed in vivo in AD, results directly from cerebrovascular pathology [37]. This ultimately leads to energy collapse followed by the functional brain failure, and as a consequence, cognitive impairment [38,39].

Another essential factor contributing to brain oxygenation is the oxygen-carrying capacity of red blood cells. It is known that the role of RBCs in tissue oxygen delivery depends on their intracellular metabolism, primarily via energy metabolism and antioxidant status [40,41], which regulate hemoglobin oxygen affinity [42] and NO-dependent hypoxic vasodilation [43–45].

Numerous studies have shown that normal RBC parameters might be significantly affected in AD patients. For instance, in AD patients RBCs have been found to exhibit morphological changes [46,47], leading to decreased deformability [48], perturbations in the physical state of membrane proteins [49,50], and oxidant/antioxidant equilibrium [51–54]. These changes are in some way due to the destabilization of intracellular metabolism in erythrocytes [55–58]. However, the mechanisms underlying the foregoing alterations in RBC metabolism in AD patients are not fully understood. Furthermore, AD is still thought to be exclusively a brain disorder, so a link between disruptions of the endogenous metabolic pathways in circulating erythrocytes that slow down oxygen release, and brain energy crisis manifested as poor brain aerobic glucose oxidation which may precede neurodegeneration is not sufficiently discussed in the literature.

In this article, we review current evidence in support of the widely known amyloid cascade hypothesis and seek to uncover new perspectives on the cause of sporadic AD development, considering brain amyloids outside pathology and suggesting a new mechanism by which neurodegeneration may occur.

Based on our and available published data [46–54] we hypothesize that abnormalities in endogenous glycolytic, antioxidant, and transport pathways in circulating erythrocytes in individuals entering old age [59] disrupt red blood cell function associated with oxygen supply to the brain. This causes deterioration in the aerobic oxidation of glucose leading to the nerve cell degeneration and impaired cognitive processes observed in AD [25].

This assumption is not in contrast with the vascular hypothesis predicting neurodegeneration as a result of impaired blood circulation and brain hypoperfusion [60–62]. On the other hand, it clearly indicates the possible existence of another unknown mechanism which restricts oxygen supply to the brain, and therefore participates in the development of hypoxia and neurodegenerative processes specific to AD.

This suggestion provides a basis for developing individual innovative technologies to restore energy metabolism, the antioxidant system and other impaired endogenous biochemical pathways in erythrocytes [63]. This will help improve RBC function to provide an adequate oxygen supply to the brain and normalize aerobic glucose utilization.

#### 2. Evidence that Contradicts the Fundamental Principles of the Amyloid Cascade Hypothesis

#### 2.1. Brain Amyloid Plaques and Soluble Aβ Oligomers are Unspecific for AD

The fact that cerebral amyloidosis is generated during many human diseases has been known for more than a century, when amyloids (A $\beta$ ) were found in the brain and cerebral vessels of a young woman with cognitive dysfunction caused by traumatic injury to the head [64]. Almost simultaneously, researchers discovered that A $\beta$  deposits are formed and stored in the brain in patients with other pathologies [65,66].

These facts indicate that cerebral amyloidosis does not depend on age and may occur in people without dementia. The latest research results are consistent with data obtained in previous studies which indicate that  $A\beta$  peptides are non-specific for AD and accumulate in the brain of children and adults with many acute and chronic central nervous system disorders [67–75], as well as in normal non-demented elderly persons [76,77]. Further, in some individuals,  $A\beta$  levels in the brain are comparable to or even higher than is typical in AD patients [78,79]. Moreover, a close correlation between severity of cognitive dysfunction and total amyloid volume in the brain of AD patients is not generally detectable [80–84]. Taken together, these results suggest that neither the density of amyloid plaques nor the absolute amount of amyloids can predict cognitive dysfunction impairment or disease onset [85]. These findings contradict the amyloid cascade hypothesis, which states that "deposition of amyloid  $\beta$  protein, the main component of the plaques, is the causative agent of Alzheimer's pathology and that the formation of neurofibrillary tangles, cell loss, vascular damage, and dementia follow as a direct result of this deposition" [20].

Soluble A $\beta$  oligomers, which according to the modified amyloid cascade hypothesis are the initiators of the fatal cascade in AD [86,87], have also been detected in the brains of healthy elderly individuals [88,89]. Moreover, the increase in soluble A $\beta$  level in AD patients compared with healthy normal subjects is negligible [90]. At the same time there is evidence indicating that the accumulation of soluble A $\beta$  oligomers in the brains of patients with AD is episodic with no direct correlation between this accumulation and the appearance of clinical symptoms of the disease [91]. In addition, after the discovery of soluble A $\beta$  oligomers in brains of patients with Down's syndrome [92] and traumatic brain injury [93,94] it can be assumed that soluble A $\beta$ s like aggregated A $\beta$  peptides are unspecific for AD prompting a search for new pathogenic factors.

## 2.2. *Aβ* Neurotoxicity: Which Peptide Is a Therapeutic Target

The toxicity of amyloid peptides is the main argument supporting the amyloid cascade hypothesis. However, it must be acknowledged that data on toxicity of amyloid peptides are obtained exclusively from in vitro experiments [95–97], while participation of amyloids in neuronal death has not yet been demonstrated in vivo.

Indeed, numerous studies conducted in vitro have shown that amyloids induce changes in neurons leading to numerous disorders incompatible with cell vitality. As an example, in primary neuronal cultures neurotoxicity is associated with channel formation in the membrane [98,99], increased metal-dependent production of reactive oxygen species, and oxidative stress [100]. Additionally, interaction between amyloids and cells leads to changes in activity of membrane-bound proteins [101] and receptors [102], in dynamic properties of lipids [103], and loss of phospholipid

asymmetry and integrity [104], that in turn increases membrane fluidity [105] and permeability [106]. The damaging effect on the membrane structures causes dysfunction of the ion pumps and a consequent increase in intracellular calcium [107] triggering a glutamate-dependent pathological cascade that results in further cell death [108]. Interestingly, in vitro both soluble A $\beta$  oligomers and aggregated A $\beta$  forms induce cell death in virtually any cultured cell type, and cell death occurs within a few minutes of exposure to amyloids. A number of questions arise about the significance of these data. For example, if amyloids are so toxic that even a short-term interaction between the cell and amyloid

causes the death of the former, how can these amyloids be "silent" in the brain of AD patients for so long time (15–25 years) before the appearance of the first clinical symptoms of the disease [109,110]? If amyloids are toxic, how can people with this disease live to a very old age and make up the largest group of total cases people aged over 95 [111]? And more, if amyloids located in the brain of elderly individuals with normal memory are toxic why do these toxic substances not affect intellectual capacity, work ability and cognitive processes to a ripe old age? It is clear that the in vitro study results do not always provide easy answer to any of the questions raised.

The issue of amyloid toxicity was not clarified by findings from studies carried out in vivo either. It has been shown that isolated plaque cores from postmortem AD brains [112] and synthetic A $\beta$  fragments produced AD-like neuropathology in vivo [113,114]. A similar effect was observed for soluble A $\beta$ s [115], which according to the reformulated "oligomeric amyloid hypothesis" are the principal neurotoxins [116]. In addition, both peptides disrupted learned behavior in normal animals [117,118]. These observations again raised the question of which hypotheses is correct and what peptide should be a therapeutic target, to which the answer would appear to be an aggregated one as all transgenic mouse models [119] as well as transgenic *Drosophila* models [120], or even transgenic strain of the nematode *Caenorhabditis elegans* [121], used for screening of anti-amyloid drugs, develop only A $\beta$  aggregates [122], ultimately leading to plaque formation [123].

The amyloid cascade hypothesis [20] and the oligomeric amyloid hypothesis [116] evidently contradict each other [124], thereby lowering the level of a huge body of data obtained via in vitro and in vivo studies. Thus, the question remains open as to the toxicity of amyloids in vivo and the expediency of designing new drugs based on contradictory hypotheses which are associated with a pathogenic factor whose toxicity has not yet been proven [22].

## 2.3. Can Transgenic Rodents Be Used as a Suitable Model for Sporadic AD

Returning to the amyloid cascade hypothesis, which was originally postulated for the genetic form of the disease [21], it should be kept in mind that detection of APP and presenilin gene mutations [125] in the genetic form of AD was the only strong evidence underlying the amyloid hypothesis [116]. However, there was no evidence proving that amyloids are the only cause of this disease, especially its sporadic form [123], in which there are no such gene mutations, but instead increased amyloid synthesis which leads to accumulation, aggregation and amyloid plaque formation, characteristic of genetic disease. Interestingly, that by creating an amyloid cascade hypothesis, D. Hardy assumed that "The mutations in APP so far described are responsible only for a small proportion of cases of Alzheimer's disease. Indeed, most cases of Alzheimer's seem to occur in a sporadic fashion, suggesting that there must be other causes of the disease. The cascade hypothesis suggests that other causes of Alzheimer's act by initially triggering APP deposition. This deposition could be caused by an induction of the APP gene through an interleukin-mediated stress response because APP increases in response to a number of neuronal stresses" [20]. A review of the literature sheds no light on when or by whom the role of genetic mutations was first considered in the pathogenesis of sporadic form of AD. Apparently confusion has arisen concerning the hypothesis: it was proposed only for the genetic form of AD, and all anti-amyloid preparations were tested on transgenic mice, using the so-called genetic mouse models of AD, but clinical trials only recruited patients with sporadic disease [21], without mutations in the abovementioned genes. Further, despite the fact that all types of transgenic models perfectly imitate the amyloid aspect of AD [126], the relationship

postulated by the amyloid cascade hypothesis between enhanced amyloid formation and accumulation of hyperphosphorylated tau protein, memory impairment, and neuronal death, is not generally confirmed, using transgenic models [127–129]. Moreover, the memory impairment and neuron death that occur before the appearance of amyloids in the brains of transgenic animals [130,131] may indicate that artificially introducing alien genetic information into the genome of animals might have unexpected pathological effects on the brain and the body as a whole. This prediction is verified by the hippocampal (and other brain regions) hypometabolism and atrophy observed prior to plaque formation [132] and premature deaths in transgenic mice [133], most probably due to the cerebral energy crisis resulting from artificial introduction of alien transgene rather than from accumulated amyloid peptides [134]. Moreover, this fact points to the presence of unrevealed problems that usually arise when creating transgenic models. In fact, overexpression of APP in transgenic animals can dramatically disrupt the mitochondrial function [135] resulting in enhanced oxidative stress [136], uncontrolled release of neurotransmitters [137] and cell death via the apoptotic pathway. This indicates a need to identify the pathological changes occurring in the brains of transgenic animals prior to augmented amyloid formation and whether changes in the behavioral responses of transgenic animals result from the accumulation of amyloids, rather than APP-inducing brain hypometabolism. The genesis of degenerative processes and memory impairment before the appearance of amyloids in the brain, and the premature death of transgenic animals preclude using these animals as adequate models of sporadic AD [126]. This view is supported by a large body of clinical trials showing negative effects, and even death in sporadic AD patients who received anti-amyloid preparations [22,138–140], despite their therapeutic effect having been confirmed in transgenic mice [141–143]. It is clear that any animal model has its advantages and disadvantages, but patients' lives depend on a model that meets the main requirement of equivalent action in animal models and patients and development of safety drugs.

Certainly, the approach in research to the problems of the sporadic form of AD seems poorly planned demonstrating a logical gap in the "hypothesis–objective–test" chain which leads to a focus only on amyloids. In fact, it is clear that if  $A\beta$  deposition occurs in response to cerebral hypometabolism in even young transgenic animals [132], and in elderly AD individuals [144,145], a search for factors that trigger pathological processes in murine and human brains, which in turn lead to APP overexpression and  $A\beta$  accumulation, is of crucial importance in current biomedical research.

#### **3.** Brain Energy Crisis and Aβ Accumulation: Cause or Consequence

Glucose is the main brain energy metabolite, the precursor of substrates that are oxidized in the mitochondrial tricarboxylic acid cycle to form ATP [146]. Glycogen is the only endogenous source of glucose in the brain, but since its reserves are extremely small and the brain is unable to synthesize glucose de novo quickly enough [147], brain cell viability and its myriad functions are completely dependent on a continuous supply of glucose via blood flow.

In addition to ATP, other important energy substrates, for example, lactate produced by glucose metabolism are used by neurons as an additional energy substrate when metabolic activity is high [148,149]. Glucose carbon can be included in lipids [150], proteins [151], and glycogen [152] and is also a precursor of some neurotransmitters such as  $\gamma$ -aminobutyric acid, GABA [153], glutamate [154], and acetylcholine [155] as well as nicotinamide adenine dinucleotide phosphate reduced (NADPH) essential for maintaining brain antioxidant capacity [156].

Thus, glucose entering the brain from the blood is the main substrate known to regulate numerous brain functions, including brain activity, learning, memory [157], and nerve cell viability in general [158,159].

Glucose oxidation in the brain is suppressed with age [160,161], but in individuals without chronic diseases or silent brain pathology, a decrease in metabolic rate usually comes with almost no symptoms, and elderly people can live long enough in relatively good physical and mental health. By contrast, in sporadic AD aerobic oxidation of glucose and the rate of ATP formation in the brain decrease

dramatically [162] resulting in damage to nerve cells and influencing neuronal function and mental states [163,164].

As is known, neurons in different brain structures are not equally sensitive to glucose shortage. Therefore, it is not a surprise that in AD all glucose-dependent neurons, localized in the hippocampus and neocortex [165], involved in the process of long-term memory storage [166], are damaged to a large extent in a state of glucose deprivation. Interestingly, significant accumulation of amyloid peptides is revealed in these brain regions in AD patients [167] suggesting that glucose deficiency is related to increased amyloid formation in the brain.

It is obvious that the causes of amyloid proliferation in the brain in the sporadic form of AD should differ from those described for the genetic form of the disease, if only because APP and presenilin gene mutations are not observed in sporadic AD.

Assuming there is a relationship between disturbances in glucose metabolism in the brain and amyloid formation, the question then arises as to which process occurs first: the formation of amyloids, which have toxic effects on neurons and disturb all types of metabolism, including energy thus resulting in neurodegeneration; or formation of amyloids in response to chronic glucose hypometabolism which serve as a beacon, signaling disturbances in energy metabolism that trigger neuronal and cognitive dysfunction [168]. However, based on the assumption that the disruption of glucose utilization in the brain is a primary cause of neurodegeneration, it is expected that APP up-regulation and accelerated Aβ formation and accumulation should occur irrespective of age or cause of glucose deficiency. Taking the predominant amyloid cascade hypothesis into account, this assumption is at least a little controversial, but it is confirmed by a huge body of evidence showing that APP overexpression, leading to intensive AB production, is observed in multiple pathologies associated with the breakdown of energy metabolism in the brains both in humans and animals. Thus, enhanced amyloid formation and accumulation occurs in the brains of people of different ages during: hypoxia [169], human and animal head trauma [170,171], spinal cord injury [172,173], neuroinflammation [174,175], middle cerebral artery occlusion [176], general anesthesia [177], influence of bacterial agents [178,179] and other examples [25]. In fact, analysis of the above pathologies leads directly to the conclusion that there is steady damage of brain cells resulting from a number of different causes. For example, anesthesia can significantly disturb hemodynamics, reducing the brain's oxygen and glucose absorption rate [180] thereby leading to severe hypoxia, a sharp inhibition of aerobic oxidation and neuronal activity [181]. Interestingly, increased APP processing and amyloid formation occur in the brain a few hours after administering isoflurane as an anesthetic [182]. Attenuated intake of glucose and oxygen to the brain together with the oxidative stress observed in ischemia-reperfusion also initiates APP processing and amyloid formation in damaged brain structures [183]. Hypoglycemia-limited brain glucose intake and also promote amyloid accumulation [184]. Taken together, these data confirm that  $A\beta$  peptides could accumulate in the brain any time when brain cells are destroyed due to a paucity of energy resources [144,185]. Some scientists therefore believe that the initial A $\beta$  formation can be considered an adaptive reaction [186,187], protecting neurons against further damage [188]. If this hypothesis is true, we must accept that the formation of amyloid plaques in the AD brain is not the start of a suicide program, but rather a signal indicating a disruption of brain metabolism and maybe even not the beginning, but the continuation of pathological process that may have gone unnoticed for several years at least [189]. Thus, unveiling the mechanisms of glucose metabolism disruption that underlie APP up-regulation in the sporadic form of AD is a task of current concern in medicine and biology as a precursor to identifying risk factors for disease development and detecting new, alternative targets (other than amyloids) for therapeutic effect.

# 4. The Possible Role of Erythrocytes in Neuronal Aerobic Glucose Metabolism Crisis in AD: The Erythrocytic Hypothesis

Erythrocytes are the only cells able to carry oxygen and maintain aerobic utilization of glucose in tissue. In fact, many people are already well informed about red blood cells in a simplified way,

viewing them only as small bags filled with hemoglobin that binds oxygen reversibly. Notably, a remarkable series of studies has previously demonstrated a pivotal relationship between RBC metabolism and binding, transport, and delivery of oxygen to tissues. Curiously, the first reports about this relationship were published more than 40 years ago [40] suggesting that these scientific findings are largely ignored. As a consequence, there is limited information regarding the interactions between metabolism and erythrocyte function. Traditionally, red blood cell indices, hemoglobin quantities, hematocrit test, sedimentation rate, average volume, and morphological changes etcetera are used to diagnose different diseases, but it is unknown how changes outside the normal range in the above-measured parameters affect RBCs' ability to deliver oxygen to tissues. It follows then that there is no tool for diagnosing metabolic illness in RBCs themselves, a disorder that may appear influenced by pathological factors present in circulation [190].

Passing through the lungs, RBCs are exposed to oxidative stress, in the kidneys which are characterized by hypertonic condition, the cells are the target of osmotic shock. To get through tight capillaries, erythrocytes experience significant mechanical compression, having to pass through capillaries with diameters smaller than their own [191]. It has been proven that cells with impaired intracellular energy metabolism are unable to withstand such exogenous attacks, becoming damaged and lysed directly into the bloodstream as evidenced by the permanent presence of cell-free plasma hemoglobin [192].

Another consequence of erythrocyte "disorder" is an increase in oxygen affinity to hemoglobin, leading to the tissue hypoxia [193] and, in particular, cerebral hypoxia. In order tofulfil their function, RBCs should be healthy which depends primarily on their metabolic and antioxidant status contributing to adequate oxygen binding capacity. For this, all metabolic pathways in mature erythrocytes should be strictly regulated and proceed at the rate necessary to maintain their viability and functional activity. One of these processes is anaerobic glycolysis, the only source of ATP production. Pentose phosphate shunt is necessary for NADPH formation, used in combined enzymatic reactions that catalyze glutathione reductase and glutathione peroxidase. These enzymes are antioxidant and responsible for maintaining low levels of hydrogen peroxide and therefore preventing methemoglobin formation. Erythrocytes also need sufficiently high activity of other antioxidant enzymes such as superoxide dismutase, which maintains a safe level of superoxide radical ( $O_2^{\bullet-}$ ), glutathione transferase, which is involved in xenobiotic detoxification [194], catalase and peroxiredoxin-2-peroxidase, which destroy hydrogen peroxide. Red blood cells are not able to synthesize ATP de novo, and energy homeostasis is maintained during high energy demand due to salvage pathways by which already existing nucleosides and purine bases can be recycled to produce adenine nucleotide triphosphates [195].

For the production of 2,3-diphosphoglycerate (2,3-DPG), the main metabolite that facilitates the release of oxygen from hemoglobin to tissue [196], and to maintain its steady state concentration, erythrocytes possess a unique glycolytic bypass called a Rapoport–Luebering shunt [42]. The activity of this pathway depends on the rate of glycolysis and a relative deficiency of enzymes catalyzing the initial reactions of glycolysis may cause insufficient synthesis of 2,3-DPG and affect the ability of RBC to release oxygen, leading to tissue hypoxia [197]. Therefore, intact intracellular metabolic pathways of erythrocytes are undoubtedly a major factor responsible for adequate oxygen delivery and release to tissues.

We have recently demonstrated differences in the parameters of various metabolic pathways in erythrocytes of patients with Alzheimer's disease (AD group) and patients with non-Alzheimer's dementia (NA group). We found that in erythrocytes of elderly control subjects (AMC, age-matched controls), glycolysis and cation transport were sharply inhibited, as evidenced by a significant decrease in all glycolytic enzyme activities, a decrease in the rate of glucose consumption, ATP concentration, the rate of pyruvate and lactate formation, and a sharp decrease in Na<sup>+</sup>, K<sup>+</sup>-ATP-ase activity compared to young adult controls (YAC). In the erythrocytes of AD and NA patients, these parameters changed to the same extent and significantly exceeded the parameters not only of AMC, but also of the YAC group indicating an increase in ion fluxes leading to acceleration of RBC glycolytic pathway [198,199].

Given that the active transport of cations through the erythrocyte membrane is controlled by the rate of ATP formation during glycolysis, up-regulation of Na<sup>+</sup>, K<sup>+</sup>-ATP-ase accompanied by a decrease in the concentration of ATP in RBCs of AD patients can be attributed to an imbalance between ATP formation and ion pumping, caused by increases in Na<sup>+</sup> input in erythrocytes of AD patients [200]. Despite glycolysis activation, this imbalance ultimately promotes an increase in adenylate catabolism as confirmed by ADP and AMP accumulation, a decrease in the cellular energy charge and the total content of adenine nucleotides. The enzyme activity of the Rapoport–Luebering shunt, in particular, diphosphoglycerate phosphatase (DPGP-ase), that regulated the steady state levels of 2,3-DPG, increased simultaneously with a significant decrease in 2,3-DPG concentration in erythrocytes of the AD group compared to both age-matched controls (AMC) and young adult controls (YC). Clearly, the affinity of hemoglobin for oxygen is influenced not only by 2,3-DPG, but most likely by other factors such as pH level,  $PCO_2$ ,  $PO_2$ ,  $CI^-$ , conformations of hemoglobin and temperature. However, the correlation between the concentration of 2,3-DPG in erythrocytes and the degree of hypoxia in tissues, and particularly in the brain, has been revealed in many pathological states of human beings and animals [201–203]. Taken together, the results obtained suggest that chronic intensification in the rate of cationic traffic and plasma membrane permeability observed in the RBCs of AD patients, could trigger a cascade of pathological reactions in erythrocytes. This then leads to accelerated hydrolysis of ATP and 2,3-DPG, responsible for regulating the affinity of hemoglobin for oxygen, one possible explanation for inadequate oxygen supply to the brain, glucose hypometabolism, and cognitive abnormalities in AD. We have also found that changes in many metabolic pathways are already present in the erythrocytes of healthy individuals entering old age but without memory problems. Thus, the cells of the AMC group were marked by disturbed homeostasis of adenine nucleotides, a slight but significant decrease in 2,3-DPG concentration [59]. Similarly, there were no differences in pro-oxidant levels [199] between AMC, AD, and NA groups, while antioxidant enzyme activity, considerably reduced in RBCs of the AMC subjects, further decreased with dementia [198].

Our findings are largely consistent with data from other researchers who show the same direction of disturbances in erythrocytes of normal elderly people and patients with AD [204]. They also indicate that the disturbed metabolism and reduced antioxidant capacity in RBCs are not hallmarks of the terminal illness, but are indicative of stable and permanent changes in intracellular metabolism in erythrocytes of the elderly, which under the influence of unknown factors may lead to the pathological lesions encountered in AD patients. One factor very likely to contribute to altered intracellular metabolic pathways and RBC morphology in older people is enhanced entry of Na<sup>+</sup> into the cells [205].

It is clear, therefore, that increased activity of Na<sup>+</sup>, K<sup>+</sup>-ATP-ase in the erythrocytes of AD patients can be considered an adaptive reaction to the increased entry of this cation into cells. At the same time, in chronic conditions, constant and multiple activation of the enzyme cannot completely compensate for the increased plasma membrane permeability that leads to pathological consequences which negatively influence RBCs' energy metabolism and antioxidant defence system, enhancing the hydrolysis of ATP, 2,3-DPG, oxidative stress and proteolysis. The combined effect of this damage disrupts the function of erythrocytes associated with oxygen delivery, and also causes changes in the morphological property of erythrocytes, decreasing their ability to deform and thus lowering the threshold for development of neuropathology.

Our erythrocytic hypothesis is thus based on the obvious link between the state of intracellular metabolism of erythrocytes and its functional ability. We suggest that abnormalities in glycolytic, antioxidant and transport pathways can disrupt the erythrocyte function associated with oxygen delivery to the brain and cause disruption of the aerobic oxidation of glucose, leading to degeneration of the nerve cells and impaired cognitive function as observed in AD.

It is obviously necessary to elucidate what factors (endogenous and exogenous) cause the premature decrease in the reserve potential of erythrocytes to withstand stress that they constantly undergo, while circulating from the lungs to the tissues.

#### 5. Encapsulation of Missing Enzymes in Erythrocytes: A Way to Reconstruct Energy Metabolism

Erythrocytes are known as cells that can be loaded with different kinds of substances (drugs, enzymes, metabolites, cofactors, etc.) through methods based on the hypotonic hemolysis-resealing procedure that allows to design intact and fully viable erythrocytes [206,207]. Based on the above methodology we developed a procedure for encapsulating missing enzymes together with the substrates and cofactors involved in the energy metabolism and antioxidant defense system. Our findings show that after reinjection into rat blood, enzyme-loaded erythrocytes were able to retain their integrity, normal energy metabolism, and survive in blood circulation for a long time [25].

This biotechnological approach provides an extraordinary opportunity to develop innovative medications for individual use that will help specifically to treat the endogenous metabolic abnormalities in the red blood cells and restore adequate oxygen supply to the brain.

### 6. Conclusions

More than 100 years have passed since Alzheimer's disease was identified, yet the pathogenesis of the disease remains unknown. Taking into account the role of the vascular hypothesis in AD pathogenesis, we assume that among the additional causes of pathological brain hypoperfusion typical in AD are structural and biochemical changes in erythrocytes, leading to inadequate oxygen supply to the brain, which inevitably precedes brain atrophy, developmental disabilities and dementia. Therefore, change in biochemical parameters in erythrocytes can be considered both an indicator of the cell state in the central nervous system, and a new target to develop innovative methods to treat this systemic disease.

The question of whether cerebral vascular damage or RBC damage is the most important cause of cerebral hypoxia is beyond the scope of this study. It is needless to say that both pathologies are intrinsically closely related to each other. However, since erythrocytes serve as the only oxygen carriers and their ability to bind, transport and deliver oxygen to tissues depends, primarily, on energy metabolism and antioxidant systems, we believe that disturbance of the endogenous processes in these cells most likely has a dramatic destabilizing impact on aerobic glucose metabolism in the brain and promotes dementia development. This assumption does not contradict the known pathological consequences of vascular damage, that leads to impaired blood circulation and brain hypoperfusion. At the same time, it clearly indicates the possible existence of additional unspecified mechanisms, restricting oxygen supply to the brain and therefore participating in the development of hypoxia and neurodegenerative processes specific to AD.

We therefore suggest that a combination of revascularization and restoration of metabolism in erythrocytes which maintain glucose/oxygen-dependent normal neuronal function could prevent brain cells from the destructive effects of energy limitation and hypoxia, and avoid dementia development. A thorough and detailed study of erythrocyte metabolism and morphology is needed not only to identify potential risk factors, but also to shed light on the molecular mechanisms that limit oxygen supply to the brain. This knowledge is important to prevent the development of hypoxia-induced neurodegenerative processes typical not only of AD, but also of many other diseases with different etiologies, and characterized by varying degrees of cognitive dysfunctions.

**Author Contributions:** E.K., L.T., C.M. provided the original conception and designed the study; E.K., L.T., A.U., G.A. analysed data and discussed the results. The manuscript was written by E.K. and approved by all the authors. All authors have read and agreed to the published version of the manuscript.

**Funding:** The study reported was funded by RFBR through the research project No 18-015-00106 and Ministerio de Ciencia e Innovación-Instituto de Salud Carlos III (FIS PI18/00150) co-funded with European Regional Development Funds (ERDF) and Fundación Ramón Areces.

**Conflicts of Interest:** The authors declare no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

## References

- Miklossy, J.; Taddei, K.; Martins, R.; Escher, G.; Kraftsik, R.; Pillevuit, O.; Lepori, D.; Campiche, M. Alzheimer disease: Curly fibers and tangles in organs other than brain. *J. Neuropathol. Exp. Neurol.* 1999, *58*, 803–814. [CrossRef]
- 2. Blass, J.P.; Hanin, I.; Barclay, L.; Kopp, U.; Reding, M.J. Red blood cell abnormalities in Alzheimer disease. *J. Am. Geriatr. Soc.* **1985**, *33*, 401–405. [CrossRef]
- 3. Kiko, T.; Nakagawa, K.; Satoh, A.; Tsuduki, T.; Furukawa, K.; Arai, H.; Miyazawa, T. Amyloid β levels in human red blood cells. *PLoS ONE* **2012**, *7*, e49620. [CrossRef] [PubMed]
- 4. Li, Q.X.; Whyte, S.; Tanner, J.E.; Evin, G.; Beyreuther, K.; Masters, C.L. Secretion of Alzheimer's disease Abeta amyloid peptide by activated human platelets. *Lab. Invest.* **1998**, *78*, 461–469. [PubMed]
- 5. Gibson, G.E.; Huang, H.M. Oxidative processes in the brain and non-neuronal tissues as biomarkers of Alzheimer's disease. *Front. Biosci.* **2002**, *7*, d1007–1015. [CrossRef] [PubMed]
- 6. Bird, T.D. Genetic factors in Alzheimer's disease. N. Engl. J. Med. 2005, 352, 862–864. [CrossRef] [PubMed]
- Sosa-Ortiz, A.L.; Acosta-Castillo, I.; Prince, M.J. Epidemiology of dementias and Alzheimer's disease. Arch. Med. Res. 2012, 43, 600–608. [CrossRef] [PubMed]
- 8. Appel, S.H. A unifying hypothesis for the cause of amyotrophic lateral sclerosis, parkinsonism, and Alzheimer disease. *Ann. Neurol.* **1981**, *10*, 499–505. [CrossRef] [PubMed]
- 9. Bartus, R.T.; Dean, R.L., 3rd; Beer, B.; Lippa, A.S. The cholinergic hypothesis of geriatric memory dysfunction. *Science* **1982**, 217, 408–414. [CrossRef]
- 10. Blass, J.P.; Zemcov, A. Alzheimer's disease. A metabolic systems degeneration? *Neurochem. Pathol.* **1984**, 2, 103–114. [CrossRef]
- 11. Beal, M.F. Does impairment of energy metabolism result in excitotoxic neuronal death in neurodegenerative illnesses? *Ann. Neurol.* **1992**, *31*, 119–130. [CrossRef] [PubMed]
- 12. Lees, G.J. Contributory mechanisms in the causation of neurodegenerative disorders. *Neuroscience* **1993**, *54*, 287–322. [CrossRef]
- 13. Markesbery, W.R. Oxidative stress hypothesis in Alzheimer's disease. *Free Radic. Biol. Med.* **1997**, 23, 134–147. [CrossRef]
- 14. Guillemin, G.J.; Williams, K.R.; Smith, D.G.; Smythe, G.A.; Croitoru-Lamoury, J.; Brew, B.J. Quinolinic acid in the pathogenesis of Alzheimer's disease. *Adv. Exp. Med. Biol.* **2003**, *527*, 167–176. [PubMed]
- 15. Tuppo, E.E.; Arias, H.R. The role of inflammation in Alzheimer's disease. *Int. J. Biochem. Cell Biol.* **2005**, *37*, 289–305. [CrossRef]
- 16. Lee, H.G.; Zhu, X.; Nunomura, A.; Perry, G.; Smith, M.A. Amyloid beta: The alternate hypothesis. *Curr. Alzheimer Res.* **2006**, *3*, 75–80. [CrossRef]
- 17. Moreira, P.I.; Cardoso, S.M.; Santos, M.S.; Oliveira, C.R. The key role of mitochondria in Alzheimer's disease. *J. Alzheimers Dis.* **2006**, *9*, 101–110. [CrossRef]
- 18. Rolston, R.K.; Perry, G.; Zhu, X.; Castellani, R.J.; Dwyer, B.E.; Lee, H.G.; Petersen, R.B.; Smith, M.A. Iron: A Pathological Mediator of Alzheimer Disease? *Agro Food Ind. Hi-Tech* **2009**, *19*, 33–36.
- 19. Manyevitch, R.; Protas, M.; Scarpiello, S.; Deliso, M.; Bass, B.; Nanajian, A.; Chang, M.; Thompson, S.M.; Khoury, N.; Gonnella, R.; et al. Evaluation of Metabolic and Synaptic Dysfunction Hypotheses of Alzheimer's Disease (AD): A Meta-Analysis of CSF Markers. *Curr. Alzheimer Res.* **2018**, *15*, 164–181. [CrossRef]
- 20. Hardy, J.A.; Higgins, G.A. Alzheimer's disease: The amyloid cascade hypothesis. *Science* **1992**, 256, 184–185. [CrossRef]
- 21. Hardy, J.; Mayer, J. The amyloid cascade hypothesis has misled the pharmaceutical industry. *Biochem. Soc. Trans.* **2011**, *39*, 920–923. [CrossRef]
- 22. Kurkinen, M. The amyloid hypothesis is too good to be true. *Alzheimers Dement. Cogn. Neurol.* **2017**, *1*, 1–9. [CrossRef]
- 23. Blass, J.P. Glucose/mitochondria in neurological conditions. Int. Rev. Neurobiol. 2002, 51, 325–376. [PubMed]
- 24. Blass, J.P.; Gibson, G.E. Cerebrometabolic aspects of delirium in relationship to dementia. *Dement. Geriatr. Cogn. Disord.* **1999**, *10*, 335–338. [CrossRef] [PubMed]
- 25. Kosenko, E.A.; Tikhonova, L.A.; Montoliu, C.; Barreto, G.E.; Aliev, G.; Kaminsky, Y.G. Metabolic Abnormalities of Erythrocytes as a Risk Factor for Alzheimer's Disease. *Front. Neurosci.* **2018**, *11*, 728. [CrossRef] [PubMed]

- 26. Iwangoff, P.; Armbruster, R.; Enz, A.; Meier-Ruge, W. Glycolytic enzymes from human autoptic brain cortex: Normal aged and demented cases. *Mech. Ageing Dev.* **1980**, *14*, 203–209. [CrossRef]
- 27. Bigl, M.; Brückner, M.K.; Arendt, T.; Bigl, V.; Eschrich, K. Activities of key glycolytic enzymes in the brains of patients with Alzheimer's disease. *J. Neural Transm. (Vienna)* **1999**, *106*, 499–511. [CrossRef]
- 28. Perry, E.K.; Perry, R.H.; Tomlinson, B.E. The influence of agonal status on some neurochemical activities of postmortem human brain tissue. *Neurosci. Lett.* **1982**, *29*, 303–307. [CrossRef]
- 29. Yates, C.M.; Butterworth, J.; Tennant, M.C.; Gordon, A. Enzyme activities in relation to pH and lactate in postmortem brain in Alzheimer-type and other dementias. *J. Neurochem.* **1990**, *55*, 1624–1630. [CrossRef]
- 30. Duarte, J.M.; Lei, H.; Mlynárik, V.; Gruetter, R. The neurochemical profile quantified by in vivo 1H NMR spectroscopy. *Neuroimage* 2012, *61*, 342–362. [CrossRef]
- Magistretti, P.J.; Allaman, I. A cellular perspective on brain energy metabolism and functional imaging. *Neuron* 2015, *86*, 883–901. [CrossRef] [PubMed]
- 32. Johnson, K.A.; Jones, K.; Holman, B.L.; Becker, J.A.; Spiers, P.A.; Satlin, A.; Albert, M.S. Preclinical prediction of Alzheimer's disease using SPECT. *Neurology* **1998**, *50*, 1563–1571. [CrossRef] [PubMed]
- 33. Ouchi, Y.; Nobezawa, S.; Okada, H.; Yoshikawa, E.; Futatsubashi, M.; Kaneko, M. Altered glucose metabolism in the hippocampal head in memory impairment. *Neurology* **1998**, *51*, 136–142. [CrossRef] [PubMed]
- 34. Killiany, R.J.; Gomez-Isla, T.; Moss, M.; Kikinis, R.; Sandor, T.; Jolesz, F.; Tanzi, R.; Jones, K.; Hyman, B.T.; Albert, M.S. Use of structural magnetic resonance imaging to predict who will get Alzheimer's disease. *Ann. Neurol.* **2000**, *47*, 430–439. [CrossRef]
- Cerami, C.; Della Rosa, P.A.; Magnani, G.; Santangelo, R.; Marcone, A.; Cappa, S.F.; Perani, D. Brain metabolic maps in Mild Cognitive Impairment predict heterogeneity of progression to dementia. *Neuroimage Clin.* 2014, 7, 187–194. [CrossRef]
- Beheshti, I.; Demirel, H.; Matsuda, H. Alzheimer's Disease Neuroimaging Initiative, Classification of Alzheimer's disease and prediction of mild cognitive impairment-to-Alzheimer's conversion from structural magnetic resource imaging using feature ranking and a genetic algorithm. *Comput. Biol. Med.* 2017, 83, 109–119. [CrossRef]
- 37. Perlmutter, L.S.; Chui, H.C. Microangiopathy, the vascular basement membrane and Alzheimer's disease. A review. *Brain Res. Bull.* **1990**, *24*, 677–686. [CrossRef]
- de la Torre, J.C.; Mussivand, T. Can disturbed brain microcirculation cause Alzheimer's disease? *Neurol. Res.* 1993, 15, 146–153. [CrossRef]
- de Leon, M.J.; Convit, A.; Wolf, O.T.; Tarshish, C.Y.; DeSanti, S.; Rusinek, H.; Tsui, W.; Kandil, E.; Scherer, A.J.; Roche, A.; et al. Prediction of cognitive decline in normal elderly subjects with 2-[F]fluoro-2-deoxy-D-glucose/poitron-emission tomography (FDG/PET). *Proc. Natl. Acad. Sci. USA* 2001, *98*, 10966–10971. [CrossRef]
- 40. Brewer, G.J.; Eaton, J.W. Erythrocyte metabolism: Interaction with oxygen transport. *Science* **1971**, 171, 1205–1211. [CrossRef]
- 41. Brewer, G.J.; Oelshlegel, F.J., Jr.; Moore, L.G.; Noble, N.A. In vivo red cell glycolytic control and DPG-ATP levels. *Ann. N. Y. Acad. Sci.* **1974**, *241*, 513–523. [CrossRef] [PubMed]
- 42. van Wijk, R.; van Solinge, W.W. The energy-less red blood cell is lost: Erythrocyte enzyme abnormalities of glycolysis. *Blood* 2005, *106*, 4034–4042. [CrossRef] [PubMed]
- 43. Collins, D.M.; McCullough, W.T.; Ellsworth, M.L. Conducted vascular responses: Communication across the capillary bed. *Microvasc. Res.* **1998**, *56*, 43–53. [CrossRef] [PubMed]
- Crawford, J.H.; Isbell, T.S.; Huang, Z.; Shiva, S.; Chacko, B.K.; Schechter, A.N.; Darley-Usmar, V.M.; Kerby, J.D.; Lang, J.D., Jr.; Kraus, D.; et al. Hypoxia, red blood cells, and nitrite regulate NO-dependent hypoxic vasodilation. *Blood* 2006, 107, 566–574. [CrossRef]
- 45. Ellsworth, M.L.; Ellis, C.G.; Goldman, D.; Stephenson, A.H.; Dietrich, H.H.; Sprague, R.S. Erythrocytes: Oxygen sensors and modulators of vascular tone. *Physiology (Bethesda)* **2009**, *24*, 107–116. [CrossRef]
- 46. Mohanty, J.G.; Eckley, D.M.; Williamson, J.D.; Launer, L.J.; Rifkind, J.M. Do red blood cell-beta-amyloid interactions alter oxygen delivery in Alzheimer's disease? *Adv. Exp. Med. Biol.* **2008**, *614*, 29–35. [CrossRef]
- 47. Mohanty, J.G.; Shukla, H.D.; Williamson, J.D.; Launer, L.J.; Saxena, S.; Rifkind, J.M. Alterations in the red blood cell membrane proteome in alzheimer's subjects reflect disease-related changes and provide insight into altered cell morphology. *Proteome Sci.* **2010**, *8*, 11. [CrossRef]

- 48. Chang, C.Y.; Liang, H.J.; Chow, S.Y.; Chen, S.M.; Liu, D.Z. Hemorheological mechanisms in Alzheimer's disease. *Microcirculation* **2007**, *14*, 627–634. [CrossRef]
- Butterfield, D.A.; Markesbery, W.R. Specificity of biophysical and biochemical alterations in erythrocyte membranes in neurological disorders-Huntington's disease, Friedreich's ataxia, Alzheimer's disease, amyotrophic lateral sclerosis, and myotonic and duchenne muscular dystrophy. *J. Neurol. Sci.* 1980, 47, 261–271. [CrossRef]
- 50. Bosman, G.J.; Bartholomeus, I.G.; de Grip, W.J. Alzheimer's disease and cellular aging: Membrane-related events as clues to primary mechanisms. *Gerontology* **1991**, *37*, 95–112. [CrossRef]
- 51. Serra, J.A.; Famulari, A.L.; Kohan, S.; Marschoff, E.R.; Dominguez, R.O.; de Lustig, E.S. Copper-zinc superoxide dismutase activity in red blood cells in probable Alzheimer's patients and their first-degree relatives. *J. Neurol. Sci.* **1994**, *122*, 179–188. [CrossRef]
- 52. Delibas, N.; Ozcankaya, R.; Altuntas, I. Clinical importance of erythrocyte malondialdehyde levels as a marker for cognitive deterioration in patients with dementia of Alzheimer type: A repeated study in 5-year interval. *Clin. Biochem.* **2002**, *35*, 137–141. [CrossRef]
- 53. Rossi, L.; Squitti, R.; Pasqualetti, P.; Marchese, E.; Cassetta, E.; Forastiere, E.; Rotilio, G.; Rossini, P.M.; Finazzi-Agró, A. Red blood cell copper, zinc superoxide dismutase activity is higher in Alzheimer's disease and is decreased by D-penicillamine. *Neurosci. Lett.* **2002**, *329*, 137–140. [CrossRef]
- 54. Torres, L.L.; Quaglio, N.B.; de Souza, G.T.; Garcia, R.T.; Dati, L.M.; Moreira, W.L.; Loureiro, A.P.; de Souza-Talarico, J.N.; Smid, J.; Porto, C.S.; et al. Peripheral oxidative stress biomarkers in mild cognitive impairment and Alzheimer's disease. *J. Alzheimers Dis.* **2011**, *26*, 59–68. [CrossRef] [PubMed]
- 55. Gov, N.S.; Safran, S.A. Red blood cell membrane fluctuations and shape controlled by ATP-induced cytoskeletal defects. *Biophys. J.* **2005**, *88*, 1859–1874. [CrossRef] [PubMed]
- Forsyth, A.M.; Wan, J.; Owrutsky, P.D.; Abkarian, M.; Stone, H.A. Multiscale approach to link red blood cell dynamics, shear viscosity, and ATP release. *Proc. Natl. Acad. Sci. USA* 2011, *108*, 10986–10991. [CrossRef] [PubMed]
- 57. Leal Denis, M.F.; Alvarez, H.A.; Lauri, N.; Alvarez, C.L.; Chara, O.; Schwarzbaum, P.J. Dynamic Regulation of Cell Volume and Extracellular ATP of Human Erythrocytes. *PLoS ONE* **2016**, *11*, e0158305. [CrossRef]
- 58. Dinarelli, S.; Longo, G.; Dietler, G.; Francioso, A.; Mosca, L.; Pannitteri, G.; Boumis, G.; Bellelli, A.; Girasole, M. Erythrocyte's aging in microgravity highlights how environmental stimuli shape metabolism and morphology. *Sci. Rep.* 2018, *8*, 5277. [CrossRef]
- Kosenko, E.A.; Aliev, G.; Kaminsky, Y.G. Relationship between chronic disturbance of 2,3-diphosphoglycerate metabolism in erythrocytes and Alzheimer disease. CNS Neurol. Disord. Drug Targets 2016, 15, 113–123. [CrossRef]
- 60. de la Torre, J.C. Hemodynamic consequences of deformed microvessels in the brain in Alzheimer's disease. *Ann. N.Y. Acad. Sci.* **1997**, *826*, 75–91. [CrossRef]
- 61. Hoyer, S.; Oesterreich, K.; Wagner, O. Glucose metabolism as the site of the primary abnormality in early-onset dementia of Alzheimer type? *J. Neurol.* **1988**, *235*, 143–148. [CrossRef] [PubMed]
- 62. Blass, J.P.; Gibson, G.E.; Hoyer, S. The role of the metabolic lesion in Alzheimer's disease. *J. Alzheimers Dis.* **2002**, *4*, 225–232. [CrossRef] [PubMed]
- 63. Bax, B.E.; Bain, M.D.; Fairbanks, L.D.; Webster, A.D.; Ind, P.W.; Hershfield, M.S.; Chalmers, R.A. A 9-yr evaluation of carrier erythrocyte encapsulated adenosine deaminase (ADA) therapy in a patient with adult-type ADA deficiency. *Eur. J. Haematol.* **2007**, *79*, 338–348. [CrossRef] [PubMed]
- 64. Atkins, R. On the Morbid Histology of the Spinal Cord in Five Cases of Insanity. *Br. Med. J.* **1878**, *2*, 96–99. [CrossRef] [PubMed]
- 65. Atkins, R. Half-yearly report on mental disease. Dublin J. Med. Sci. 1875, 60, 314–331. [CrossRef]
- 66. Blocq, P.; Marinesco, G. Sur les lésions et la pathogénie de l'épilepsie dite essentielle. *Sem. Med.* **1892**, *12*, 445–456.
- 67. Glenner, G.G.; Wong, C.W. Alzheimer's disease and Down's syndrome: Sharing of a unique cerebrovascular amyloid fibril protein. *Biochem. Biophys. Res. Commun.* **1984**, *122*, 1131–1135. [CrossRef]
- 68. Roberts, G.W.; Allsop, D.; Bruton, C. The occult aftermath of boxing. *J. Neurol. Neurosurg. Psychiatry* **1990**, *53*, 373–378. [CrossRef]
- 69. Arai, H.; Lee, V.M.; Hill, W.D.; Greenberg, B.D.; Trojanowski, J.Q. Lewy bodies contain beta-amyloid precursor proteins of Alzheimer's disease. *Brain Res.* **1992**, *585*, 386–390. [CrossRef]

- 70. Jordan, B.D. Chronic traumatic brain injury associated with boxing. *Semin. Neurol.* **2000**, *20*, 179–185. [CrossRef]
- 71. Smith, D.H.; Chen, X.H.; Iwata, A.; Graham, D.I. Amyloid beta accumulation in axons after traumatic brain injury in humans. *J. Neurosurg.* **2003**, *98*, 1072–1077. [CrossRef] [PubMed]
- 72. Liberski, P.P. Amyloid plaques in transmissible spongiform encephalopathies (prion diseases). *Folia Neuropathol.* **2004**, *42* (Suppl. B), 109–119.
- Omalu, B.I.; DeKosky, S.T.; Minster, R.L.; Kamboh, M.I.; Hamilton, R.L.; Wecht, C.H. Chronic traumatic encephalopathy in a National Football League player. *Neurosurgery* 2005, 57, 128–134, discussion 128–134. [CrossRef] [PubMed]
- 74. Calderón-Garcidueñas, L.; Mora-Tiscareño, A.; Ontiveros, E.; Gómez-Garza, G.; Barragán-Mejía, G.; Broadway, J.; Chapman, S.; Valencia-Salazar, G.; Jewells, V.; Maronpot, R.R.; et al. Air pollution, cognitive deficits and brain abnormalities: A pilot study with children and dogs. *Brain Cogn.* 2008, 68, 117–127. [CrossRef] [PubMed]
- 75. Calderón-Garcidueñas, L.; Solt, A.C.; Henríquez-Roldán, C.; Torres-Jardón, R.; Nuse, B.; Herritt, L.; Villarreal-Calderón, R.; Osnaya, N.; Stone, I.; García, R.; et al. Long-term air pollution exposure is associated with neuroinflammation, an altered innate immune response, disruption of the blood-brain barrier, ultrafine particulate deposition, and accumulation of amyloid beta-42 and alpha-synuclein in children and young adults. *Toxicol. Pathol.* **2008**, *36*, 289–310. [CrossRef] [PubMed]
- 76. Joachim, C.L.; Mori, H.; Selkoe, D.J. Amyloid beta-protein deposition in tissues other than brain in Alzheimer's disease. *Nature* **1989**, *341*, 226–230. [CrossRef]
- 77. Mann, D.M.; Jones, D.; South, P.W.; Snowden, J.S.; Neary, D. Deposition of amyloid beta protein in non-Alzheimer dementias: Evidence for a neuronal origin of parenchymal deposits of beta protein in neurodegenerative disease. *Acta Neuropathol.* **1992**, *83*, 415–419. [CrossRef]
- 78. Funato, H.; Yoshimura, M.; Kusui, K.; Tamaoka, A.; Ishikawa, K.; Ohkoshi, N.; Namekata, K.; Okeda, R.; Ihara, Y. Quantitation of amyloid beta-protein (A beta) in the cortex during aging and in Alzheimer's disease. *Am. J. Pathol.* **1998**, *152*, 1633–1640.
- 79. Blass, J.P. Immunologic treatment of Alzheimer's disease. N. Engl. J. Med. 1999, 341, 1694–1695. [CrossRef]
- Terry, R.D.; Masliah, E.; Salmon, D.P.; Butters, N.; DeTeresa, R.; Hill, R.; Hansen, L.A.; Katzman, R. Physical basis of cognitive alterations in Alzheimer's disease: Synapse loss is the major correlate of cognitive impairment. *Ann. Neurol.* **1991**, *30*, 572–580. [CrossRef]
- 81. Giannakopoulos, P.; Herrmann, F.R.; Bussière, T.; Bouras, C.; Kövari, E.; Perl, D.P.; Morrison, J.H.; Gold, G.; Hof, P.R. Tangle and neuron numbers, but not amyloid load, predict cognitive status in Alzheimer's disease. *Neurology* **2003**, *60*, 1495–1500. [CrossRef] [PubMed]
- 82. Guillozet, A.L.; Weintraub, S.; Mash, D.C.; Mesulam, M.M. Neurofibrillary tangles, amyloid, and memory in aging and mild cognitive impairment. *Arch. Neurol.* **2003**, *60*, 729–736. [CrossRef] [PubMed]
- Erten-Lyons, D.; Woltjer, R.L.; Dodge, H.; Nixon, R.; Vorobik, R.; Calvert, J.F.; Leahy, M.; Montine, T.; Kaye, J. Factors associated with resistance to dementia despite high Alzheimer disease pathology. *Neurology* 2009, 72, 354–360. [CrossRef] [PubMed]
- Karran, E.; De Strooper, B. The amyloid cascade hypothesis: Are we poised for success or failure? *J. Neurochem.* 2016, 139 (Suppl. 2), 237–252. [CrossRef]
- Jack, C.R., Jr.; Lowe, V.J.; Weigand, S.D.; Wiste, H.J.; Senjem, M.L.; Knopman, D.S.; Shiung, M.M.; Gunter, J.L.; Boeve, B.F.; Kemp, B.J.; et al. Alzheimer's Disease Neuroimaging Initiative, Serial PIB and MRI in normal, mild cognitive impairment and Alzheimer's disease: Implications for sequence of pathological events in Alzheimer's disease. *Brain* 2009, *132*, 1355–1365. [CrossRef]
- McLean, C.A.; Cherny, R.A.; Fraser, F.W.; Fuller, S.J.; Smith, M.J.; Beyreuther, K.; Bush, A.I.; Masters, C.L. Soluble pool of Abeta amyloid as a determinant of severity of neurodegeneration in Alzheimer's disease. *Ann. Neurol.* 1999, 46, 860–866. [CrossRef]
- 87. Wirths, O.; Multhaup, G.; Bayer, T.A. A modified beta-amyloid hypothesis: Intraneuronal accumulation of the beta-amyloid peptide—The first step of a fatal cascade. *J. Neurochem.* **2004**, *91*, 513–520. [CrossRef]
- Kuo, Y.M.; Emmerling, M.R.; Vigo-Pelfrey, C.; Kasunic, T.C.; Kirkpatrick, J.B.; Murdoch, G.H.; Ball, M.J.; Roher, A.E. Water-soluble Abeta (N-40, N-42) oligomers in normal and Alzheimer disease brains. *J. Biol. Chem.* 1996, 271, 4077–4081. [CrossRef]

- 89. Tabaton, M.; Piccini, A. Role of water-soluble amyloid-beta in the pathogenesis of Alzheimer's disease. *Int. J. Exp. Pathol.* **2005**, *86*, 139–145. [CrossRef]
- Wang, J.; Dickson, D.W.; Trojanowski, J.Q.; Lee, V.M. The levels of soluble versus insoluble brain Abeta distinguish Alzheimer's disease from normal and pathologic aging. *Exp. Neurol.* 1999, 158, 328–337. [CrossRef]
- 91. de la Torre, J.C. Is Alzheimer's disease a neurodegenerative or a vascular disorder? Data, dogma, and dialectics. *Lancet Neurol.* **2004**, *3*, 184–190. [CrossRef]
- Teller, J.K.; Russo, C.; DeBusk, L.M.; Angelini, G.; Zaccheo, D.; Dagna-Bricarelli, F.; Scartezzini, P.; Bertolini, S.; Mann, D.M.; Tabaton, M.; et al. Presence of soluble amyloid beta-peptide precedes amyloid plaque formation in Down's syndrome. *Nat. Med.* **1996**, *2*, 93–95. [CrossRef] [PubMed]
- DeKosky, S.T.; Abrahamson, E.E.; Ciallella, J.R.; Paljug, W.R.; Wisniewski, S.R.; Clark, R.S.; Ikonomovic, M.D. Association of increased cortical soluble abeta42 levels with diffuse plaques after severe brain injury in humans. *Arch. Neurol.* 2007, *64*, 541–544. [PubMed]
- 94. Johnson, V.E.; Stewart, W.; Smith, D.H. Traumatic brain injury and amyloid-β pathology: A link to Alzheimer's disease? *Nat. Rev. Neurosci.* **2010**, *11*, 361–370. [CrossRef] [PubMed]
- 95. Pike, C.J.; Walencewicz, A.J.; Glabe, C.G.; Cotman, C.W. In vitro aging of beta-amyloid protein causes peptide aggregation and neurotoxicity. *Brain Res.* **1991**, *563*, 311–314. [CrossRef]
- 96. Meske, V.; Hamker, U.; Albert, F.; Ohm, T.G. The effects of beta/A4-amyloid and its fragments on calcium homeostasis, glial fibrillary acidic protein and S100beta staining, morphology and survival of cultured hippocampal astrocytes. *Neuroscience* **1998**, *85*, 1151–1160. [CrossRef]
- 97. Orellana, J.A.; Shoji, K.F.; Abudara, V.; Ezan, P.; Amigou, E.; Sáez, P.J.; Jiang, J.X.; Naus, C.C.; Sáez, J.C.; Giaume, C. Amyloid β-induced death in neurons involves glial and neuronal hemichannels. *J. Neurosci.* 2011, 31, 4962–4977. [CrossRef]
- Quist, A.; Doudevski, I.; Lin, H.; Azimova, R.; Ng, D.; Frangione, B.; Kagan, B.; Ghiso, J.; Lal, R. Amyloid ion channels: A common structural link for protein-misfolding disease. *Proc. Natl. Acad. Sci. USA* 2005, 102, 10427–10432. [CrossRef]
- 99. Yoshiike, Y.; Kayed, R.; Milton, S.C.; Takashima, A.; Glabe, C.G. Pore-forming proteins share structural and functional homology with amyloid oligomers. *Neuromol. Med.* **2007**, *9*, 270–275. [CrossRef]
- 100. White, A.R.; Multhaup, G.; Maher, F.; Bellingham, S.; Camakaris, J.; Zheng, H.; Bush, A.I.; Beyreuther, K.; Masters, C.L.; Cappai, R. The Alzheimer's disease amyloid precursor protein modulates copper-induced toxicity and oxidative stress in primary neuronal cultures. *J. Neurosci.* **1999**, *19*, 9170–9179. [CrossRef]
- Mark, R.J.; Hensley, K.; Butterfield, D.A.; Mattson, M.P. Amyloid beta-peptide impairs ion-motive ATPase activities: Evidence for a role in loss of neuronal Ca2+ homeostasis and cell death. *J. Neurosci.* 1995, 15, 6239–6249. [CrossRef] [PubMed]
- 102. Joslin, G.; Krause, J.E.; Hershey, A.D.; Adams, S.P.; Fallon, R.J.; Perlmutter, D.H. Amyloid-beta peptide, substance P, and bombesin bind to the serpin-enzyme complex receptor. *J. Biol. Chem.* **1991**, 266, 21897–21902. [PubMed]
- 103. Müller, W.E.; Koch, S.; Eckert, A.; Hartmann, H.; Scheuer, K. beta-Amyloid peptide decreases membrane fluidity. *Brain Res.* **1995**, *674*, 133–136. [CrossRef]
- McLaurin, J.; Chakrabartty, A. Membrane disruption by Alzheimer beta-amyloid peptides mediated through specific binding to either phospholipids or gangliosides. Implications for neurotoxicity. *J. Biol. Chem.* 1996, 271, 26482–26489. [CrossRef]
- 105. Kayed, R.; Sokolov, Y.; Edmonds, B.; McIntire, T.M.; Milton, S.C.; Hall, J.E.; Glabe, C.G. Permeabilization of lipid bilayers is a common conformation-dependent activity of soluble amyloid oligomers in protein misfolding diseases. J. Biol. Chem. 2004, 279, 46363–46366. [CrossRef]
- 106. Friedman, R.; Pellarin, R.; Caflisch, A. Amyloid aggregation on lipid bilayers and its impact on membrane permeability. *J. Mol. Biol.* **2009**, *387*, 407–415. [CrossRef]
- 107. Pollard, H.B.; Rojas, E.; Arispe, N. A new hypothesis for the mechanism of amyloid toxicity, based on the calcium channel activity of amyloid beta protein (A beta P) in phospholipid bilayer membranes. *Ann. N. Y. Acad. Sci.* **1993**, 695, 165–168. [CrossRef]
- 108. Khachaturian, Z.S. Calcium hypothesis of Alzheimer's disease and brain aging. *Ann. N. Y. Acad. Sci.* **1994**, 747, 1–11. [CrossRef]

- 109. Bateman, R.J.; Xiong, C.; Benzinger, T.L.; Fagan, A.M.; Goate, A.; Fox, N.C.; Marcus, D.S.; Cairns, N.J.; Xie, X.; Blazey, T.M.; et al. Dominantly Inherited Alzheimer Network, Clinical and biomarker changes in dominantly inherited Alzheimer's disease. *N. Engl. J. Med.* **2012**, *367*, 795–804. [CrossRef]
- 110. Mullane, K.; Williams, M. Alzheimer's therapeutics: Continued clinical failures question the validity of the amyloid hypothesis-but what lies beyond? *Biochem. Pharmacol.* **2013**, *85*, 289–305. [CrossRef]
- 111. Wimo, A.; Winblad, B.; Jönsson, L. An estimate of the total worldwide societal costs of dementia in 2005. *Alzheimers Dement.* 2007, *3*, 81–91. [CrossRef] [PubMed]
- 112. Frautschy, S.A.; Baird, A.; Cole, G.M. Effects of injected Alzheimer beta-amyloid cores in rat brain. *Proc. Natl. Acad. Sci. USA* **1991**, *88*, 8362–8366. [CrossRef] [PubMed]
- 113. Kowall, N.W.; Beal, M.F.; Busciglio, J.; Duffy, L.K.; Yankner, B.A. An in vivo model for the neurodegenerative effects of beta amyloid and protection by substance P. *Proc. Natl. Acad. Sci. USA* 1991, *88*, 7247–7251. [PubMed]
- 114. Geula, C.; Wu, C.K.; Saroff, D.; Lorenzo, A.; Yuan, M.; Yankner, B.A. Aging renders the brain vulnerable to amyloid beta-protein neurotoxicity. *Nat. Med.* **1998**, *4*, 827–831. [CrossRef]
- 115. Shankar, G.M.; Li, S.; Mehta, T.H.; Garcia-Munoz, A.; Shepardson, N.E.; Smith, I.; Brett, F.M.; Farrell, M.A.; Rowan, M.J.; Lemere, C.A.; et al. Amyloid-beta protein dimers isolated directly from Alzheimer's brains impair synaptic plasticity and memory. *Nat. Med.* **2008**, *14*, 837–842. [CrossRef]
- 116. Hardy, J.; Selkoe, D.J. The amyloid hypothesis of Alzheimer's disease: Progress and problems on the road to therapeutics. *Science* 2002, 297, 353–356. [CrossRef]
- 117. Giovannelli, L.; Casamenti, F.; Scali, C.; Bartolini, L.; Pepeu, G. Differential effects of amyloid peptides beta-(1–40) and beta-(25–35) injections into the rat nucleus basalis. *Neuroscience* **1995**, *66*, 781–792. [CrossRef]
- 118. Cleary, J.P.; Walsh, D.M.; Hofmeister, J.J.; Shankar, G.M.; Kuskowski, M.A.; Selkoe, D.J.; Ashe, K.H. Natural oligomers of the amyloid-beta protein specifically disrupt cognitive function. *Nat. Neurosci.* 2005, *8*, 79–84.
- 119. Duyckaerts, C.; Potier, M.C.; Delatour, B. Alzheimer disease models and human neuropathology: Similarities and differences. *Acta Neuropathol.* 2008, *115*, 5–38. [CrossRef]
- 120. Iijima-Ando, K.; Iijima, K. Transgenic Drosophila models of Alzheimer's disease and tauopathies. *Brain Struct. Funct.* 2010, 214, 245–262. [CrossRef]
- 121. Luo, Y. Alzheimer's disease, the nematode Caenorhabditis elegans, and ginkgo biloba leaf extract. *Life Sci.* **2006**, *78*, 2066–2072. [CrossRef] [PubMed]
- 122. Alexander, A.G.; Marfil, V.; Li, C. Use of Caenorhabditis elegans as a model to study Alzheimer's disease and other neurodegenerative diseases. *Front. Genet.* **2014**, *5*, 279. [CrossRef] [PubMed]
- Pimplikar, S.W. Reassessing the amyloid cascade hypothesis of Alzheimer's disease. *Int. J. Biochem. Cell Biol.* 2009, 41, 1261–1268. [CrossRef] [PubMed]
- 124. Robinson, S.R.; Bishop, G.M. The search for an amyloid solution. *Science* **2002**, *298*, 962–964. [CrossRef] [PubMed]
- 125. Hatami, A.; Monjazeb, S.; Glabe, C. The Anti-Amyloid-β Monoclonal Antibody 4G8 Recognizes a Generic Sequence-Independent Epitope Associated with α-Synuclein and Islet Amyloid Polypeptide Amyloid Fibrils. *J. Alzheimers Dis.* **2016**, *50*, 517–525. [CrossRef] [PubMed]
- 126. Kokjohn, T.A.; Roher, A.E. Amyloid precursor protein transgenic mouse models and Alzheimer's disease: Understanding the paradigms, limitations, and contributions. *Alzheimers Dement.* 2009, 5, 340–347. [CrossRef] [PubMed]
- 127. Holcomb, L.; Gordon, M.N.; McGowan, E.; Yu, X.; Benkovic, S.; Jantzen, P.; Wright, K.; Saad, I.; Mueller, R.; Morgan, D.; et al. Accelerated Alzheimer-type phenotype in transgenic mice carrying both mutant amyloid precursor protein and presenilin 1 transgenes. *Nat. Med.* **1998**, *4*, 97–100. [CrossRef]
- 128. Janus, C.; D'Amelio, S.; Amitay, O.; Chishti, M.A.; Strome, R.; Fraser, P.; Carlson, G.A.; Roder, J.C.; St. George-Hyslop, P.; Westaway, D. Spatial learning in transgenic mice expressing human presenilin 1 (PS1) transgenes. *Neurobiol. Aging* 2000, 21, 541–549. [CrossRef]
- 129. Lesné, S.; Kotilinek, L.; Ashe, K.H. Plaque-bearing mice with reduced levels of oligomeric amyloid-beta assemblies have intact memory function. *Neuroscience* **2008**, *151*, 745–749. [CrossRef]
- 130. Holcomb, L.A.; Gordon, M.N.; Jantzen, P.; Hsiao, K.; Duff, K.; Morgan, D. Behavioral changes in transgenic mice expressing both amyloid precursor protein and presenilin-1 mutations: Lack of association with amyloid deposits. *Behav. Genet.* **1999**, *29*, 177–185. [CrossRef]

- 131. Gruart, A.; López-Ramos, J.C.; Muñoz, M.D.; Delgado-García, J.M. Aged wild-type and APP, PS1, and APP + PS1 mice present similar deficits in associative learning and synaptic plasticity independent of amyloid load. *Neurobiol. Dis.* 2008, 30, 439–450. [CrossRef] [PubMed]
- 132. Gonzalez-Lima, F.; Berndt, J.D.; Valla, J.E.; Games, D.; Reiman, E.M. Reduced corpus callosum, fornix and hippocampus in PDAPP transgenic mouse model of Alzheimer's disease. *Neuroreport* **2001**, *12*, 2375–2379. [CrossRef] [PubMed]
- Moechars, D.; Lorent, K.; Van Leuven, F. Premature death in transgenic mice that overexpress a mutant amyloid precursor protein is preceded by severe neurodegeneration and apoptosis. *Neuroscience* 1999, *91*, 819–830. [CrossRef]
- 134. Dodart, J.C.; May, P. Overview on rodent models of Alzheimer's disease. *Curr. Protoc. Neurosci.* 2005, 9, 9.22.1–9.22.16. [CrossRef]
- Findlay, J.A.; Hamilton, D.L.; Sri, S.; Vargas-Caballero, M.; Ashford, M.L.J.; Smith, P.J.S. Manipulation of Amyloid Precursor Protein Processing Impacts Brain Bioenergetics and Glucose Metabolism. *Biophys. J.* 2017, 112, 324a. [CrossRef]
- 136. Carlson, G.A.; Borchelt, D.R.; Dake, A.; Turner, S.; Danielson, V.; Coffin, J.D.; Eckman, C.; Meiners, J.; Nilsen, S.P.; Younkin, S.G.; et al. Genetic modification of the phenotypes produced by amyloid precursor protein overexpression in transgenic mice. *Hum. Mol. Genet.* **1997**, *6*, 1951–1959. [CrossRef]
- 137. Lee, H.W.; Park, J.W.; Sandagsuren, E.U.; Kim, K.B.; Yoo, J.J.; Chung, S.H. Overexpression of APP stimulates basal and constitutive exocytosis in PC12 cells. *Neurosci. Lett.* **2008**, *436*, 245–249. [CrossRef]
- 138. Bishop, G.M.; Robinson, S.R.; Smith, M.A.; Perry, G.; Atwood, C.S. Call for Elan to publish Alzheimer's trial details. *Nature* **2002**, *416*, 677. [CrossRef]
- 139. Check, E. Nerve inflammation halts trial for Alzheimer's drug. Nature 2002, 415, 462. [CrossRef]
- 140. Hopkins, C.R. ACS chemical neuroscience molecule spotlight on ELND006: Another γ-secretase inhibitor fails in the clinic. *ACS Chem. Neurosci.* **2011**, *2*, 279–280. [CrossRef]
- 141. Bard, F.; Cannon, C.; Barbour, R.; Burke, R.L.; Games, D.; Grajeda, H.; Guido, T.; Hu, K.; Huang, J.; Johnson-Wood, K.; et al. Peripherally administered antibodies against amyloid beta-peptide enter the central nervous system and reduce pathology in a mouse model of Alzheimer disease. *Nat. Med.* 2000, *6*, 916–919. [CrossRef]
- 142. Morgan, D.; Diamond, D.M.; Gottschall, P.E.; Ugen, K.E.; Dickey, C.; Hardy, J.; Duff, K.; Jantzen, P.; DiCarlo, G.; Wilcock, D.; et al. A beta peptide vaccination prevents memory loss in an animal model of Alzheimer's disease. *Nature* 2000, 408, 982–985. [CrossRef] [PubMed]
- 143. Elvang, A.B.; Volbracht, C.; Pedersen, L.Ø.; Jensen, K.G.; Karlsson, J.J.; Larsen, S.A.; Mørk, A.; Stensbøl, T.B.; Bastlund, J.F. Differential effects of gamma-secretase and BACE1 inhibition on brain Abeta levels in vitro and in vivo. *J. Neurochem.* 2009, 110, 1377–1387. [CrossRef] [PubMed]
- 144. Gabuzda, D.; Busciglio, J.; Chen, L.B.; Matsudaira, P.; Yankner, B.A. Inhibition of energy metabolism alters the processing of amyloid precursor protein and induces a potentially amyloidogenic derivative. *J. Biol. Chem.* 1994, 269, 13623–13628. [PubMed]
- 145. Hoyer, A.; Bardenheuer, H.J.; Martin, E.; Plaschke, K. Amyloid precursor protein (APP) and its derivatives change after cellular energy depletion. An in vitro-study. J. Neural Transm. (Vienna) 2005, 112, 239–253. [CrossRef] [PubMed]
- 146. Hoyer, S. Age-related changes in cerebral oxidative metabolism. Implications for drug therapy. *Drugs Aging* **1995**, *6*, 210–218. [CrossRef]
- 147. Bhattacharya, S.B.; Datta, A.G. Is brain a gluconeogenic organ? Mol. Cell Biochem. 1993, 125, 51–57. [CrossRef]
- 148. Bélanger, M.; Allaman, I.; Magistretti, P.J. Brain energy metabolism: Focus on astrocyte-neuron metabolic cooperation. *Cell Metab.* **2011**, *14*, 724–738. [CrossRef]
- 149. Wyss, M.T.; Jolivet, R.; Buck, A.; Magistretti, P.J.; Weber, B. In vivo evidence for lactate as a neuronal energy source. *J. Neurosci.* **2011**, *31*, 7477–7485. [CrossRef]
- 150. Siperstein, M.D. Inter-relationships of glucose and lipid metabolism. *Am. J. Med.* **1959**, *26*, 685–702. [CrossRef]
- 151. Yeshao, W.; Gu, J.; Peng, X.; Nairn, A.C.; Nadler, J.L. Elevated glucose activates protein synthesis in cultured cardiac myocytes. *Metabolism* **2005**, *54*, 1453–1460. [CrossRef] [PubMed]
- 152. Souza Ade, A.; da Silva, G.S.; Velez, B.S.; Santoro, A.B.; Montero-Lomelí, M. Glycogen synthesis in brain and astrocytes is inhibited by chronic lithium treatment. *Neurosci. Lett.* **2010**, *482*, 128–132. [CrossRef] [PubMed]

- 153. Plum, F.; Posner, J.B. The diagnosis of stupor and coma. Contemp. Neurol. 1972, 10, 1–286.
- 154. Hamberger, A.C.; Chiang, G.H.; Nylén, E.S.; Scheff, S.W.; Cotman, C.W. Glutamate as a CNS transmitter. I. Evaluation of glucose and glutamine as precursors for the synthesis of preferentially released glutamate. *Brain Res.* 1979, 168, 513–530. [CrossRef]
- 155. Gibson, G.E.; Jope, R.; Blass, J.P. Decreased synthesis of acetylcholine accompanying impaired oxidation of pyruvic acid in rat brain minces. *Biochem. J.* **1975**, *148*, 17–23. [CrossRef]
- 156. Camandola, S.; Mattson, M.P. Brain metabolism in health, aging, and neurodegeneration. *EMBO J.* **2017**, *36*, 1474–1492. [CrossRef]
- 157. Korol, D.L.; Gold, P.E. Glucose, memory, and aging. Am. J. Clin. Nutr. 1998, 67, 764S–771S. [CrossRef]
- 158. Cryer, P.E. Hypoglycemia, functional brain failure, and brain death. *J. Clin. Invest.* **2007**, *117*, 868–870. [CrossRef]
- 159. Daulatzai, M.A. Cerebral hypoperfusion and glucose hypometabolism: Key pathophysiological modulators promote neurodegeneration, cognitive impairment, and Alzheimer's disease. *J. Neurosci. Res.* **2017**, *95*, 943–972. [CrossRef]
- 160. Hoyer, S. The abnormally aged brain. Its blood flow and oxidative metabolism. A review—part II. *Arch. Gerontol. Geriatr.* **1982**, *1*, 195–207. [CrossRef]
- 161. Dastur, D.K. Cerebral blood flow and metabolism in normal human aging, pathological aging, and senile dementia. *J. Cereb. Blood Flow Metab.* **1985**, *5*, 1–9. [CrossRef] [PubMed]
- 162. Hoyer, S. Senile dementia and Alzheimer's disease. Brain blood flow and metabolism. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **1986**, *10*, 447–478. [CrossRef]
- 163. Suh, S.W.; Aoyama, K.; Chen, Y.; Garnier, P.; Matsumori, Y.; Gum, E.; Liu, J.; Swanson, R.A. Hypoglycemic neuronal death and cognitive impairment are prevented by poly(ADP-ribose) polymerase inhibitors administered after hypoglycemia. *J. Neurosci.* 2003, 23, 10681–10690. [CrossRef] [PubMed]
- 164. Auer, R.N. Hypoglycemic brain damage. Metab. Brain Dis. 2004, 19, 169–175. [CrossRef]
- 165. Auer, R.; Kalimo, H.; Olsson, Y.; Wieloch, T. The dentate gyrus in hypoglycemia: Pathology implicating excitotoxin-mediated neuronal necrosis. *Acta Neuropathol.* **1985**, *67*, 279–288. [CrossRef]
- 166. Graham, K.S.; Hodges, J.R. Differentiating the roles of the hippocampal complex and the neocortex in long-term memory storage: Evidence from the study of semantic dementia and Alzheimer's disease. *Neuropsychology* **1997**, *11*, 77–89. [CrossRef]
- 167. Butterfield, D.A.; Drake, J.; Pocernich, C.; Castegna, A. Evidence of oxidative damage in Alzheimer's disease brain: Central role for amyloid beta-peptide. *Trends Mol. Med.* **2001**, *7*, 548–554. [CrossRef]
- 168. Gentleman, S.M.; Nash, M.J.; Sweeting, C.J.; Graham, D.I.; Roberts, G.W. Beta-amyloid precursor protein (beta APP) as a marker for axonal injury after head injury. *Neurosci. Lett.* **1993**, *160*, 139–144. [CrossRef]
- Baiden-Amissah, K.; Joashi, U.; Blumberg, R.; Mehmet, H.; Edwards, A.D.; Cox, P.M. Expression of amyloid precursor protein (beta-APP) in the neonatal brain following hypoxic ischaemic injury. *Neuropathol. Appl. Neurobiol.* **1998**, 24, 346–352. [CrossRef]
- Roberts, G.W.; Gentleman, S.M.; Lynch, A.; Murray, L.; Landon, M.; Graham, D.I. Beta amyloid protein deposition in the brain after severe head injury: Implications for the pathogenesis of Alzheimer's disease. *J. Neurol. Neurosurg. Psychiatry* 1994, 57, 419–425. [CrossRef]
- Li, G.L.; Farooque, M.; Holtz, A.; Olsson, Y. Changes of beta-amyloid precursor protein after compression trauma to the spinal cord: An experimental study in the rat using immunohistochemistry. *J. Neurotrauma* 1995, 12, 269–277. [CrossRef] [PubMed]
- Ahlgren, S.; Li, G.L.; Olsson, Y. Accumulation of beta-amyloid precursor protein and ubiquitin in axons after spinal cord trauma in humans: Immunohistochemical observations on autopsy material. *Acta Neuropathol.* 1996, 92, 49–55. [CrossRef] [PubMed]
- 173. Cornish, R.; Blumbergs, P.C.; Manavis, J.; Scott, G.; Jones, N.R.; Reilly, P.L. Topography and severity of axonal injury in human spinal cord trauma using amyloid precursor protein as a marker of axonal injury. *Spine* (*Phila Pa* 1976) **2000**, *25*, 1227–1233. [CrossRef] [PubMed]
- 174. Vandenabeele, P.; Fiers, W. Is amyloidogenesis during Alzheimer's disease due to an IL-1-/IL-6-mediated "acute phase response" in the brain? *Immunol. Today* **1991**, *12*, 217–219. [CrossRef]
- 175. Thirumangalakudi, L.; Prakasam, A.; Zhang, R.; Bimonte-Nelson, H.; Sambamurti, K.; Kindy, M.S.; Bhat, N.R. High cholesterol-induced neuroinflammation and amyloid precursor protein processing correlate with loss of working memory in mice. *J. Neurochem.* 2008, 106, 475–485. [CrossRef]

- 176. van Groen, T.; Puurunen, K.; Mäki, H.M.; Sivenius, J.; Jolkkonen, J. Transformation of diffuse beta-amyloid precursor protein and beta-amyloid deposits to plaques in the thalamus after transient occlusion of the middle cerebral artery in rats. *Stroke* **2005**, *36*, 1551–1556. [CrossRef]
- 177. Parikh, S.S.; Chung, F. Postoperative delirium in the elderly. Anesth. Analg. 1995, 80, 1223–1232.
- 178. Balin, B.J.; Appelt, D.M. Role of infection in Alzheimer's disease. J. Am. Osteopath. Assoc. 2001, 101, S1-6.
- 179. Miklossy, J.; Kis, A.; Radenovic, A.; Miller, L.; Forro, L.; Martins, R.; Reiss, K.; Darbinian, N.; Darekar, P.; Mihaly, L.; et al. Beta-amyloid deposition and Alzheimer's type changes induced by Borrelia spirochetes. *Neurobiol. Aging* 2006, 27, 228–236. [CrossRef]
- Alkire, M.T.; Haier, R.J.; Shah, N.K.; Anderson, C.T. Positron emission tomography study of regional cerebral metabolism in humans during isoflurane anesthesia. *Anesthesiology* 1997, *86*, 549–557. [CrossRef]
- 181. Hodes, J.E.; Soncrant, T.T.; Larson, D.M.; Carlson, S.G.; Rapoport, S.I. Selective changes in local cerebral glucose utilization induced by phenobarbital in the rat. *Anesthesiology* **1985**, *63*, *633–639*. [CrossRef] [PubMed]
- 182. Xie, Z.; Dong, Y.; Maeda, U.; Alfille, P.; Culley, D.J.; Crosby, G.; Tanzi, R.E. The common inhalation anesthetic isoflurane induces apoptosis and increases amyloid beta protein levels. *Anesthesiology* 2006, 104, 988–994. [CrossRef] [PubMed]
- 183. Tesco, G.; Koh, Y.H.; Kang, E.L.; Cameron, A.N.; Das, S.; Sena-Esteves, M.; Hiltunen, M.; Yang, S.H.; Zhong, Z.; Shen, Y.; et al. Depletion of GGA3 stabilizes BACE and enhances beta-secretase activity. *Neuron* 2007, 54, 721–737. [CrossRef] [PubMed]
- 184. Shi, J.; Xiang, Y.; Simpkins, J.W. Hypoglycemia enhances the expression of mRNA encoding beta-amyloid precursor protein in rat primary cortical astroglial cells. *Brain Res.* **1997**, 772, 247–251. [CrossRef]
- 185. Velliquette, R.A.; O'Connor, T.; Vassar, R. Energy inhibition elevates beta-secretase levels and activity and is potentially amyloidogenic in APP transgenic mice: Possible early events in Alzheimer's disease pathogenesis. J. Neurosci. 2005, 25, 10874–10883. [CrossRef]
- Smith, M.A.; Joseph, J.A.; Perry, G. Arson. Tracking the culprit in Alzheimer's disease. *Ann. N. Y. Acad. Sci.* 2000, 924, 35–38. [CrossRef]
- 187. Lee, H.G.; Casadesus, G.; Zhu, X.; Takeda, A.; Perry, G.; Smith, M.A. Challenging the amyloid cascade hypothesis: Senile plaques and amyloid-beta as protective adaptations to Alzheimer disease. *Ann. N. Y. Acad. Sci.* 2004, 1019, 1–4. [CrossRef]
- Robinson, S.R.; Bishop, G.M. Abeta as a bioflocculant: Implications for the amyloid hypothesis of Alzheimer's disease. *Neurobiol. Aging* 2002, 23, 1051–1072. [CrossRef]
- Amieva, H.; Le Goff, M.; Millet, X.; Orgogozo, J.M.; Pérès, K.; Barberger-Gateau, P.; Jacqmin-Gadda, H.; Dartigues, J.F. Prodromal Alzheimer's disease: Successive emergence of the clinical symptoms. *Ann. Neurol.* 2008, 64, 492–498. [CrossRef]
- 190. de Back, D.Z.; Kostova, E.B.; van Kraaij, M.; van den Berg, T.K.; van Bruggen, R. Of macrophages and red blood cells; a complex love story. *Front. Physiol.* **2014**, *5*, 9. [CrossRef]
- 191. Lang, K.S.; Lang, P.A.; Bauer, C.; Duranton, C.; Wieder, T.; Huber, S.M.; Lang, F. Mechanisms of suicidal erythrocyte death. *Cell Physiol. Biochem.* **2005**, *15*, 195–202. [CrossRef] [PubMed]
- Reiter, C.D.; Wang, X.; Tanus-Santos, J.E.; Hogg, N.; Cannon, R.O., 3rd; Schechter, A.N.; Gladwin, M.T. Cell-free hemoglobin limits nitric oxide bioavailability in sickle-cell disease. *Nat. Med.* 2002, *8*, 1383–1389.
  [CrossRef] [PubMed]
- MacDonald, R. Red cell 2,3-diphosphoglycerate and oxygen affinity. *Anaesthesia* 1977, 32, 544–553. [CrossRef]
  [PubMed]
- 194. Ansari, G.A.; Singh, S.V.; Gan, J.C.; Awasthi, Y.C. Human erythrocyte glutathione S-transferase: A possible marker of chemical exposure. *Toxicol. Lett.* **1987**, *37*, 57–62. [CrossRef]
- 195. Schuster, S.; Kenanov, D. Adenine and adenosine salvage pathways in erythrocytes and the role of S-adenosylhomocysteine hydrolase. A theoretical study using elementary flux modes. *FEBS J.* 2005, 272, 5278–5290. [CrossRef] [PubMed]
- 196. Benesch, R.; Benesch, R.E. The effect of organic phosphates from the human erythrocyte on the allosteric properties of hemoglobin. *Biochem. Biophys. Res. Commun.* **1967**, *26*, 162–167. [CrossRef]
- 197. Nakamura, J.; Koh, N.; Sakakibara, F.; Hamada, Y.; Wakao, T.; Hara, T.; Mori, K.; Nakashima, E.; Naruse, K.; Hotta, N. Polyol pathway, 2,3-diphosphoglycerate in erythrocytes and diabetic neuropathy in rats. *Eur. J. Pharmacol.*1995, 294, 207–214. [CrossRef]

- 198. Kosenko, E.A.; Aliev, G.; Tikhonova, L.A.; Li, Y.; Poghosyan, A.C.; Kaminsky, Y.G. Antioxidant status and energy state of erythrocytes in Alzheimer dementia: Probing for markers. CNS Neurol. Disord. Drug Targets 2012, 11, 926–932. [CrossRef]
- 199. Kaminsky, Y.G.; Reddy, V.P.; Ashraf, G.M.; Ahmad, A.; Benberin, V.V.; Kosenko, E.A.; Aliev, G. Age-related defects in erythrocyte 2,3-diphosphoglycerate metabolism in dementia. *Aging Dis.* 2013, 4, 244–255. [CrossRef]
- 200. Diamond, J.M.; Matsuyama, S.S.; Meier, K.; Jarvik, L.F. Elevation of erythrocyte countertransport rates in Alzheimer's dementia. *N. Engl. J. Med.* **1983**, *309*, 1061–1062.
- Larsen, V.H.; Waldau, T.; Gravesen, H.; Siggaard-Andersen, O. Erythrocyte 2,3-diphosphoglycerate depletion associated with hypophosphatemia detected by routine arterial blood gas analysis. *Scand. J. Clin. Lab. Invest. Suppl.* 1996, 224, 83–87. [CrossRef] [PubMed]
- 202. Papassotiriou, I.; Kister, J.; Griffon, N.; Abraham, D.J.; Kanavakis, E.; Traeger-Synodinos, J.; Stamoulakatou, A.; Marden, M.C.; Poyart, C. Synthesized allosteric effectors of the hemoglobin molecule: A possible mechanism for improved erythrocyte oxygen release capability in hemoglobinopathy H disease. *Exp. Hematol.* **1998**, *26*, 922–926. [PubMed]
- Cholevas, V.; Challa, A.; Lapatsanis, P.D.; Andronikou, S. Changes in red cell phosphate metabolism of preterm and fullterm infants with perinatal problems during their first month of life. *Eur. J. Pediatr.* 2008, 167, 211–218. [CrossRef] [PubMed]
- 204. Smith, C.D.; Carney, J.M.; Starke-Reed, P.E.; Oliver, C.N.; Stadtman, E.R.; Floyd, R.A.; Markesbery, W.R. Excess brain protein oxidation and enzyme dysfunction in normal aging and in Alzheimer disease. *Proc. Natl. Acad. Sci. USA* 1991, *88*, 10540–10543. [CrossRef] [PubMed]
- 205. Ronquist, G.; Waldenström, A. Imbalance of plasma membrane ion leak and pump relationship as a new aetiological basis of certain disease states. *J. Intern. Med.* **2003**, 254, 517–526. [CrossRef] [PubMed]
- 206. Sprandel, U.; Hubbard, A.R.; Chalmers, R.A. Towards enzyme therapy using carrier erythrocytes. *J. Inherit. Metab. Dis.* **1981**, *4*, 99–100. [CrossRef]
- 207. DeLoach, J.R.; Sprandel, U.; Green, R. Carrier erythrocytes: A prospectus for the future. *Bibl. Haematol.* **1985**, *51*, 157–159.



© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).